Seminar

JOURNAL OF HEPATOLOGY

Advanced preclinical models for evaluation of drug-induced liver injury – consensus statement by the European Drug-Induced Liver Injury Network [PRO-EURO-DILI-NET]

Jose C. Fernandez-Checa[1,2,3,4,5,*], Pierre Bagnaninchi[6,7], Hui Ye[8,9], Pau Sancho-Bru[3,4], Juan M. Falcon-Perez[4,10,11], Felix Royo[10], Carmen Garcia-Ruiz[1,2,3,4,5], Ozlen Konu[12,13,14], Joana Miranda[15], Oleg Lunov[16], Alexandr Dejneka[16], Alistair Elfick[17], Alison McDonald[17], Gareth J. Sullivan[18,19,20], Guruprasad P. Aithal[21], M. Isabel Lucena[4,22], Raul J. Andrade[4,23], Bernard Fromenty[24], Michel Kranendonk[25], Francisco Javier Cubero[4,8,9,#], Leonard J. Nelson[6,7,26,*,#]

Summary

Drug-induced liver injury (DILI) is a major cause of acute liver failure (ALF) and one of the leading indications for liver transplantation in Western societies. Given the wide use of both prescribed and over the counter drugs, DILI has become a major health issue for which there is a pressing need to find novel and effective therapies. Although significant progress has been made in understanding the molecular mechanisms underlying DILI, our incomplete knowledge of its pathogenesis and inability to predict DILI is largely due to both discordance between human and animal DILI in preclinical drug development and a lack of models that faithfully recapitulate complex pathophysiological features of human DILI. This is exemplified by the hepatotoxicity of acetaminophen (APAP) overdose, a major cause of ALF because of its extensive worldwide use as an analgesic. Despite intensive efforts utilising current animal and *in vitro* models, the mechanisms involved in the hepatotoxicity of APAP are still not fully understood. In this expert Consensus Statement, which is endorsed by the European Drug-Induced Liver Injury Network, we aim to facilitate and outline clinically impactful discoveries by detailing the requirements for more realistic human-based systems to assess hepatotoxicity and guide future drug safety testing. We present novel insights and discuss major players in APAP pathophysiology, and describe emerging *in vitro* and *in vivo* pre-clinical models, as well as advanced imaging and *in silico* technologies, which may improve prediction of clinical outcomes of DILI.

Keywords: Acetaminophen; iPSCs; liver-on-a-chip; humanized models.

Received 20 April 2021; received in revised form 2 June 2021; accepted 11 June 2021; available online 24 June 2021

Introduction

Drug-induced liver injury (DILI) is an infrequent, multifaceted and potentially life-threatening adverse reaction to medications and other chemical compounds that represents one of the most challenging liver disorders with regards to its prediction, diagnosis and management.[1–4] Idiosyncratic DILI (iDILI) leads to hospitalisation of 23% of affected individuals,[5] accounting for 11% of acute liver failure (ALF) cases in advanced economies, with acetaminophen (paracetamol, APAP) overdose, the prototypical example of intrinsic, predictable DILI, representing 50% of all attributable ALF cases.[6] In addition, 8% of acute DILI cases remain unresolved.[7] As a consequence, DILI jeopardises patient safety and represents a major concern for regulatory authorities; it is both a cause of drug attrition during clinical development and a leading reason for drug withdrawal from the market. The treatment of iDILI is not evidence-based[8] and often relies on *ad hoc* treatment with steroids or ursodeoxycholic acid, particularly in more severe cases.[9] N-acetylcysteine (NAC) has proven effective in very specific instances, such as APAP intoxication,. Clinical aspects of DILI have been covered in a recent review.[1]

DILI pathogenesis is considered a multifactorial process involving several factors other than the generation of toxic intermediate(s) from parental drug metabolism, such as environmental, physiological and genetic factors as well as altered immunological responses. Thus, there is a need for the identification of mechanisms that contribute to DILI in order to develop protective/preventive therapeutic interventions.[6,10] While some of these mechanisms are dose related, others derive from individual susceptibility to the toxic effects of a certain drug, leading to the classification of DILI as either intrinsic, which is considered predictable,

[1]*Cell Death and Proliferation, Institute of Biomedical Research of Barcelona (IIBB), Consejo Superior Investigaciones Científicas (CSIC), Spain;*
[2]*Liver Unit, Hospital Clínic, Barcelona, Spain;*
[3]*Instituto Investigaciones Biomédicas August Pi i Sunyer (IDIBAPS), Universitat de Barcelona, Spain;*
[4]*Centro de Investigación Biomédica en Red de Enfermedades Hepáticas y Digestivas (CIBERehd), Instituto de Salud Carlos III, Madrid, 28029, Spain;*
[5]*USC Research Center for ALPD, Keck School of Medicine, Los Angeles, United States, CA 90033;*
[6]*Center for Regenerative Medicine, Institute for Regenerative and Repair, The University of Edinburgh, Edinburgh, UK, EH16 4UU;*
[7]*School of Engineering, Institute for*

Bioengineering, The University of Edinburgh, Faraday Building, Colin Maclaurin Road, EH9 3 DW, Scotland, UK;
[8]*Department of Immunology, Ophthalmology & ENT, Complutense University School of Medicine, 28040 Madrid, Spain;*
[9]*Health Research Institute Gregorio Marañón (IiSGM), 28007 Madrid, Spain;*
[10]*Exosomes Laboratory, Center for Cooperative Research in Biosciences (CIC bioGUNE), Basque Research and Technology Alliance (BRTA), Derio, Bizkaia, 48160, Spain;*
[11]*IKERBASQUE, Basque Foundation for Science, Bilbao, Bizkaia, 48015, Spain;*
[12]*Department of Molecular Biology and Genetics, Faculty of Science, Bilkent University, Ankara, Turkey;*
[13]*Interdisciplinary Neuroscience Program, Bilkent University, Ankara, Turkey;*
[14]*UNAM-Institute of Materials Science and Nanotechnology, Bilkent University, Ankara, Turkey;*
[15]*Research Institute for iMedicines (iMed.ULisboa), Faculty of Pharmacy, Universidade de Lisboa, 1649-003 Lisbon, Portugal;*
[16]*Department of Optical and Biophysical Systems, Institute of Physics of the Czech Academy of Sciences, Prague, Czech Republic;*
[17]*Institute for Bioengineering, School of Engineering, The University of Edinburgh, Edinburgh EH8 3DW, UK*
[18]*University of Oslo and the Oslo University Hospital, Oslo, Norway;*
[19]*Hybrid Technology Hub-Center of Excellence, Institute of Basic Medical Sciences, University of Oslo, Oslo, Norway;*
[20]*Department of Pediatric Research, Oslo University Hosptial, Oslo, Norway;*
[21]*National Institute for Health Research (NIHR) Nottingham Biomedical Research Centre, Nottingham University Hospital NHS Trust and University of Nottingham, Nottingham, UK;*
[22]*Servicio de Farmacología Clínica, Instituto de Investigación Biomédica de Málaga-IBIMA, Hospital Universitario Virgen de la Victoria, UICEC SCReN, Universidad de Málaga, Málaga, Spain;*
[23]*Unidad de Gestión Clínica de Enfermedades Digestivas, Instituto de Investigación, Biomédica de Málaga-IBIMA, Hospital Universitario Virgen de la Victoria, Universidad de Málaga, Malaga, Spain;*
[24]*INSERM, Univ Rennes, INRAE, Institut NUMECAN (Nutrition Metabolisms and Cancer) UMR_A 1341, UMR_S 1241, F-35000 Rennes, France;*
[25]*Center for Toxicogenomics and Human Health (ToxOmics), Genetics, Oncology and Human*

reproducible and dose-dependent or idiosyncratic, which is unpredictable and not necessarily dose-dependent. DILI covers a broad clinical and histological phenotypic spectrum, including hepatocellular damage, cholestasis, and acute steatosis, which are often detected late in phase III clinical trials or post-marketing.

APAP hepatotoxicity is the archetypal model of DILI and probably the most relevant to human DILI, with billions of analgesic doses consumed annually. APAP hepatotoxicity in humans can be modelled in rodents after administration of an acute or cumulative overdose, often after fasting. However, despite intensive efforts, the mechanisms involved in the hepatotoxicity of APAP are not fully understood, highlighting the imposed limitations of interspecies variability and differences in metabolism between humans and rodents, whilst existing *in vitro* hepatic cell systems based on human cell lines or rodent hepatocytes are suboptimal. These factors have severely hampered pre-clinical efforts to accurately predict DILI and to unravel hidden mechanisms that occur *in vivo*.

In this review, we summarise the pathophysiology of DILI exemplified by the APAP paradigm, describing existing pre-clinical models for DILI. In addition, we assess emerging models, including the development of multi-parametric approaches and humanised models for better DILI prediction. We also highlight how advanced technological integration and innovation could enhance phenotypic profiling, leading to a better understanding of DILI. Future avenues are also explored including novel approaches in delineating mechanistic DILI and the utility of disruptive technologies, such as liver on-chip, to advance DILI prediction.

DILI pathophysiology

The APAP paradigm

APAP-induced liver damage is characterised by haemorrhagic centrilobular necrosis and high plasma transaminase levels in both humans and animals.[2,10] Although APAP is normally metabolised to its glucuronidated and sulphated non-toxic metabolites in the liver, APAP overdose saturates these pathways and excess APAP is metabolised mainly by cytochrome P450 (CYP)2E1 but also by CYPA2, CYP2A6, CYP2D6 and CYP3A4 into the highly reactive metabolite N-acetyl-*p*-benzoquinoneimine (NAPQI). This highly toxic byproduct is rapidly conjugated with glutathione (GSH) resulting in non-toxic mercapturic acid and cysteine conjugates that are excreted in the urine. In APAP overdose or in conditions of GSH limitation (*e.g.* fasting), free unconjugated NAPQI reacts with sulfhydryl groups on cysteine and lysine residues, generating adducts with proteins (APAP-protein adducts) in hepatocytes, and particularly in mitochondria, leading to mitochondrial dysfunction and cell death.[11–13] Despite being the most comprehensively studied and understood hepatotoxic drug, our understanding of the underlying mechanisms involved in APAP hepatotoxicity are still incomplete. Indeed, APAP can also elicit an idiosyncratic response in humans[2] and the use of APAP even at therapeutic doses can have deleterious effects.[14,15] Although models of iDILI are lacking, it is postulated that reactive drug metabolites may elicit an immune response in susceptible individuals.[16] The complex and multifactorial nature of APAP hepatotoxicity extends to DILI itself. Clearly, developing more realistic human models to foster a better understanding of the mechanistic basis of DILI is imperative. In turn, improved models that allow for more accurate prediction of pre-clinical DILI may help uncover effective therapeutic interventions.

Other DILI-causing drugs

Besides APAP, other classes of drug such as non-steroidal anti-inflammatory drugs (NSAIDs) and statins are important causes of DILI, although with a relatively low overall incidence rate. Some of the molecular and cellular mechanisms underlying NSAID-DILI have been identified: (i) mitochondrial injury, (ii) induction of cholestasis, (iii) protein adduct formation by reactive drug metabolites, and (iv) possible direct consequences of cyclo-oxygenase 2 (COX) inhibition.[17] In isolated rat liver mitochondria, diclofenac decreased hepatic ATP content and impaired ATP synthesis causing mitochondrial permeability transition (MPT), leading to generation of reactive oxygen species (ROS), mitochondrial swelling and oxidation of NADPH and protein thiols.[18] Besides diclofenac, indomethacin, celecoxib and ibuprofen NSAIDs can induce endoplasmic reticulum (ER) stress response-related proteins, particularly CHOP, leading to apoptosis.[19]

Statins are generally well tolerated and adverse effects are relatively rare.[20] Mitochondrial dysfunction due to a significant increase in ROS, causing lipid peroxidation and the inhibition of the respiratory chain (complex I and III) which trigger apoptosis, may explain the mechanisms of statin-induced hepatotoxicity.[21]

Emerging mechanisms and signalling cascades governing APAP hepatotoxicity

APAP overdose has major clinical relevance as the primary cause of ALF in advanced economies and a major reason for liver transplantation, and it is regarded as a model hepatotoxin. In the following section we briefly summarise the role of major players that contribute to APAP-induced liver damage. A key question in the pathophysiology of DILI is how a toxin or its (reactive) intermediate metabolites trigger cell damage, which has been intensively investigated in the case of APAP. In this regard, cell-specific (*i.e.* hepatocytes, immune cells) signalling cascades governing APAP hepatotoxicity have attracted much attention since activation/

inhibition of these pathways could be of pivotal importance in patients who do not respond to standard treatment.

MAPK family: JNK

The c-Jun-N-terminal kinase (JNK) is a serine/threonine kinase that belongs to the mitogen-activated protein kinase (MAPK) family, which has been shown to play a causal role in APAP hepatotoxicity by mediating an amplification loop in APAP-induced mitochondrial targeting and oxidative stress.[22] In the liver, 2 JNK genes, *Jnk1* and *Jnk2*, are expressed.[23] Antagonising JNK activation using the classical inhibitor SP600125 has protective effects against APAP-induced liver injury, by significantly reducing necrosis both *in vivo* and *in vitro*.[24,25] Although SP600125 may have effects independent of JNK inhibition, combined *Jnk*1 and *Jnk*2 germ-line deletion or knockdown by antisense oligonucleotides in adult mice markedly protected against APAP hepatotoxicity.[26] In addition, simultaneous deletion of *Jnk1* and *Jnk*2 in adult hepatocytes in *Jnk*1+2$^{f/f}$ mice following injection with an associated adenovirus expressing Cre recombinase driven by the hepatocyte-specific promoter TBG (AAV-TBG-Cre) protected against APAP-mediated liver injury.[27] In contrast with these findings, a recent report in mice with hepatocyte-specific *Jnk1* and *Jnk2* deletion (*Jnk*$^{\Delta hepa}$) questioned the role of JNK in APAP-induced hepatotoxicity, as *Jnk*$^{\Delta hepa}$ mice developed greater liver injury than wild-type animals after APAP overdose, suggesting a beneficial role for combined JNK1 and JNK2 activation in hepatocytes.[28] Whilst the reasons underlying these opposing findings remain to be fully unravelled, in the latter study *Jnk2* was globally deleted in all cell types and *Jnk1* was specifically knocked down in hepatocytes but not in non-parenchymal cells (NPCs), implying opposing roles for Jnk1 in different types of liver cells, as well as in infiltrating inflammatory cells.

The specific contribution of JNK1 and JNK2 in DILI remains controversial. No differences in APAP hepatotoxicity in *Jnk1* knockout mice were observed,[26] despite a clear pro-apoptotic and pro-fibrogenic function of *Jnk1* in tumour necrosis factor (TNF)-induced cell death,[29] and in liver fibrosis.[30] Regarding *Jnk2*, increased susceptibility towards APAP, TNF and lipopolysaccharide-induced liver injury was reported upon *Jnk2* deficiency,[31] whilst *Jnk2* disruption protected against APAP-induced liver injury.[26] Recent findings have shown that both hepatocyte *Jnk*2 knockout and knockdown ameliorated ibuprofen-mediated DILI.[32] Recent investigations highlighted the critical role of immune cells in APAP-induced ALF, including activation of resident hepatic macrophages (Kupffer cells [KCs]) following hepatocyte necrosis as well as massive CCR2-dependent recruitment of monocytes.[33]

JNK activation factors

Preclinical findings in constitutive and conditional knockout mice have shown that JNK can be activated by many factors, ranging from various pathogens and cytokines, including transforming growth factor-β (TGF-β), interleukin-1β and TNF to oxidative stress and DNA damage in both hepatocytes and infiltrating cells.[22,34] Phosphorylation of JNK is mediated by MAP2Ks,[35] which, in turn, are phosphorylated and activated by MAP3K. The best characterised MAP3Ks are the apoptosis signal-regulating kinase-1 (ASK1) and mixed-linage kinase 3 (MLK3). ASK1 participates in APAP-induced JNK activation,[36] which is achieved by dissociation from thioredoxin-1 (Trx-1) (Fig. 1). MLK3, a member of the Ser/Thr protein kinase family, mediates the initial phase of JNK activation.[37] Glycogen synthase kinase 3β (GSK-3β) is also involved in the early-phase of JNK activation. Inhibition of GSK-3β in mice prevented JNK activation and ameliorated APAP-derived toxicity.[25] The MAP2Ks (MKK4 and MKK7) are capable of phosphorylating JNKs at Thr/Tyr residues.[38] Furthermore, MKK4 activates both JNK and p38 kinases, while MKK7 only activates JNK. An additional factor that contributes to sustained JNK phosphorylation in APAP hepatotoxicity is impaired MAPK phosphatase (Mkp) activity. Mkp deficiency in mice has been shown to exacerbate APAP-induced liver injury along with sustained JNK activation, while Mkp activation prevents JNK activation and subsequent APAP hepatotoxicity.[39,40]

JNK amplification loop

In recent studies utilising novel mouse liver models, a feedforward self-sustaining signalling pathway referred to as the JNK amplification loop[41,42] was reported to maintain sustained JNK activation, leading to liver damage and dysfunction in response to APAP. Activated JNK (p-JNK) translocates to mitochondria and binds to the Sab (SH3BP5) protein on the outer mitochondrial membrane,[43,44] impairing mitochondrial respiration and enhancing the release of ROS.[45] ROS release, in turn, activates ASK1 and MKK4, which sustain JNK activity and amplify the toxic effect. The binding of JNK to the outer mitochondrial membrane via Sab further induces MPT, thus changing the permeabilisation of the mitochondrial outer membrane and allowing the exit of molecules less than 1,500 Da, including cytochrome c, apoptosis inducing factor (AIF) and endonuclease G.[46] Although release of cytochrome c and AIF is a hallmark of apoptosis, as they activate caspase 3/7 and lead to nuclear DNA cleavage, respectively,[47,48](Fig. 1), the major form of cell death in APAP toxicity is necrosis.[49] This may be because the marked injury in the mitochondria and the pronounced reduction in ATP cannot sustain activation of the apoptotic cascade. It should be noted that, currently, other less familiar modes of

Toxicology, NOVA Medical School, Faculty of Medical Sciences, Universidade NOVA de Lisboa, Lisbon, Portugal;
[26]*Institute of Biological Chemistry, Biophysics and Bioengineering (IB3), School of Engineering and Physical Sciences (EPS), Heriot-Watt University, Edinburgh EH12 2AS, Scotland, UK*

[#]Contributed equally

• Corresponding authors. Addresses: IIBB-CSIC-IDIBAPS, Centre Esther Koplowitz (CEK), C/ Roselló, 153, 08036 Barcelona, Spain; Tel.: +34 93 227 5709 (J.-C. Fernandez-Checa), or School of Engineering, Institute for Bioengineering, The University of Edinburgh, Faraday Building, Colin Maclaurin Road, EH9 3 DW, Scotland, UK. Tel.: +44 (0) 131 651 7181 (L.J. Nelson).

E-mail addresses: checa229@yahoo.com (J.C. Fernandez-Checa), l.nelson@ed.ac.uk (L.J. Nelson).

https://doi.org/10.1016/j.jhep.2021.06.021

Key point

Gaps in our understanding of DILI and the complexity of underlying mechanisms coupled with interspecies differences have hampered efforts to develop reproducible animal models.

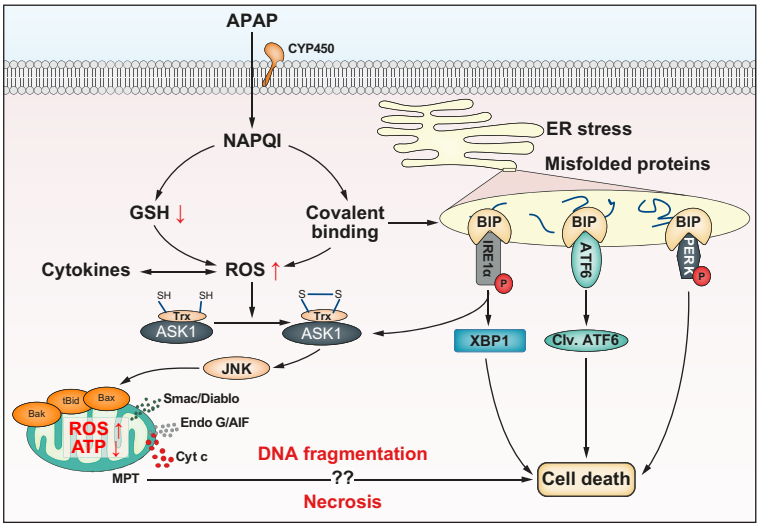

Fig. 1. Pathophysiology of APAP-induced liver injury. APAP toxicity is caused mainly by the excess formation of NAPQI. Enhanced NAPQI depletes hepatic GSH, covalently binds to proteins and forms protein adducts. ROS accumulation oxidises and removes Trx-1 from Trx-ASK1 complexes, leading to activation of ASK1 and subsequently the apoptosis signalling cascade. Activated JNK translocates into the mitochondria and alters the mitochondrial membrane potential, which triggers DNA fragmentation and cell death. Opening of the MPT contributes to the predominant APAP-induced necrotic cell death, compared to the minor role of the release of Cyt c, and apoptosis factors AIF, Smac/endo G. The increase of misfolded or unfolded proteins in the ER lumen triggers the ER stress-mediated UPR, which has 3 different effectors: PERK, ATF6 and IRE1α. If the UPR cannot efficiently restore ER homeostasis, it will ultimately induce the elevated expression of CHOP and lead to cell death. AIF, apoptosis inducing factor; APAP, acetaminophen; ASK1, apoptosis signal-regulating kinase-1; ATF6, activating transcription factor 6; Bak, BCL2 antagonist/killer; Bax, BCL2 associated X, apoptosis regulator; CHOP, CCAAT-enhancer-binding protein homologous protein; Cyt c, cytochrome c; ER, endoplasmic reticulum; GSH, glutathione; IRE1α, inositol-requiring enzyme 1α; JNK, c-Jun-N-terminal kinase; MPT, mitochondrial permeability transition; NAPQI, N-acetyl-p-benzoquinone imine; PERK, protein kinase RNA-like ER kinase; ROS, reactive oxygen species; tBid, truncated Bid, Trx-1, thioredoxin-1; UPR, unfolded protein response; XBP1, X-box binding protein 1.

Key point

Currently, none of the existing models are approved by regulatory agencies in Europe and the US, given the limited predictive value of current preclinical systems.

cell death including pyroptosis, necroptosis and ferroptosis with alternate mechanistic pathways are under active investigation with respect to their contribution to APAP hepatotoxicity.[50] Moreover, although autophagy is another form of cell death, it is considered a protective mechanism against APAP hepatotoxicity (see below).

ER stress and mitochondrial cholesterol accumulation

The ER stress-mediated unfolded protein response (UPR) is an adaptive stress response resulting in accumulation of unfolded or misfolded proteins in the ER lumen.[51] ER stress can be detected late after APAP challenge (500 mg/kg) in murine models, and becomes highly significant 12 hours following APAP administration.[52] The ER stress response has 3 signalling arms: (i) protein kinase RNA-like ER kinase (PERK), (ii) activating transcription factor 6 (ATF6), and (iii) inositol-requiring enzyme 1α (IRE1α). These pathways are maintained in an inactive state through binding to BiP (HSPA5) in non-stressed cells. Upon APAP-mediated ER stress, IRE1α, PERK and ATF6 become activated, triggering an inflammatory response and cell death mediated via ASK1 and JNK[52](Fig. 1). However, in mice with genetically deleted *XBP1 (*X-box binding protein 1*)*, constitutive IRE1α hyperactivation in hepatocytes resulted in reduced JNK activation and protection from APAP through suppression of CYP activity.[53] Recently, the steroidogenic acute regulatory protein 1 (STARD1), a mitochondrial cholesterol transport protein, has been identified as a key player in ER-stress mediated APAP hepatotoxicity.[27]

In this respect, STARD1 promotes cholesterol trafficking and accumulation in mitochondria, which in turn leads to mitochondrial GSH depletion and contributes to mitochondrial dysfunction, exacerbated ROS generation and necrotic cell death (Fig. 2). An intriguing finding is the protection of mice with liver-specific STARD1 deletion despite preserved mitochondrial Sab/p-JNK activation, suggesting that the deleterious effect of p-JNK in mitochondrial dysfunction and hepatocyte cell death is dependent on STARD1.[27] In addition, hepatocyte-specific deletion of Sab or p-JNK1+2 was also protective against APAP hepatotoxicity, preventing APAP-induced ER stress and subsequent STARD1 upregulation. Further mechanistic studies utilising human models and clinical samples (see below) will likely lead to confirmation of the molecular basis for the complementary role of STARD1/mitochondrial cholesterol and the Sab/p-JNK axis in APAP hepatotoxicity – and the upstream events involved in p-JNK1/2 induced ER stress.

Furthermore, as mitochondrial dysfunction contributes to APAP hepatotoxicity, removal of damaged mitochondria through mitophagy has emerged as a critical mechanism in APAP-induced ALF.[13] Besides transcriptional regulation, autophagy can also be modulated by lysosomal lipid composition. Indeed, accumulation of lipids (*e.g.* cholesterol) in lysosomes has been shown to impair the fusion of autophagosomes (containing disrupted mitochondria) with lysosomes, contributing to perpetuation of damaged mitochondria, which sensitises to APAP hepatotoxicity.[54] Thus, it is not only the intermediates of APAP metabolism (*e.g.* NAPQI) acting directly on mitochondria that determine APAP hepatotoxicity, but also secondary factors that delay mitochondrial turnover via mitophagy. The latter can be of particular clinical relevance in non-alcoholic steatohepatitis (NASH), which can potentiate DILI[55] (see Section Implications of DILI in clinical contexts). In fact, it has previously been reported that patients with NASH exhibit increased expression of STARD1,[56] suggesting that a subset of patients with advanced non-alcoholic fatty liver disease (NAFLD), with enhanced free cholesterol content and STARD1 expression, may develop liver injury on APAP ingestion.

In summary, it is clear that the JNK signalling pathway is a critical component in DILI, particularly in APAP pathogenesis. Since JNK differentially regulates

important biologic targets, this cascade can be either beneficial or detrimental in different cells and tissues, and compensatory mechanisms need to be modulated or even discarded. In addition, upstream or downstream pathways regulating the JNK-specific role in cell death during APAP hepatotoxicity are pivotal to developing new therapeutic interventions in patients with DILI.

Other signalling pathways and mechanisms

Apart from MAPK, other pathways have been reported to modulate APAP hepatotoxicity. Inhibition of protein Kinase C (PKC) prevents APAP hepatotoxicity by blocking ROS-mediated hepatic necrosis.[57] The receptor interacting protein kinases (RIPKs) that modulate necroptosis have a controversial role in DILI which is the subject of intensive research. RIPK3-deficient mice were protected from early phase APAP toxicity, which also resulted in the prevention of ROS-JNK associated signalling.[58] In contrast, other studies found no evidence that RIPK3 or the pseudokinase MLKL participate in APAP-mediated injury.[59] Indeed, RIPK1 inhibition reversed APAP-induced JNK activation and liver damage, a possible mechanism associated with ASK1 and ER stress.[36,60] As these studies used global RIPK1/3 deletion, conditional deletion in adult mice would be needed in order to unequivocally demonstrate the role of RIPK1/3 in APAP hepatotoxicity.

Liver sinusoidal endothelial cells (LSECs) form the wall of hepatic sinusoids, regulate hepatic vascular tone and contribute to the maintenance of a low portal pressure. LSECs help maintain hepatic stellate cell quiescence, and thus essentially inhibit intrahepatic vasoconstriction and fibrosis development. In line with their key role in hepatic homeostasis, LSECs play a key role in the initiation and progression of chronic liver disease and DILI.[61] Pioneering studies identified LSEC as a target for APAP toxicity,[62] with further investigations revealing the ability of APAP to cause LSEC apoptosis via Trail,[63] leading to hepatic congestion and haemorrhagic lesions. Quite intriguingly, recent findings revealed that the accumulation of free cholesterol in the endolysosomes of LSECs exacerbates APAP hepatotoxicity via Toll-like receptor 9/inflammasome pathway.[64] These findings highlight that hepatic steatosis, and in particular increased liver cholesterol, emerge as a risk factor for APAP hepatotoxicity (see section NAFLD and ageing as a susceptibility state for DILI).

Adaptive and cellular protective mechanisms: Autophagy | Keap1/Nrf2

Macroautophagy (autophagy) is a non-selective bulk degradation process aimed at recycling cellular components and damaged organelles in response to a variety of stimuli, such as nutrient deprivation and toxic stress, including APAP hepatotoxicity. Using primary mouse hepatocytes and GFP/light chain 3 transgenic mice, Ni and colleagues reported that APAP-induced autophagy correlated with recycling of damaged mitochondria.[65] APAP suppressed mTOR complex 1 and APAP-induced autophagy was blocked by NAC, suggesting APAP mitochondrial protein binding and the subsequent production of ROS elicited APAP-mediated autophagy. Importantly, pharmacological inhibition of autophagy further exacerbated APAP-induced hepatocytotoxicity, while induction of autophagy by rapamycin inhibited APAP-induced liver injury. The hepatoprotective role of autophagy in APAP hepatotoxicity was due to the elimination of damaged mitochondria by a more selective process called mitophagy,[65] which impacted removal of APAP-protein adducts.[66] Interestingly, the APAP-induced mitophagy appears to be predominant in zone 3 of the liver compared to zone 1 (coinciding with the site of APAP metabolism), suggesting mitophagy as an adaptive mechanism to promote cell survival and restrict the expansion of necrotic areas.[67] In line with these findings, adiponectin has emerged as an adaptive mechanism to ameliorate APAP

Key point

APAP hepatotoxicity is multifaceted and molecular pathways incompletely understood, although disruption of mitochondrial function is a well-recognised player in APAP-mediated liver injury. Novel and emerging mechanisms have been identified in this critical step, although they remain to be validated in human DILI.

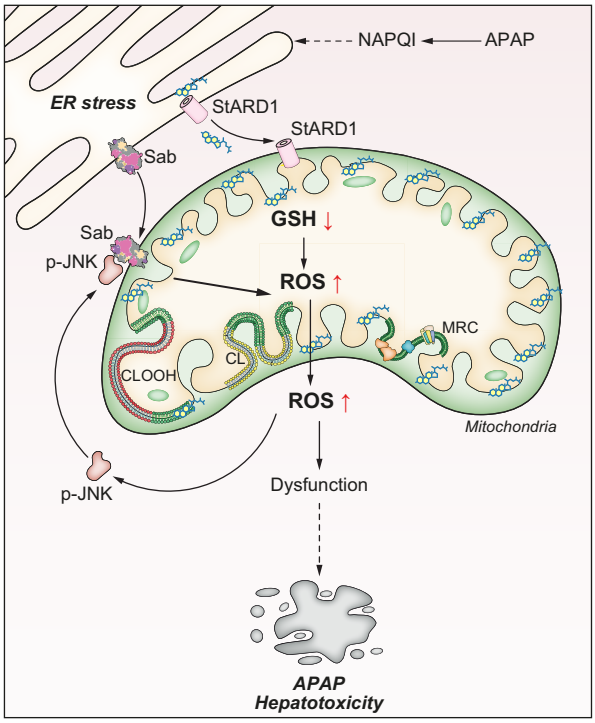

Fig. 2. Schematic role of Sab and STARD1 in APAP hepatotoxicity. Both Sab (SH3BP5) and STARD1 are induced upon APAP metabolism and act in mitochondria. Whereas Sab functions as a docking site for activation of JNK to mediate mitochondrial dysfunction and ROS generation, STARD1 activation by APAP-mediated ER stress causes the accumulation of cholesterol in mitochondrial membranes (orange structures), which contributes to the limitation of mitochondrial antioxidant defence and potentiation of ROS generation. Intriguingly, depletion of Sab and STARD1 independently protect against APAP hepatotoxicity, suggesting that both proteins exert complementary roles in APAP-induced liver injury. APAP, acetaminophen; ER, endoplasmic reticulum; GSH, glutathione; JNK, c-Jun-N-terminal kinase; MRC, mitochondrial respiratory chain; ROS, reactive oxygen species; STARD1, steroidogenic acute regulatory protein 1.

hepatotoxicity by promoting mitophagy through stimulation of autophagosome formation by AMPK-dependent activation of Unc-51-like kinase 1.[68]

Besides mitophagy, ROS generation can be offset by an antioxidant stress response controlled by nuclear erythroid-2-related factor 2 (Nrf2).[69] The Kelch-like ECH-associated protein 1(Keap1)/Nrf2 system is recognised as an important cytoprotective pathway combating cellular oxidative injury.[70] Liver conditional Keap1 knockout or activators of Nrf2 provide protection against APAP-induced liver injury, while deletion of Nrf2 results in hypersensitivity to APAP hepatotoxicity.[71] Indeed, farrerol, a 2,3-dihydro-flavonoid isolated from rhododendron, has been shown to confer rapid (within 1 hour) protection against APAP hepatotoxicity by activation of Nrf2 and autophagy.[72] Thus, whether targeting autophagy and Nrf2 in combination with NAC may be a relevant approach to reduce APAP-mediated ALF remains to be investigated.

Emerging mechanisms and approaches in DILI

Extracellular vesicles

Like other cell types, hepatocytes secrete extracellular vesicles (EVs), both under physiological and pathological conditions, including in chronic injury, such as liver fibrosis, and in DILI.[73] EVs are membrane-bound vesicles released into the extracellular milieu, protected by a lipid bilayer, which also include protein receptors and signal triggering molecules. The EVs carry diverse cargo that include proteins, active enzymes, coding and non-coding RNA, DNA, and metabolites.[74–76] Three different types of EV can be released from cells (exosomes, microvesicles [MVs], and apoptotic bodies) and are closely related to the mechanism of biogenesis. Exosomes are the smallest EVs (30-150 nm),[77] which are formed in multivesicular bodies (MVBs) of the endocytic and secretory pathway.[78,79] Microvesicles (50-3,000 nm) are formed directly by outward budding of the plasma membrane.[80] Whilst apoptotic bodies are EVs (>500 nm) that originate from cells undergoing apoptosis.[81] In the liver, the first descriptions of hepatocyte-derived vesicles were obtained from primary culture of rat hepatocytes,[82] and isolated hepatic stem cell cultures.[83] Many researchers have since contributed to the characterisation of liver-derived EVs in different contexts of liver disease.[84–86]

Of relevance to DILI, hepatocyte-derived EVs can be influenced by drug metabolism, affecting the protein cargo composition, morphology and number.[87,88] Following APAP or diclofenac exposure released EVs have been shown to contain liver-specific mRNA (*e.g.* ALB gene),[89] liver-specific miRNA (such as miR-122),[84] and liver specific proteins such CPS1, MAT1 and COMT.[87,89–93] Several studies have further unravelled the diverse cargo of EVs in DILI (Fig. 3), including the presence of CYPs, such as CYP2A1, 2B3, UDP-glucuronosultransferases (UGT), and 2B2 isoforms, and sulfotransferase 1A1 in rats.[82] Notably, CYPs 1A2, 2B6, 2E1, 3A4 and UGT 1A1, and other isoforms have been detected in circulating EVs isolated from the plasma of patients with DILI.[93,94]

Apart from CYPs, EVs may also harbour other active enzymes, such as Arginase 1 (Arg1)[95] or the carboxylesterase 3 (CES3).[90] Due to this capability of transporting active liver enzymes, it is likely that circulating EVs are also involved in the pathogenesis of DILI, since they can reach different tissues, such as the lung or brain,[96] and modify acceptor cell responses, such as the contractile capability of blood endothelium[95] – potentially playing a role in the pathogenesis of lung hypertension related to liver damage. Moreover, the presence of active CYP2E1 in EVs suggest that they may exacerbate APAP-induced toxicity in hepatocytes and monocytes.[97] The presence of drug-induced protein modifications within the EV cargo can potentially have negative effects through covalent binding to certain drugs, such as amoxicillin or flucloxacillin. These protein-adducts can also induce the activation of dendritic cells when exposed to EVs released by hepatocytes, indicating that EVs could play a role in drug-induced autoimmune hepatitis.[98]

AOP framework

As DILI is a highly heterogeneous process, the mechanistic characterisation underlying this wide spectrum of hepatotoxic manifestations is a requisite to improve prediction. Effective experimental approaches for DILI evaluation require novel preclinical test systems that faithfully mimic these heterogeneous pathways. Context-specific *in vitro* models for assessing hepatotoxicity have been characterised recently[99] – and several new technological approaches are being developed in the search for more predictive systems (see Sections Towards capturing hepatocellular complexity and Advanced technologies).

Although a substantial amount of mechanistic data on DILI is currently known (*e.g.* obtained through genome-wide association studies [GWAS][100] or transcriptomics approaches[101]), there remain significant gaps in our understanding of hepatotoxic outcomes following chemical exposure. The lack of detailed understanding of the mechanistic pathways underlying the multifactorial nature of DILI has impeded the development of improved treatment and cell systems to test novel therapies. Support for the AOP concept[102] has gained momentum and, together with parallel improvements in test systems, could help to bridge this gap in knowledge.

AOP is a mechanistic representation of critical toxicological effects that propagate over different

Key point

Novel approaches and emerging mechanisms in APAP hepatotoxicity and DILI may be of significance for the discovery of potential treatments.

layers of biological organisation, from the initial interaction of a chemical compound with a molecular target, to an adverse outcome at the individual or population level (Fig. 4). An AOP describes a sequence of events starting with a molecular initiation event (the molecular target), with progression through a series of key events, linked by key event relations, which may occur at the sub-cellular/cellular-/or tissue-level, up to the whole organism. It describes only toxicodynamic interactions and pathways, and as such, is compound agnostic, *i.e.* independent of any specific chemical or its dose level. AOP is a valuable approach to incorporate mechanistic knowledge as demonstrated for APAP, chlorpromazine and other DILI-causing drugs[102,103] – it is a multi-scale data integration tool in which newly obtained mechanistic data can be used to feed the linear AOP structure (see below, and Fig. 4). Moreover, AOP can enhance our mechanistic knowledge,[102] as it can identify deficits in existing tests and models intended to predict DILI. The AOP framework could be particularly helpful in delineating underlying causes and mechanisms in iDILI. As such, piecing together currently fragmented data sets from studies on iDILI in an AOP scheme will fill knowledge gaps, enabling the design of effective experimental approaches to unveil pathway(s) and help reduce the unpredictability of iDILI. Additionally, AOP is geared towards the indication of potential DILI biomarkers as early indicators of DILI, as practical read-outs in experimental approaches, as well in pharmacovigilance. Initial AOPs for several forms of DILI have recently been established[104] and contain substantial mechanistic information on liver fibrosis, steatosis, and cholestasis.

The linear schematisation of current mechanistic information in AOP generates relevant data on metabolic alterations, such as bile acid homeostasis,[105] mitochondrial dysfunction,[106] or the role of innate immune responses.[107] In addition, this workflow points to AOP as a practical tool for the design, development, and validation of improved experimental models for DILI prediction. Indeed, a recent AOP approach that integrates mechanistic knowledge of multiple data sources enabled selection of a number of *in vitro* assays as effective predictors of DILI risk.[108]

Toxicogenomic approaches can further reveal fundamental molecular mechanisms and improve prediction of toxicity through integration of cross-omics technology including epigenomics, transcriptomics, proteomics and metabolomics, along with physiologically based pharmacokinetic experiments. This has been used in 3D human liver and heart microtissues with advanced *in silico* bioinformatics to predict DILI.[109]

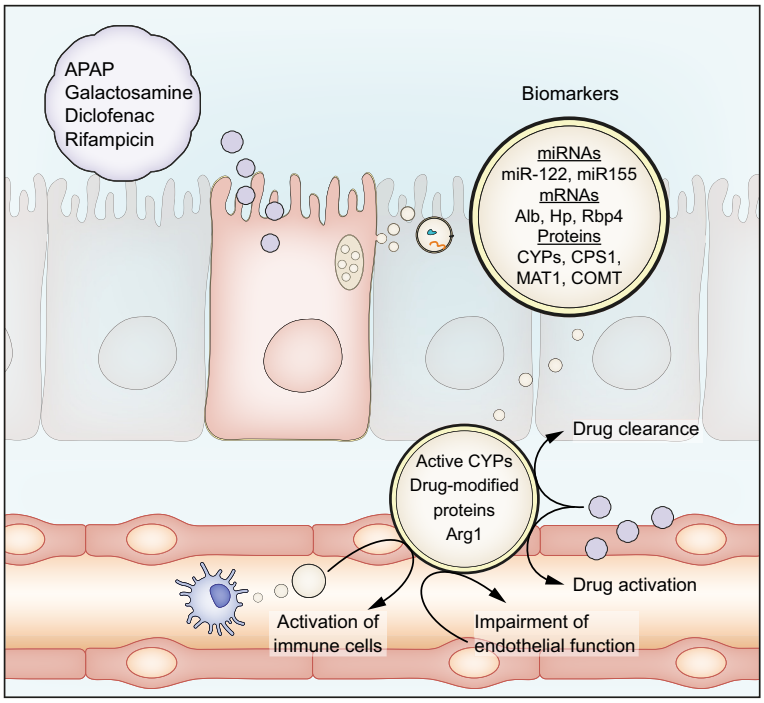

Fig. 3. Role of extracellular vesicles in DILI. Hepatotoxic drugs induce the release of extracellular vesicles from hepatocytes carrying a differential cargo which can be isolated from plasma or urine, providing a unique source of low-invasive biomarkers. Extracellular vesicles carry cargo including active enzymes that can modify the microenvironment, participating in drug clearance, but also forming active drug-protein adducts that increase toxic effects, or deplete metabolites from blood. Drug-modified proteins can trigger an immune response when extracellular vesicles are presented to dendritic cells. Studies thus far suggest that extracellular vesicles play an important role in the pathogenesis of DILI, but also offer an opportunity for drug diagnosis (as biomarkers) and therapy. Alb, albumin; APAP, acetaminophen; Arg1, arginase 1; COMT, catechol-O-methyltransferase; CPS1, carbamoyl-phosphate synthase 1; CYP, cytochrome P450; DILI, drug-induced liver injury; Hp, haptoglobin; MAT1, methionine adenosyltransferase 1; miR, microRNA; Rbp4, retinol binding protein 4.

Toxicogenomics approaches

Next generation genomic technologies are now being used as powerful tools in the armament to investigate DILI. Toxicogenomics allows for detailed analysis of altered gene and protein expression profiles and across biological scales (which are also relevant to AOP: molecular-single cell – population levels) in response to xenobiotic exposure.[110] Its potential for application is enhanced by the availability of accessible databases that can facilitate and harness generated Omics and imaging data (*e.g.* Open TG-GATEs[111]). GWAS have resulted in highly revealing findings, such as HLA polymorphisms related to DILI;[112] whilst transcriptomics-based Big Data-driven analysis has identified adverse outcomes at cellular and organism levels.[101] The relative risk for complex diseases such as coronary artery disease can be predicted by calculating the polygenic risk score (PRS). Recently, Koido *et al.* used a GWAS-based PRS prediction strategy to demonstrate that genetic variation in susceptibility to DILI resides in hepatocytes. The authors used PRS, which sums up the effects of hundreds or thousands of variants, combined with genomic, cellular and organoid 'polygenicity-in-a-dish' approaches to delineate a

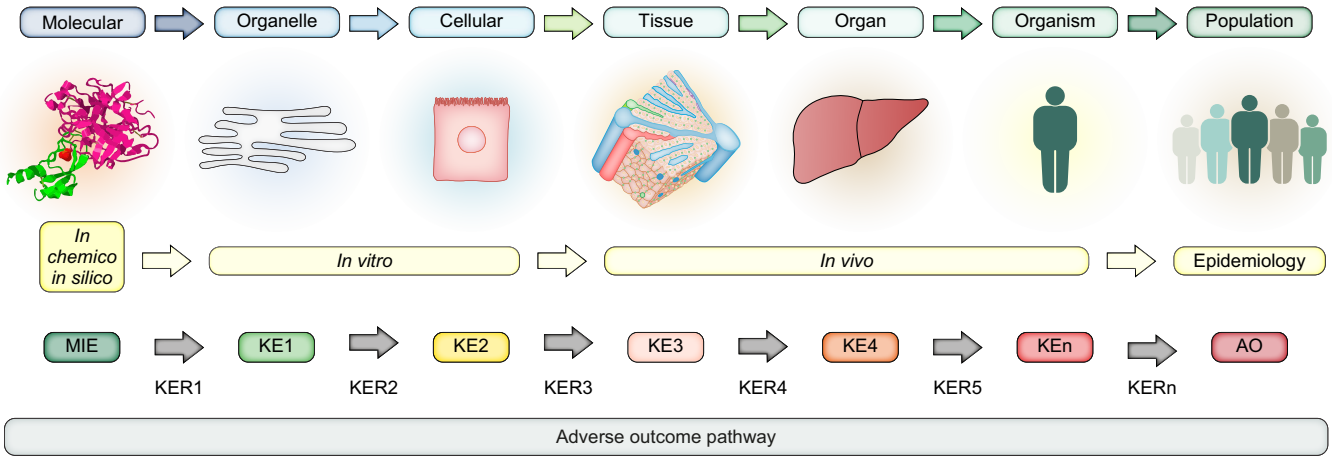

Fig. 4. Adverse outcome pathway in DILI. General representation of the structure of the adverse outcome pathway, applicable in DILI research. The structure is fed with information obtained from different levels of biological experimentation. It can be considered as a multi-scale data integration tool, helpful in identifying knowledge gaps and prone toward indicating potential biomarkers. (adapted from: Vinken *et al.*, 2017[104]). AO, adverse outcome; DILI, drug-induced liver injury; KE, key event; KER, key event relation; MIE, molecular initiation event.

spectrum of DILI-causing agents including fasiglifam (TAK-875).[113] This demonstrates the utility of toxicogenomics approaches for DILI and other rare diseases.[113]

Recent application of single-cell transcriptomics in DILI has, for the first time, enabled the identification of unique subsets of MYC-dependent, activated liver-resident cellular types or 'states' (Kupffer, stellate, and liver sinusoidal endothelial cells) in APAP-induced ALF in mice, which correlated with human ALF.[114] This approach may enable pathway-specific therapeutic interventions for ALF. Metabolomics can also be highly informative for DILI prediction, whereby detection of endogenous metabolites/reactive metabolites can be complemented with the development of AOP for the design of more effective approaches for DILI prediction.[115]

Thus, whilst current methods to study DILI pathology have involved mostly well-defined endpoint assays such as immunostaining, multiparametric image analysis (cell viability), ultrastructural imaging, quantitative reverse-transcription PCR, western blot or flow cytometry techniques, powerful complementary assays are increasingly becoming available that, when coupled with emerging human-based multi-cellular models, can shed light on human-specific toxicity mechanisms.

Towards capturing hepatocellular complexity

Choice of cells - not all cells are equal

Conventional hepatic culture models for drug discovery assays mostly use rodent primary hepatocytes or human immortalised cell lines. However, these rapidly lose polarity and differentiated phenotype[116,117] and are not representative of normal liver tissue. Such models often lack the functional repertoire of primary human hepatocytes (PHHs), including the ability to metabolise drugs (CYP activity). However, PHHs have a short culture life-span, and exhibit phenotypic variability and instability in culture with intermittent supply and high unit costs.[118] The multi-billion dollar drug development process is often hampered by the fact that candidate drugs, which show promise in preclinical animal models, subsequently do not show efficacy in humans, due to interspecies metabolic differences.[116] DILI is a leading cause of drug withdrawal from the market, highlighting the fact that current preclinical models of toxicity are not universally predictive of drug effects in humans.[119] DILI accounts for a 30% of attrition of all pharmaceutical compounds,[120] therefore a robust and scalable human hepatic *in vitro* cell culture platform would enable physiologically relevant preclinical data for drug screening for DILI.

HepaRG cell line

The human liver-derived HepaRG cell line is now considered the closest surrogate to PHHs for DILI applications. The HepaRG cell line is a unique and sustainable intrinsic human co-culture model system for reproducible measurements of drug uptake, metabolism and toxicity. The hepatic HepaRG bipotential progenitor cell line can differentiate into mature hepatocyte-like cells (HLCs) and biliary epithelial cells (BECs). Various liver-specific phenotypic functions[15,121] are stably expressed in HepaRGs, including the major CYPs – at levels comparable to those found in PHHs, with high functional stability for several weeks. This cell line has been used as a scaffold-free spheroid to screen toxicity profiles and thresholds of a number of compounds.[122] Altogether, HepaRGs provide a high-fidelity, sustainable organotypic model system for exploring mechanisms of APAP toxicity and

other forms of DILI such as chlorpromazine.[122–124] Coupling organotypic human HepaRG cells with various combinations of NPC types: hepatic stellate cells (HSCs), KCs, and LSECs provides a rational approach to providing context-specific models to investigate DILI, *viz*: i) Immunomodulatory (HepaRG:KCs), ii) vascular (HepaRG:LSECs); and iii) fibrogenic (HepaRG:HSCs) models. Stepwise integration of these cell types within a microphysiological system as well as novel 2D-3D platforms (see below, and Section Advanced technologies), could be an important step in enhancing our understanding of DILI pathophysiology to solve the prediction dilemma in drug development. In principle, this approach may begin to discern what factors are lacking from current models to improve model relevance for DILI and thus unravel novel toxicity mechanisms leading to DILI.

Alternative *in vitro* hepatic models: iPSC-derived hepatic tissue

Alternative strategies to provide liver cell surrogates are found in 2 forms of human pluripotent stem cells (hPSCs): human embryonic stem cells (hESCs) and induced pluripotent stem cells (iPSCs). Both have the potential to serve as a source of HLCs and other key cellular players for drug discovery and DILI research. However, ethical considerations, with moratoria or outright bans on hESC use in many countries, have prevented their widespread adoption. Therefore, hiPSCs, which are derived through the reprogramming of somatic cells, such as fibroblasts, are the mainstay of HLC-based, and multicellular human liver models (Fig. 5).[125–127]

iPSCs have the potential to expand indefinitely and differentiate into any cell type (Fig. 5). These characteristics make iPSCs an ideal source from which to obtain patient-specific cell types or to generate cells with specific genomic features resembling those of a particular human population using genome editing technologies.[128] Moreover, the pluripotent state of iPSC means they can act as a single source for the generation of the different hepatic cell types facilitating multicellular *in vitro* systems with the same genetic background or pre-existing disease.[129,130] This technology enables production of highly novel cellular models for studying unique and unexplored aspects of DILI such as cell-specific responses and multiorgan interaction.[131–134]

Early protocols for HLC generation were based on addition of stage-specific morphogenic cues mimicking hepatic embryonic development. More recently, growth factor-free approaches have been reported using small molecules that activate or mimic the effect of growth factors with a significant reduction in costs.[135–138] The iPSC-derived HLCs exhibit many hepatic functions, including serum protein production, urea synthesis and xenobiotic metabolism. Human iPSC-derived HLCs have similar attributes to the hepatoma cell line HepG2[139] and HepaRG cells (at least in 3D culture),[140] with lower metabolic activity compared with PHHs, and exhibit a mixed adult/foetal phenotype.[141] To improve HLCs' functionality, strategies include generation of HLCs in 3D using collagen matrices to achieve cellular polarity, induction of mature hepatocyte genes by small molecules, or mimicking liver maturation (postpartum) by exposing HLCs to bile acid synthesis components, drug metabolism, amino acid transport or microbiome composition.[142–146] Remarkably, supplementation of the growth medium with high concentrations of defined amino acids drove metabolic maturity (PHH levels of CYP activity) of both HLCs and HepG2 cells.[140]

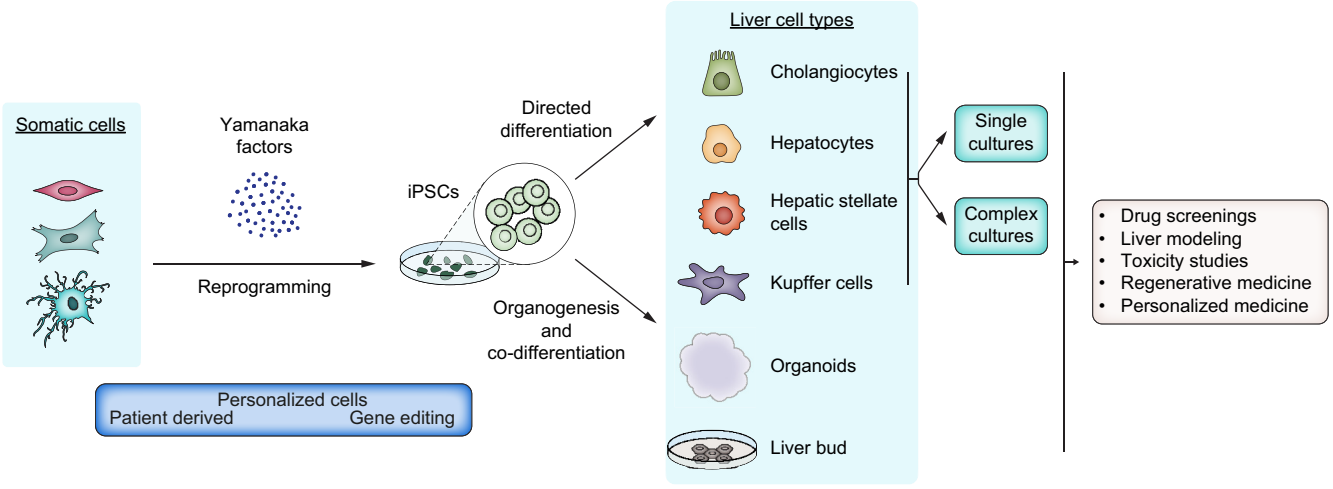

Fig. 5. Representation of methodological pathway for the use of iPSCs as *in vitro* liver models. Reprogrammed iPSCs can be differentiated following 2 strategies, directed differentiation to liver cell surrogates or by organogenic induction. Cells resulting from both strategies can be used in 2D or 3D for biotechnological and biomedical applications. iPSCs, induced pluripotent stem cells.

As all liver cell types are involved in disease processes and DILI, different approaches have evolved to generate HSCs, LSECs, and KCs. iPSC-derived human cholangiocytes with functional characteristics of primary cholangiocytes[147] have been developed and used to model disease (Alagille syndrome, chronic cholestasis due to reduced intrahepatic bile ducts) and for drug validation, highlighting hiPSCs' utility.[148,149]

NPCs are essential for liver homeostasis and immunological function, and play a key role in DILI. Therefore, generation of NPCs has been an intense area of research for development of complex *in vitro* systems. Tissue-resident human macrophages with KC characteristics have recently been generated, which exhibit low mismatch-background inflammatory response when co-cultured with hepatocytes.[150] HSCs also play a crucial role in response to injury/wound healing in the liver and are the main cell type responsible for not only extracellular matrix (ECM) production and degradation, but also ECM deposition and remodelling in fibrosis. In this regard, Coll and collaborators[151] generated HSC-like cells displaying features of quiescent HSCs that could be activated by inflammatory and pro-fibrogenic stimuli, such as lipopolysaccharide or TGF-β.[145]

A feature of pluripotent stem cells is their ability to recapitulate aspects of liver organogenesis/development in the dish. In a landmark study, Takebe *et al.* combined human umbilical vein endothelial cells (HUVECs) and mesenchymal stem cells with specified iPSC-derived hepatic endoderm. This approach resulted in the formation of a 3D structure resembling a liver bud, which upon transplantation into mice protected against DILI.[152] *In vitro* organogenic recapitulation of the liver by iPSCs has been used to generate liver organoids containing HLCs and other NPCs, which could be useful for liver disease modelling, toxicity testing and drug screening.[153–155]

As an alternative to primary hepatocytes, iPSC-derived cultures of HLCs have several applications for early preclinical hepatotoxicity assessment and drug screening in 2D and 3D culture systems.[156] iPSC-derived 3D organoids demonstrated a toxic response to clinically relevant concentrations of drugs withdrawn from the market due to hepatotoxicity.[157]

Key point

To elucidate critical pathogenic features of DILI, including genetic and immune factors, more faithful human *in vitro* models should include organotypic cultures and more focused investigational studies.

DILI is frequently characterised by common pathogenic mechanisms observed in chronic liver disease, such as inflammation, fibrosis and cholestasis. In order to link these responses to toxicity, more complex *in vitro* systems that capture aspects of *vivo* architecture, and contain different liver cell populations such as HSCs, cholangiocytes and inflammatory cells are required.

3D liver cell models

New *in vitro* cell and tissue engineering technologies are being developed to improve hepatocyte performance and are expected to generate more robust data on the potential risks of environmental agents and pharmaceuticals to humans. To achieve more efficient DILI prediction models, it will be necessary to develop new test systems that expand capabilities of target molecules more efficiently, reduce animal testing, increase drug development efficiency and are able to predict adverse effects.[158–166]

The major shortcomings of the currently available 2D *in vitro* liver systems are insufficient hepatocyte-like function and metabolic competence. A valid alternative for *in vitro* toxicology testing comprises more predictive cell models closer to the *in vivo* environment. These are summarised in Table 1, whilst bioengineering aspects of 3D liver systems, including the use of dynamic or static bioreactor devices are discussed further in the supplementary information (Sections 1.1 and 1.2).

Advanced technologies

Current hepatic-based microphysiological systems

Development of reliable human hepatic organotypic culture systems that are compatible with medium-high throughput screening (HTS) would have a significant impact on streamlining the drug development pipeline. Commercially available bioengineered liver models, including Emulate Inc., HμREL® Biochip, RegeneMed, Hepregen and LiverChip systems (Table 1), are based on hepatocyte-stromal cell interactions providing biomimetic cues to enhance hepatic phenotype/functionality. However, these systems utilise either heterologous hepatic co-cultures (rodent, primate or PHHs), combined with complex multi-step microfabrication manufacturing processes (*e.g.* soft-lithography, microfluidics), significantly increasing unit costs. Furthermore, the Hepregen system contains 3T3-J2 mouse fibroblasts, seeded on rat collagen-I, which can stabilise the function of the co-cultured PHHs. Such systems, however, are bio-incompatible, as they may introduce confounding variables in drug metabolism assays, given the presence of xeno-derived proteins and the fact that fibroblasts are not abundant in the functional liver acinus. Distinct challenges therefore remain with regard to realisation of a standardised, cost-effective, fully customised and widely available, organotypic *in vitro* human model.

'Liver-on-a-chip' models

Organ-on-a-chip (OoC) models are being developed as potentially improved experimental devices to overcome the limitations of current *in vitro* models of DILI. They are multicellular models connected by microfluidic flow that mimic features and functions of the organ represented. OoC models are rapidly emerging as an alternative to animal models to study human disease, while

Table 1. Advantages and limitations of complex *in vitro* hepatotoxicity cell culture systems.

	Advantages	Disadvantages	
Cell models			
ECM sandwich cultures[266–268]	Low complexity Hepatocytes regain polarity, maintain proper basolateral and canalicular transporters localisation and functional bile canaliculi Enables estimation of transport clearance, enzyme-transporter interplay, and bile acid mediated hepatotoxicity	Leakage, bile canaliculi damage and development of cholestasis in a time-dependent manner	[266–268]
Stirred bioreactors	Low complexity Scalable Long-term culture Co-culture of different cell types Enables perfusion Enables online monitoring	Requires specialised equipment Shear stress Variation in size/cell number/shape	[169,269–273]
Hollow-fiber bioreactors	Moderate throughput Counter-directional flow Scalable Long-term culture Possibility of PBPK studies Real-time monitoring	Complex system Microscopic evaluation is only possible in the end of the experiment Requires high number of cells Cell sampling not possible	[269,271,273–276]
Multi-well perfused bioreactors	High throughput Cells form 3D tissue constructs Sustained liver-like cell functionality Physiological shear stress Good correlation with in vivo clearance rates Ability for microscopic examination	Uses greater cell numbers and larger media volumes	[172,277]
Single-organ OoC models			
HμREL® Biochip	Moderate throughput Allows for multiple cell types and interaction between cell types Preservation of cell viability and metabolic competency Microscopic imaging and oxygen sensing Physiologically relevant ratios of liquid:cells and shear stress Requires less media and cells than traditional culture Good correlation with *in vivo* clearance rates	A complex system to establish and maintain Sample removal difficult No 3D tissue constructs	[278]
Multi-organ OoC models	Long term culture Improved cell functionality More physiologic model Enables tissue communication	Complex system Requires specialised equipment	
Microfluidic devices (*e.g. LiverChip system*)	Long term culture Laminar flow of cell culture media mimics the blood flow hemodynamics Stable low shear pressure Possibility to study multiple organs interaction Possibility of PBPK studies Real-time monitoring of metabolic function	A complex system to develop and establish Very low-sample and cell amounts	[278–282]
Liver bioprinting	Allows to build/design specific structures including endothelial and other cell types	A complex system to establish The printing process induces stress on cells	[279]

OoC, organ-on-a-chip. Adapted from [272,279–281,283].

academic research and industrial drug discovery have implemented this approach for drug target identification, validation, as well as efficacy and safety testing. Compared with 2D micro-engineered or bioprinted co-culture models, OoC models are generally less amenable to HTS due to their inherent complexity, and the need to incorporate biosensors for longitudinal real-time monitoring of biological events. Instead, they aim presently to address more complex physiological outcomes, including the preclinical phase of drug development.

Complex events of liver drug metabolism, as described for APAP, are a predominant feature of adverse drug reaction events leading to DILI. Therefore, to emulate organ physio-/pathophysiology in OoC models, integration of increasingly sophisticated and more realistic hepatic models, in combination with microfluidics and miniaturisation,

are crucial. This goal requires the convergence of tissue engineering processes and technologies to attain physiologically relevant systems. 3D bioprinting technology is a relatively new and rapidly evolving technology that is strategically placed to significantly enhance development and utility of biomimetic OoC models for preclinical applications. In addition, 3D bioprinting can be implemented as a stand-alone system to fabricate multicellular human hepatic models for HTS-amenable screening formats. Recent work has addressed major issues including limited structural complexity and resolution of many 3D-bioprinter systems, enabling assembly of complex vascular networks within 3D-printed hydrogels.[167] Many novel integrative bioengineering approaches have been adopted, with new designs and innovations continuing to evolve at a rapid pace. For example, Bhise *et al.*[168] developed an integrated Liver-on-a-chip platform for drug toxicity assessment, based on a bioreactor interfaced with a 3D-bioprinter. Hepatocyte (HepG2/C3A cells) spheroid-laden hydrogel constructs were bioprinted directly into a 'bioreactor' chip. This system exhibited a functional hepatic phenotype with an *in vivo*-like response to APAP toxicity. Ever more sophisticated, multi-cellular OoC models are now emerging. Digital light processing-based 3D-bioprinting systems can rapidly print tricultures of hiPSC-derived HLCs, endothelial and mesenchymal cells on hexagonal 3D-hydrogel scaffolds. The biomimetic liver lobule patterns demonstrated a robust functional metabolic profile (CYP expression) and suitability for hepatotoxicity screening and DILI prediction, as well as downstream personalised drug-screening applications.[169] Ingber's group recently developed a species-specific Liver-Chip that recapitulates complex liver cytoarchitecture, phenotypic profile and species-specific drug toxicities using rat, dog, and human cells.[170] Crucially, this system could identify both species-specific toxicity of drugs, such as APAP, and identify toxic events in hepatocyte and vascular channels.

Various levels of *in vivo*-like complexity have been achieved with improved PHH stability and functionality based on urea, albumin production, and CYP activity.[171] This has also been demonstrated using hepatocytes co-cultured with stromal cells present *in vivo* (*e.g.* liver KCs; sinusoidal epithelial cells).[172–174]

Whilst an organotypic human liver C3A cell line/HUVEC co-culture system demonstrated profound susceptibility to APAP-induced toxicity in endothelial cells (reflecting the situation found *in vivo*) compared with the monocultures; it is speculated that the vascular signals were likely hepatoprotective in the (APAP-resistant) co-cultures.[175] Proteomic analysis of LSECs may provide mechanistic insights allowing for the identification of sensitive and specific biomarkers through comparison and validation of omics data from preclinical animal models, *in vitro* human models and clinical biospecimens (see also sections AOP framework and Toxicogenomics approaches).

OoC models may also find a particularly relevant niche in the investigation of multi-organ systems, allowing for the examination of how bidirectional signals (*e.g.* metabolic, pro-inflammatory) in drug metabolism can affect other organs, and to study drug pharmacokinetics and ADME (absorption, distribution, metabolism, excretion). In particular, OoC models have attracted the interest of the pharmaceutical industry by demonstrating the ability to predict metabolic drug clearance rates in accordance with clinical data.[176] The circulation of drugs and metabolites between the liver and intestine has been explored using "liver-gut" models that replicate the intestinal barrier function. The parent compound phenacetin passed through the gut barrier and was metabolised to APAP by hepatic cells,[177] while a model including KCs mimicked inflammatory gut-liver interactions.[178]

Examples of dual-organ OoC models include liver/kidney interactions that recapitulated the nephrotoxicity of ifosfamide when metabolised by liver cells,[179] whilst skin and tumour compartments proved efficacious for substance testing.[180] More complex multi-organ models are under development, including: i) Liver/cardiac/muscle/neuronal system to investigate drug toxicity (doxorubicin, atorvastatin, valproic acid, APAP and N-acetyl-m-aminophenol),[181] and ii) a gut/skin/liver/kidney system in which organ-level functions were maintained for 28 days.[182] Recently, improved drug and toxicological readout was demonstrated with liver/lung/cardiac organoids derived from primary and iPSCs linked with microfluidics.[183]

Increasing OoC complexity has recently been explored with up to 10 interconnected organs to explore drug and metabolite bio-distribution together with a pharmacokinetic model.[184] At present, OoC technology is still in its infancy, and while it is demonstrating important tissue engineering principles and proof of concept, complexity and interactions resulting from multi-organ models are presently very challenging to decipher, whilst being inherently incompatible with HTS. This makes OoC models (and eventually multi-organ models) more suited to mechanistic studies and to predict safety and efficiency of compounds, as well as their pharmacokinetics, later in the drug discovery pipeline.

Non-invasive technologies to screen DILI models

High-content live cell confocal microscopy is particularly suited to screen DILI models. Real-time stress response pathways such as oxidative stress, UPR and DNA damage can be evaluated quantitatively at the single cell level,[185] and in individual 3D spheroids to screen for DILI.[186] In addition, novel

fluorescence dyes (*e.g.* thioflavin T) that react with aggregated proteins can be utilised for the determination of unfolded protein aggregations and thus to monitor ER stress induced by hepatotoxic drugs in live cell imaging settings.[187,188]

Mitochondrial stress and lysosomal dysfunction are also important mechanistic targets of DILI (see earlier section on emerging mechanisms and signalling cascades). Endosomes, cellular vesicle motion and mitochondrial fission-fusion are highly dynamic events (>3 μm/s) that require both super-resolution and fast acquisition (<<30 ms for 100 nm resolution). While PALM, STORM or STED super-resolution microscopy cannot currently provide such fast live imaging, a novel implementation of SIM using spinning disk confocal microscope optics has been developed[189,190] to achieve a spatial and temporal resolution of respectively, 120 nm,[189,190] and 6 μm/s.[191]

Optical screening of 3D organotypic models for DILI

3D tissue-like DILI models can be very challenging for optical microscopy. Novel high-resolution and super-resolution optical imaging methods that achieve 3D optical sectioning in real-time have very recently been developed,[192–197] providing insight into 3D *in vitro* model systems of DILI.[185,186,192–197] Finally, optical coherence tomography is particularly suited to imaging dense tissue-like structures at mm depth. Indeed, we have recently demonstrated label-free and non-destructive measurement of the hepatotoxic response to APAP in 3D human liver spheroids which correlated well with cellular metabolic activity assays.[198]

Super-resolution fluorescence nanoscopy

A new technological revolution in microscopic imaging called super-resolution fluorescence nanoscopy has been developed, enabling molecular scale resolution, localisation (<2 nm) and tracking of molecules, using a light microscope.[199] This affordable and flexible system (MINFLUX nanoscopy) will open up enormous possibilities in DILI, including 3D phenotypic profiling, imaging of protein complexes (drug-protein adducts) in pharmacological, ADME and toxicological studies, with simultaneous 2-colour (fluorophore) staining and recording. The Adaptive Optics system permits sharp deep tissue images down to 250 μm; while live-cell imaging of 3D-organoids to a depth of 37 μm into the sample can be attained using the easy3D STED imaging system. Fluorescence nanoscopy has already shown its applicability as a discovery tool in key areas transferable to mechanistic DILI studies. Indeed, nanoscopy studies of mitochondrial apoptotic mechanisms have demonstrated the assembly of Bax (BCL2 associated X, apoptosis regulator)/Bak (BCL2 antagonist/killer) proteins in the mitochondrial outer membrane, revealing a structural mechanism of membrane rupture and intracellular tracking of cancer-derived exosomes in *in vivo* mouse models.[196,200]

Impedance biosensing

As a non-invasive alternative to optical imaging, impedance-based cellular assays[201] have the advantage of enabling label-free and real-time monitoring of *in vitro* liver models, and they provide unique dynamics and quantitative insights into the impact of hepatotoxic drugs on cell-cell junctions.[15,202] Recent advances in impedance-based cellular assays also allow for the measurement of 3D models.[203] Advances in non-invasive imaging technologies in parallel with improved cell systems are powerful tools for improving DILI prediction and revealing critical DILI events, such as cellular reactive metabolite formation or oxidative stress at the molecular level and in real-time.

Emerging *in vivo* models of DILI

Mouse models with humanised livers

The biology and metabolism between mice and humans differ and hence the pharmacokinetics and toxicity profile of drugs can be substantially different between humans and mice. Although some limitations have been partially addressed using alternative approaches such as human hepatic cell lines, liver microsomes, PHHs or engineered human micro-livers,[204] these models are of limited predictive value regarding the pharmacokinetics and toxicity of drug metabolism *in vivo* and hence of limited relevance to human safety.[205] The development of chimeric models with bio-artificial livers repopulated with human adult hepatocytes could be an important advance for predicting human pharmacokinetics, drug interactions and *in vivo* safety (Fig. 6). This will be briefly described in the following sections.

Key point

Non-invasive technologies to screen DILI models may improve cell systems' readout and stand as a powerful tool for improving DILI prediction.

Generation of chimeric mice with humanised livers

Several models of chimeric mice suitable for repopulation with human adult hepatocytes have been developed over the years. The first chimeric mice with a partially humanised liver was described almost 20 years ago using a urokinase-type plasminogen activator-transgenic SCID (uPA$^{+/+}$/SCID) mouse.[124,206] The degree of repopulation of human hepatocytes in these initial studies was modest (about 15%), which was sufficient in the context of hepatitis viral infection, but inadequate to investigate human phamacokinetics (ADME) in mice *in vivo*. Tateno *et al.* generated chimeric uPA$^{+/+}$/SCID mice, replacing 70% of the liver with PHHs following anti-human complement factor treatment (estimated by serum levels of human albumin and cytokeratin 8/18 immunostaining[207]). Another model used TK-NOG mice expressing a herpes simplex virus type 1 thymidine kinase transgene in the liver of highly immunodeficient NOG mice.[208] Mouse hepatocyte deletion was

Key point

More realistic human-based *in vitro* models and humanised rodent models are urgently required to improve mechanistic understanding in order to de-risk DILI.

performed by exposure to ganciclovir followed by xenotransplantation of human hepatocytes. Both models exhibited substantial repopulation of human hepatocytes in the liver of chimeric mice and were useful in investigating the expression and activities of enzymes involved in drug metabolism.

Azuma *et al.*[209] also generated robust expansion of human hepatocytes in *Fah*$^{-/-}$/*Rag2*$^{-/-}$/*Il2rg*$^{-/-}$ (FRG) mice (humanised liver FRG mice). Fumaryl acetoacetate hydrolase (Fah) is involved in the tyrosine catabolic pathway, and genetic deletion of Fah acts as a molecular switch to control the demise of *Fah*$^{-/-}$ murine hepatocytes as its ablation causes massive damage to the endogenous mouse hepatocytes, driven by the accumulation of fumaroyl acetoacetate. The injection of human Fah$^{+/+}$ hepatocytes through the spleen, leads to the gradually repopulation of the liver of FRG mice over time.

Initial studies reported the use of these chimeric mice models to investigate the expression, levels and activities of human drug metabolising enzymes and transporters. For instance, the expression and enzyme activities of several CYPs in the livers of humanised uPA/SCID mice were similar to those in the donor liver or even greater than those found in cryopreserved human hepatocytes.[205,207,210] Furthermore, protein and enzyme activity levels of human UGT, sulfotransferase, *N*-acetyltransferase and glutathione-*s*-transferase in the humanised livers of uPA/SCID mice were reported to be similar to those in the donor liver.[211] Similar findings in terms of expression and enzymatic activities of CYPs with respect to the donor livers were reported in the liver of TK-NOG mice repopulated with human hepatocytes.[212] Therefore, these data validate the functional retention of human drug metabolising enzymes and transporters in the humanised livers of chimeric mice, further highlighting the utility in predicting relevant drug-drug interactions in humans.[213,214]

Drug metabolism and DILI in chimeric mice with humanised CYP and human liver chimeric mouse models

In addition to mouse models with humanised livers, several human CYP-transgenic mouse models have been generated. Most human CYP family members that are involved in xenobiotic metabolism, including members of the CYP1-CYP4 gene families, have been introduced into the mouse genome as a transgene (summarised recently in[215]). Although these models are potentially useful, metabolism of drugs in these transgenic humanised models reflects the action of a single human CYP transgene. Hence the relevance to human drug metabolism and safety may be of limited

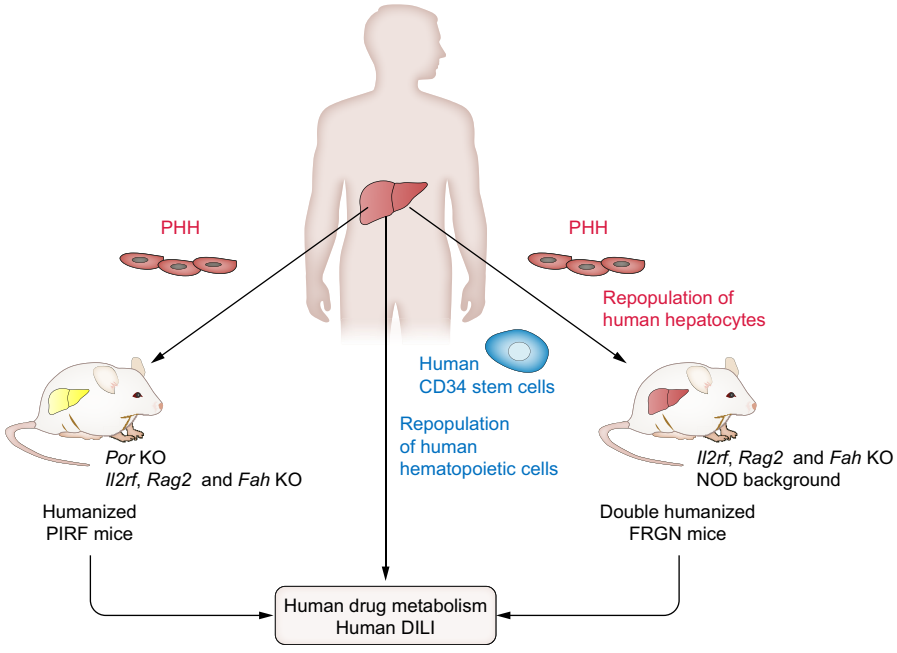

Fig. 6. Chimeric mice with humanised livers to model human DILI. Given the scarce availability of PHHs for drug toxicity screening, an alternative approach to potentially study human drug metabolism and impact in DILI is the xenotransplantation of adult PHHs into immunosuppressed mice engineered to selectively kill mouse hepatocytes while PHHs gradually repopulate the mouse liver. The FRGN model, which is amenable to double humanisation with PHHs and human hematopoietic cells, can be used to model human DILI. On the other hand, PIRF mice in which the *Por* gene has been deleted in a hepatocyte-specific manner, along with the deletion of *Il2rf*, *Rag2* and *Fah* genes, enable the xenotransplantation of PHHs and the study of human drug metabolism, which can be used to predict human DILI. DILI, drug-induced liver injury; KO, knockout; PHHs, primary human hepatocytes.

significance for human drug metabolism as this process may involve the function of multiple CYPs.

Moreover, the advantage of chimeric mouse models with humanised livers over CYP transgenic humanised models is that the former has been shown to generate human-specific metabolites and hence are of potential relevance for clinical drug development.[215] For instance, chimeric mice with humanised livers were recently used to study the metabolism of fenclozic acid, a drug that was developed as an alternative to high-dose therapy with aspirin in the mid 70s, and while it showed a good safety profile in experimental animals, it had to be withdrawn from late-clinical development because of hepatotoxicity. Interestingly, although fenclozic is off the market, these studies are useful to illustrate the ability of the chimeric mice to generate human-specific metabolites, such as the presence of fenclozic acid with side-chain extension in the plasma, which is not detected in conventional mice.[216] However, a drawback of the chimeric mice with humanised livers is that the remaining murine hepatocytes contain an expanded set of CYPs that form the major class of drug-metabolising enzymes. To exploit the potential of the human hepatocytes repopulating the livers of chimeric mice, and to provide xenobiotic metabolism, Barzi *et al.* generated a chimeric model in which the NADPH-cytochrome P450 oxidoreductase gene (POR) was knocked out in a liver-restricted manner in *Il2rg*$^{-/-}$/*Rag2*$^{-/-}$/*Fah*$^{-/-}$ (PIRF) mice. This provided a model with the advantage that drug metabolism in this engineered liver reflected the predominant activities of human CYPs.[217] Indeed, in response to the anticancer drug gefitinib or the retroviral drug atazanavir, the *POR*-deleted humanised PIRF mice developed higher levels of the major human metabolites and were consequently able to better predict human drug metabolism.[237]

Despite the relevance of APAP in human DILI, few studies have investigated APAP hepatotoxicity in chimeric mice with humanised livers. In this regard, Sato *et al.* examined the susceptibility of uPA^{++}/SCID mice whose livers were repopulated with human adult hepatocytes to APAP hepatotoxicity compared with control mice.[218] APAP administration resulted in vacuolation of hepatocytes and hepatocellular degeneration, leading to the detection of some areas of TUNEL-positive cells in the human hepatocyte zones. The hepatotoxic effects of APAP in the chimeric livers were milder than the severe liver injury observed in the control mice.[218] Further analysis indicated that APAP-related changes correlated with human CYP2E1 expression. In addition to these findings, a recent study reported on APAP hepatotoxicity in chimeric FRG mice bred on a NOD background (FRGN). These mice underwent xenotransplantation with human adult hepatocytes that had been pre-sensitised with valproic acid (VPA) pretreatment.[27] Comparable with wild-type mice, VPA pretreatment sensitised humanised FRGN mice to APAP hepatotoxicity, although the degree of injury was somewhat lower than that seen in wild-type mice, in line with findings in uPA^{++}/SCID chimeric mice. A caveat from these studies is that the degree of APAP-induced hepatotoxicity was milder with respect to wild-type mice. Given immature hepatocytes such as oval cells are reported to be resistant to APAP toxicity,[219] it is conceivable that the reduced toxicity of APAP in the chimeric mice (uPA$^{+/+}$-SCID or FRGN) may be due to functional immaturity of the repopulating human hepatocytes. In addition, whether a reduced presence of inflammatory cells in the liver of chimeric mice (*e.g.* macrophages or neutrophils) contribute to the milder hepatotoxicity remains to be investigated. In this regard, FRGN but not FRG mice are an amenable model for double humanisation following reconstitution with hepatocytes and haematopoietic cells. This is achieved by treatment with human CD34^{+} stem cells[220] and may be a useful approach to faithfully reproduce the observed hepatotoxicity of APAP in humans and to pinpoint the interactions between human hepatocytes and inflammatory cells. The impact of double humanization of FRGN mice with both human hepatocytes and hematopoietic cells in DILI still remains to be established.

Zebrafish, as a DILI model

Zebrafish is a vertebrate model organism widely used in development and genetics, which potentially provides a powerful tool for modelling DILI[221–223] (Fig. 7). Advantages of the zebrafish model include: a significant level of genomic, histological and functional similarity with humans; transparency of embryos and larvae, allowing for thorough imaging of the liver *in vivo*; and the availability of large numbers of offspring, increasing the feasibility and statistical power of drug screening experiments. Multiple types of assays have been described to characterise DILI in the zebrafish, including the detection of accumulated lipids in zebrafish larvae/liver as well as quantification of changes in liver size and numbers of liver cells using transgenic zebrafish lines expressing hepatic-specific fluorescent proteins.[224–226] Further applications of zebrafish for DILI modelling are discussed below and in the supplementary information (section 1.3).

Implications of DILI in clinical contexts

NAFLD and ageing as a susceptibility state for DILI

NAFLD, also now referred to as metabolic dysfunction-associated fatty liver disease (MAFLD), is currently the most prevalent chronic liver disease worldwide due to its association with the obesity epidemic. NAFLD is a spectrum of liver disorders beginning with steatosis, which can

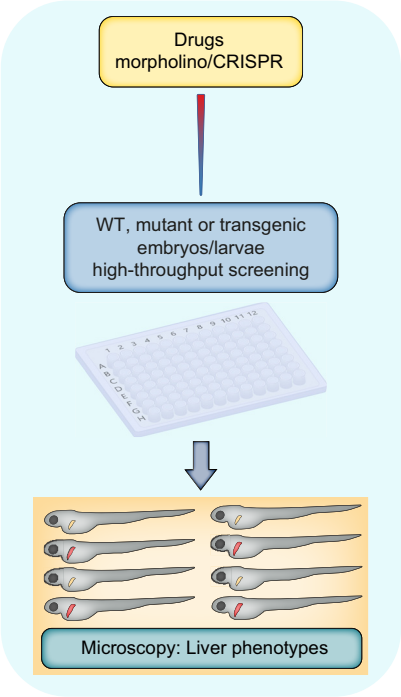

Fig. 7. Zebrafish to model DILI. Large-scale phenotypic assays in zebrafish embryos and larvae can be used for screening of drug- or genetically induced liver damage, using wild-type, mutant or transgenic zebrafish injected with different drugs of interest or morpholinos/CRISPR constructs to knockdown/knockout specific genes in a high throughput screening format. The effects of treatments are scored by microscopy manually or robotically starting from day 3 post-fertilisation (3dpf) to observe differences between the control and treatment groups in terms of differences in liver size and/or lipid accumulation. DILI, drug-induced liver injury; WT, wild-type.

progress to non-alcoholic steatohepatitis (NASH), cirrhosis and ultimately hepatocellular carcinoma. Although many drugs can induce steatosis as an early sign of potential hepatotoxicity, in this section we focus on growing evidence indicating that NAFLD can sensitise patients to DILI.

There are increasing clinical reports suggesting that patients suffering from obesity and NAFLD may be more susceptible to DILI.[55,227–231] This paradigm implies 2 possible scenarios. First, drugs such as APAP (in the context of overdose), halothane and isoflurane may cause more severe and/or more frequent ALF in individuals with NAFLD.[228,232] Second, pharmaceuticals such as irinotecan, methotrexate and tamoxifen seem to be more hepatotoxic in obese patients than in lean individuals by triggering the transition from steatosis to NASH, and/or worsening pre-existing steatosis, necroinflammation and fibrosis.[55,233,234] In spite of these emerging and distinct clinical situations, well-designed prospective clinical studies are urgently needed in order to identify the full repertoire of drugs which pose a particular risk in patients with NAFLD.[55,233] However, detection of DILI using the standard clinico-biological parameters could be difficult in patients with NASH,[235] while not all drugs necessarily pose a specific risk in NAFLD. For instance, hepatotoxicity induced by amiodarone and statins do not seem to be more frequent in patients with NAFLD.[233] Interestingly, the cytotoxicity induced by amiodarone, atorvastatin and lovastatin was not greater in a cellular model of NAFLD using the HepaRG cell line.[236] In contrast, the antiretroviral ritonavir was found to be less cytotoxic in this model, although clinical investigations are warranted to determine whether this observation can be confirmed in patients.

Except for a few drugs,[55,233] the mechanisms whereby some drugs are more hepatotoxic in NAFLD are complex and not well understood. Some drugs could induce more severe ALF in individuals with NAFLD because this disease is associated with altered activity of CYPs and other xenobiotic-metabolising enzymes (XMEs), which can increase the generation of toxic metabolites or conversely impair detoxification pathways.[55,228,233] For instance, human NAFLD is often associated with increased CYP2E1 activity and reduced CYP3A4 activity and also with higher glucuronide formation for some drugs such as APAP and lorazepam.[237–239] For drugs and other xenobiotics triggering the transition from simple fatty liver to NASH, or aggravating pre-existing liver lesions, experimental data strongly suggest a significant role for mitochondrial dysfunction, ER stress and ROS overproduction.[55,233,240]

In this context, preclinical models of NAFLD can be useful for distinct purposes. First, they can be used to confirm the specific toxicity of some pharmaceuticals in NAFLD, which might have been revealed during clinical investigations. Second, these experimental models can help to decipher the mechanisms whereby some drugs or other xenobiotics are more hepatotoxic in this liver disease. Lastly, these models might also be useful in preclinical safety studies, in particular for drug candidates that would be essentially prescribed in obese patients.

Numerous rodent models of NAFLD have been useful to study drug-induced hepatotoxicity.[233,241,242] However, it should be stressed that some of these models do not fully tally with the clinical situation, particularly in the context of NAFLD. For instance, leptin deficiency in genetically obese and diabetic ob/ob mice curbs the development of liver fibrosis,[243] and thus these mice are not appropriate to determine whether drugs are able to aggravate liver fibrosis;[233] whilst ob/ob mice do not present augmented hepatic CYP2E1 activity, which limits liver injury induced by APAP.[228,244] Moreover, mice fed a methionine choline-deficient diet consistently lose weight and can develop hypoglycaemia.[233,245] Finally, it should be mentioned that numerous types of energy-dense diets can be used to induce obesity

associated with simple fatty liver or NASH, but the degree of the different histopathological lesions can greatly vary between diets.[241,242,246] However, the extent of obesity and related metabolic disorders (*e.g.* insulin resistance and diabetes) as well as the severity of some liver lesions (*e.g.* steatosis and necroinflammation) are likely to influence the activity of different XMEs such as CYPs and UGTs.[55,239] Finally, zebrafish larvae fed lipid-enriched diets can also be used to evaluate hepatotoxicity in obesity and NAFLD. Although to the best of our knowledge this model has not been used for pharmaceuticals, recent investigations showed that obese zebrafish larvae were more sensitive to the hepatic toxicity of a mixture of benzo[a]pyrene and ethanol.[247] Interestingly, results collected in this zebrafish model were reproduced in a cellular model of NAFLD progression, as mentioned below.[247]

As with *in vivo* models, numerous *in vitro* NAFLD models have been established for various research purposes, particularly in the field of pharmacology and toxicology.[233,248–251] These cellular models of NAFLD are based on different types of cells (*i.e.* primary hepatocytes or cell lines, such as HuH7, HepG2, and HepaRG), fatty acids (used individually or in mixture), and duration of lipid overload (from a few hours to 15 days).[233,248–251] Interestingly, human iPSC-derived hepatocytes have recently been used in both 2D and 3D format to model NAFLD.[252] Another promising approach is the use of human iPSC-derived hepatocytes from patients with NAFLD including NASH, which might reproduce the inter-individual differences classically observed in DILI.[253,254] However, some experimental conditions might not be optimal for determining whether a drug is more toxic in the setting of NAFLD. For instance, the human hepatoma cell lines HuH7 and HepG2 do not have the full repertoire of XMEs,[255,256] whereas rodent hepatocytes do not have the same profile of drug metabolism as human hepatocytes, as discussed in the section 'Towards capturing hepatocellular complexity'. In addition, numerous studies have been performed in cells incubated with fatty acids for only a short duration of time (from a few hours to 2 or 3 days). Thus effects of prolonged or repeat-dose xenobiotic exposure are excluded, while this period may not be long enough to induce NAFLD-related alterations of XME expression and activity.[233]

Recently, a cellular model of NAFLD was established using differentiated and metabolically competent HepaRG cells incubated with 100 μM stearic acid for 7 days[257] (or with a mixture of stearic and oleic acids [150 μM each] for 14 days[236,240,247]). Notably, these *in vitro* models of NAFLD were characterised by enhanced CYP2E1 activity and reduced CY3A4 activity, thus reproducing what has been consistently observed in clinical studies, as previously mentioned. Of note, incubation of HepaRG cells with 100 μM stearic acid for 48 hours did not change CYP2E1 and CYP3A4 activities,[257] thus underscoring the importance of the duration of fatty acid exposure. Interestingly, a comparison by gene set enrichment analysis between the transcriptome GSE102536 dataset obtained in lipid-laden HepaRG cells[247] and the GSE61260 dataset obtained from biopsies of obese patients with fatty liver[258] revealed a highly significant correlation (p <0.001) concerning the upregulated genes (B. Fromenty and S. Bucher, unpublished data). These models disclosed higher cytotoxicity of APAP,[257] troglitazone[236] and a mixture of benzo[a]pyrene and ethanol[240,247] in NAFLD cells compared with the non-steatotic cells. Regarding APAP toxicity,[257] these *in vitro* investigations confirmed previous studies carried out in obese mice and humans with NAFLD.[228,232,244] Furthermore, mechanistic investigations showed that higher CYP2E1 activity in lipid-loaded HepaRG cells was, at least in part, responsible for higher APAP cytotoxicity.[257] Finally, it would be interesting to add cholesterol to fat-laden HepaRG cells in order to determine whether this lipid derivative could further enhance APAP cytotoxicity. Indeed, feeding wild-type mice a cholesterol-enriched diet (0.5%), which induces microvesicular steatosis and cholesterol accumulation in mitochondria, sensitises to APAP hepatotoxicity without fasting (JCFC and CGR, unpublished observations). As for NAFLD, there is evidence that older people might be at risk of DILI, at least with specific pharmaceuticals such as antimicrobials and cardiovascular agents.[259,260] In addition to the role of some specific medications, polypharmacy is deemed to be a risk factor for DILI in old age, although this does not seem to be related to impaired intrinsic drug metabolism.[259–261] Although old mice can be used as a preclinical model,[262] cellular models of hepatocyte ageing might also be useful. For instance, by using the senescence β-galactosidase assay, the occurrence of an ageing process has been observed in long-term confluent HepaRG cells,[263] and cellular senescence favours lipid deposition in the liver.[264,265] By using appropriate pre-clinical models, it would thus be interesting to determine whether NAFLD and ageing further increase the risk of DILI with some drugs.

Application of emerging mechanisms and approaches to human DILI

In the supplementary information, we select and briefly highlight a few examples of how some of the emerging mechanisms and approaches described above could be of value in human DILI.

Conclusions and future perspectives

Early pre-clinical identification of the toxic events leading to DILI is the primary goal and driver of major efforts in the pharmaceutical industry and academia to develop more realistic human-based models for DILI prediction. DILI represents an

unexpected liver injury caused by either prescribed or over-the counter drugs, which entails damage to hepatocytes as well as NPCs. Severe DILI is a serious clinical outcome and a major cause of ALF requiring liver transplantation. Besides its clinical relevance, DILI can be a primary reason for drug withdrawal from the market. Unlike intrinsic DILI, which is predictable, reproducible, and dose-dependent, iDILI is unpredictable, not strictly dose-dependent, and although rare it accounts for 10% to 15% of ALF cases in the United States. Due to its central role in biotransformation (metabolism) of xenobiotics entering the gastrointestinal tract, the liver is the main target of DILI and hepatocyte cell death stands as the major manifestation of DILI. The mechanisms inflicting hepatocellular demise in response to drugs are still not fully understood – representing a multifactorial process wherein activation of an immune response often contributes to overall death of hepatocytes and the spread of the damage to other NPCs. The limitation of our understanding of the underlying mechanisms and interplay between different players involved in DILI have hampered the delineation of effective therapies and the ability to accurately predict preclinical DILI development. This reflects the use of inadequate models used for DILI research. Indeed, unfortunately most experimental models currently used for DILI mechanistic studies do not adequately reflect the complexity of human biology and barely reproduce the features of DILI described in humans, highlighting the need to establish improved models for preclinical evaluation of DILI. Ideally these improved approaches should include experimental models that exhibit a higher concordance with human outcome through introduction of biological variation and complexity, leading to delineation of mechanistic and predictive signals that are relevant to DILI. In parallel, this paradigm shift in approaches to DILI must embrace a technological 'bioconvergence' encompassing multidisciplinary approaches across biology, engineering and medicine, such as coupling non-invasive imaging, multi-omics approaches, and conceptual frameworks (AOP) to organise modes and mechanisms of action, combined with microphysiological and other emerging 2D-3D multicellular platforms. Stepwise integration of appropriate human hepatic (acinar unit) cell types within microphysiological devices, as well as in novel 2D-3D platforms, and experimental decoupling of the acinar unit could be an important step in enhancing our understanding of DILI pathophysiology from single-cell to organ level – to solve the prediction dilemma in drug development. In principle, this approach may begin to discern what factors are lacking from current models to improve model relevance for DILI and thus better unravel mechanisms leading to DILI.

Bioconvergence offers a rich landscape for innovation, and includes the development of highly differentiated iPSC-derived hepatic tissues, which are accepted by regulatory agencies and the pharmaceutical industry because of the potential of this cell resource to populate OoC models and to develop multi-cellular organotypic 3D liver models with personalised medicine capability. Importantly, the use of chimeric mice with humanised livers and CYP biotransformation potential could offer transformational insights into specific aspects of DILI such as immune signals – and as a comparator system with next generation human-based *in vitro* models. This range of integrative approaches complemented with the development of state-of-the art non-invasive imaging methods for screening 2D-3D models within a flexible regulatory acceptance framework could improve the prediction of DILI and even iDILI with the possibility of identifying new targets for intervention and treatment.

Abbreviations

ADME, absorption, distribution, metabolism, excretion; AIF, apoptosis inducing factor; ALF, acute liver failure; AOP, adverse outcome pathways; ASK1, apoptosis signal-regulating kinase-1; ATF6, activating transcription factor 6; CYP, cytochrome P450; DILI, drug-induced liver injury; ECM, extracellular matrix; ER, endoplasmic reticulum; EVs, extracellular vesicles; GSH, glutathione; GSK-3β, glycogen synthase kinase 3β; HLCs, hepatocyte-like cells; HSCs, hepatic stellate cells; HTS, high throughput screening; HUVECs, human umbilical vein endothelial cells; iDILI, idiosyncratic DILI; IRE1α, inositol-requiring enzyme 1α; iPSC, inducible pluripotent stem cell; JNK, c-Jun-N-terminal kinase; KCs, Kupffer cells; LSEC, liver sinusoidal endothelial cells; MAPK, mitogen-activated protein kinase; Mpk, MAPK phosphatase; MPT, mitochondrial permeability transition; NAC, N-acetylcysteine; NAFLD, non-alcoholic fatty liver disease; NAPQI, N-acetyl-p-benzoquinone imine; NPCs, non-parenchymal cells; NSAID, non-steroidal anti-inflammatory drugs; OoC, organ-on-a-chip; PERK, protein kinase RNA-like ER kinase; PHHs, primary human hepatocytes; PRS, polygenic risk score; ROS, reactive oxygen species; STARD1, steroidogenic acute regulatory protein 1; TGF-β, transforming growth factor-β; TNF, tumour necrosis factor; Trx-1, thioredoxin-1; UGT, UDP-glucuronosultransferases; UPR, unfolded protein response; XME, xenobiotic-metabolising enzymes.

Financial support

This work is supported by COST Action CA17112, www.cost.eu. LJN is supported by AMMF 2018/117; with support from grants - BBSRC (Grant BB/L023687/1), EPSRC (IAA Grant PIII013). MK and JM were partly funded by the Fundação para a Ciencia e a Tecnologia (FCT, Portugal), through grants UID/BIM/0009/2020 for the Research Center for Toxicogenomics and Human Health-(ToxOmics) and PTDC/MED-TOX/29183/2017 and UIDB/04138/2020

for the iMed.ULisboa, respectively. OK is supported by the Scientific and Technological Research Council of Turkey (TUBITAK 116Z388) and COST Action CA17112. PSB is supported by the Fondo de Investigación Sanitaria Carlos III, cofinanced by the Fondo Europeo de Desarrollo Regional (FEDER), Unión Europea, "Una manera de hacer Europa" (FIS PI20/00765, PI17/00673), DTS18/00088 and Miguel Servet (CPII16/00041); AGAUR 2019_PROD_00055; NIHAAA 1U01AA026972-01. OL is supported by the Czech Ministry of Education, Youth and Sports (Project No. LTC19040). FJC is supported by the MINECO Retos SAF2016-78711 EXOHEP-CM S2017/BMD-3727, NanoLiver-CM Y2018/NMT-4949, ERAB Ref. EA 18/14, AMMF 2018/117, UCM-25-2019 and COST Action CA17112, RYC-2014-15242 and Gilead Liver Research 2018. JCFC and CGR are supported from grants and SAF2017-85877R, PID2019-111669RB-100 the CIBEREHD; the center grant P50AA011999 Southern California Research Center for ALPD and Cirrhosis funded by NIAAA/NIH; as well as support from AGAUR of the Generalitat de Catalunya SGR-2017-1112, the "ER stress-mitochondrial cholesterol axis in obesity-associated insulin resistance and comorbidities"-Ayudas FUNDACION BBVA and the Red Nacional 2018-102799-T de Enfermedades Metabólicas y Cáncer, Project 201916/31 from Fundació Marato TV3, and European Cooperation in Science & Technology (COST) ACTION CA17112 Prospective European Drug-Induced Liver Injury Network, www.cost.eu. GJS was partially supported by the Research Council of Norway through its Centres of Excellence funding scheme (Project number 262613).

Conflict of interest

The authors declare no conflicts of interest that pertain to this work.

Please refer to the accompanying ICMJE disclosure forms for further details.

Authors' contributions

Thematic structure and Introduction (LN, JCFC, FJC); DILI pathophysiology and emerging mechanisms in APAP (JCFC, FJC, CGR, HY); Emerging mechanisms and approaches (JMF-P, FR; LN); Towards capturing cellular complexity (LN, PSB, GS, JM, MK), Current hepatic-based microphysiological and advanced systems/non-invasive technologies to screen DILI models (LN, POB, OL, AD, AM, AE); Emerging in vivo models for DILI (OK, JCFC, CGR); Implications of DILI in clinical contexts (BF, JCFC; GPA, RJA, MIL).

Acknowledgements

The authors want to thank the support from the COST Action CA17112 and the valuable discussion with other experts and members of the PRO-EURO-DILI Network.

Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.jhep.2021.06.021.

References

Author names in bold designate shared co-first authorship

[1] Andrade RJ, Chalasani N, Bjornsson ES, Suzuki A, Kullak-Ublick GA, Watkins PB, et al. Drug-induced liver injury. Nat Rev Dis Prim 2019;5:58.
[2] Kaplowitz N. Idiosyncratic drug hepatotoxicity. Nat Rev Drug Discov 2005;4:489–499.
[3] Lucena MI, Kaplowitz N, Hallal H, Castiella A, Garcia-Bengoechea M, Otazua P, et al. Recurrent drug-induced liver injury (DILI) with different drugs in the Spanish Registry: the dilemma of the relationship to autoimmune hepatitis. J Hepatol 2011;55:820–827.
[4] Newsome PN, Henderson NC, Nelson LJ, Dabos C, Filippi C, Bellamy C, et al. Development of an invasively monitored porcine model of acetaminophen-induced acute liver failure. BMC Gastroenterol 2010;10:34.
[5] Bjornsson ES, Bergmann OM, Bjornsson HK, Kvaran RB, Olafsson S. Incidence, presentation, and outcomes in patients with drug-induced liver injury in the general population of Iceland. Gastroenterology 2013;144:1419–1425. 1425 e1411-1413; quiz e1419-1420.
[6] Reuben A, Tillman H, Fontana RJ, Davern T, McGuire B, Stravitz RT, et al. Outcomes in adults with acute liver failure between 1998 and 2013: an observational cohort study. Ann Intern Med 2016;164:724–732.
[7] Medina-Caliz I, Robles-Diaz M, Garcia-Munoz B, Stephens C, Ortega-Alonso A, Garcia-Cortes M, et al. Definition and risk factors for chronicity following acute idiosyncratic drug-induced liver injury. J Hepatol 2016;65:532–542.
[8] Niu H, Sanabria-Cabrera J, Alvarez-Alvarez I, Robles-Diaz M, Stankeviciute S, Aithal GP, et al. Prevention and management of idiosyncratic drug-induced liver injury: systematic review and meta-analysis of randomised clinical trials. Pharmacol Res 2021;164:105404.
[9] Stephens C, Robles-Diaz M, Medina-Caliz I, Garcia-Cortes M, Ortega-Alonso A, Sanabria-Cabrera J, et al. Comprehensive analysis and insights gained from long-term experience of the Spanish DILI Registry. J Hepatol 2021.
[10] Kullak-Ublick GA, Andrade RJ, Merz M, End P, Benesic A, Gerbes AL, et al. Drug-induced liver injury: recent advances in diagnosis and risk assessment. Gut 2017;66:1154–1164.
[11] Yuan L, Kaplowitz N. Mechanisms of drug-induced liver injury. Clin Liver Dis 2013;17:507–518. vii.
[12] Jaeschke H, Bajt ML. Intracellular signaling mechanisms of acetaminophen-induced liver cell death. Toxicol Sci 2006;89:31–41.
[13] Moles A, Torres S, Baulies A, Garcia-Ruiz C, Fernandez-Checa JC. Mitochondrial-lysosomal Axis in acetaminophen hepatotoxicity. Front Pharmacol 2018;9:453.
[14] Watkins PB, Kaplowitz N, Slattery JT, Colonese CR, Colucci SV, Stewart PW, et al. Aminotransferase elevations in healthy adults receiving 4 grams of acetaminophen daily: a randomized controlled trial. JAMA 2006;296:87–93.
[15] Gamal W, Treskes P, Samuel K, Sullivan GJ, Siller R, Srsen V, et al. Low-dose acetaminophen induces early disruption of cell-cell tight junctions in human hepatic cells and mouse liver. Sci Rep 2017;7:37541.
[16] Uetrecht J. Mechanisms of idiosyncratic drug-induced liver injury. Adv Pharmacol 2019;85:133–163.
[17] Boelsterli UA. Mechanisms of NSAID-induced hepatotoxicity: focus on nimesulide. Drug Saf 2002;25:633–648.
[18] O'Connor N, Dargan PI, Jones AL. Hepatocellular damage from non-steroidal anti-inflammatory drugs. QJM 2003;96:787–791.

[19] Tsutsumi S, Gotoh T, Tomisato W, Mima S, Hoshino T, Hwang HJ, et al. Endoplasmic reticulum stress response is involved in nonsteroidal anti-inflammatory drug-induced apoptosis. Cell Death Differ 2004;11:1009–1016.
[20] Bjornsson ES. Hepatotoxicity of statins and other lipid-lowering agents. Liver Int 2017;37:173–178.
[21] Karahalil B, Hare E, Koc G, Uslu I, Senturk K, Ozkan Y. Hepatotoxicity associated with statins. Arh Hig Rada Toksikol 2017;68:254–260.
[22] Seki E, Brenner DA, Karin M. A liver full of JNK: signaling in regulation of cell function and disease pathogenesis, and clinical approaches. Gastroenterology 2012;143:307–320.
[23] Johnson GL, Nakamura K. The c-jun kinase/stress-activated pathway: regulation, function and role in human disease. Biochim Biophys Acta 2007;1773:1341–1348.
[24] Matsumaru K, Ji C, Kaplowitz N. Mechanisms for sensitization to TNF-induced apoptosis by acute glutathione depletion in murine hepatocytes. Hepatology 2003;37:1425–1434.
[25] Shinohara M, Ybanez MD, Win S, Than TA, Jain S, Gaarde WA, et al. Silencing glycogen synthase kinase-3beta inhibits acetaminophen hepatotoxicity and attenuates JNK activation and loss of glutamate cysteine ligase and myeloid cell leukemia sequence 1. J Biol Chem 2010;285:8244–8255.
[26] Gunawan BK, Liu ZX, Han D, Hanawa N, Gaarde WA, Kaplowitz N. c-Jun N-terminal kinase plays a major role in murine acetaminophen hepatotoxicity. Gastroenterology 2006;131:165–178.
[27] **Torres S**, **Baulies A**, Insausti-Urkia N, Alarcon-Vila C, Fucho R, Solsona-Vilarrasa E, et al. Endoplasmic reticulum stress-induced upregulation of STARD1 promotes acetaminophen-induced acute liver failure. Gastroenterology 2019;157:552–568.
[28] Cubero FJ, Zoubek ME, Hu W, Peng J, Zhao G, Nevzorova YA, et al. Combined activities of JNK1 and JNK2 in hepatocytes protect against toxic liver injury. Gastroenterology 2016;150:968–981.
[29] Chang L, Kamata H, Solinas G, Luo JL, Maeda S, Venuprasad K, et al. The E3 ubiquitin ligase itch couples JNK activation to TNFalpha-induced cell death by inducing c-FLIP(L) turnover. Cell 2006;124:601–613.
[30] Zhao G, Hatting M, Nevzorova YA, Peng J, Hu W, Boekschoten MV, et al. Jnk1 in murine hepatic stellate cells is a crucial mediator of liver fibrogenesis. Gut 2014;63:1159–1172.
[31] Cederbaum AI, Yang L, Wang X, Wu D. CYP2E1 sensitizes the liver to LPS- and TNF alpha-induced toxicity via elevated oxidative and nitrosative stress and activation of ASK-1 and JNK mitogen-activated kinases. Int J Hepatol 2012;2012:582790.
[32] Zoubek ME, Woitok MM, Sydor S, Nelson LJ, Bechmann LP, Lucena MI, et al. Protective role of c-Jun N-terminal kinase-2 (JNK2) in ibuprofen-induced acute liver injury. J Pathol 2019;247:110–122.
[33] Mossanen JC, Krenkel O, Ergen C, Govaere O, Liepelt A, Puengel T, et al. Chemokine (C-C motif) receptor 2-positive monocytes aggravate the early phase of acetaminophen-induced acute liver injury. Hepatology 2016;64:1667–1682.
[34] Han D, Ybanez MD, Ahmadi S, Yeh K, Kaplowitz N. Redox regulation of tumor necrosis factor signaling. Antioxid Redox Signal 2009;11:2245–2263.
[35] Tournier C, Dong C, Turner TK, Jones SN, Flavell RA, Davis RJ. MKK7 is an essential component of the JNK signal transduction pathway activated by proinflammatory cytokines. Genes Dev 2001;15:1419–1426.
[36] Nakagawa H, Maeda S, Hikiba Y, Ohmae T, Shibata W, Yanai A, et al. Deletion of apoptosis signal-regulating kinase 1 attenuates acetaminophen-induced liver injury by inhibiting c-Jun N-terminal kinase activation. Gastroenterology 2008;135:1311–1321.
[37] Sharma M, Gadang V, Jaeschke A. Critical role for mixed-lineage kinase 3 in acetaminophen-induced hepatotoxicity. Mol Pharmacol 2012;82:1001–1007.
[38] Kallunki T, Deng T, Hibi M, Karin M. c-Jun can recruit JNK to phosphorylate dimerization partners via specific docking interactions. Cell 1996;87:929–939.
[39] Wancket LM, Meng X, Rogers LK, Liu Y. Mitogen-activated protein kinase phosphatase (Mkp)-1 protects mice against acetaminophen-induced hepatic injury. Toxicol Pathol 2012;40:1095–1105.
[40] Kouam AF, Yuan F, Njayou FN, He H, Tsayem RF, Oladejo BO, et al. Induction of Mkp-1 and nuclear translocation of Nrf2 by limonoids from khaya grandifoliola C.DC protect L-02 hepatocytes against acetaminophen-induced hepatotoxicity. Front Pharmacol 2017;8:653.
[41] Win S, Than TA, Min RW, Aghajan M, Kaplowitz N. c-Jun N-terminal kinase mediates mouse liver injury through a novel Sab (SH3BP5)-dependent pathway leading to inactivation of intramitochondrial Src. Hepatology 2016;63:1987–2003.
[42] Zhang J, Min RWM, Le K, Zhou S, Aghajan M, Than TA, et al. The role of MAP2 kinases and p38 kinase in acute murine liver injury models. Cell Death Dis 2017;8:e2903.
[43] Hanawa N, Shinohara M, Saberi B, Gaarde WA, Han D, Kaplowitz N. Role of JNK translocation to mitochondria leading to inhibition of mitochondria bioenergetics in acetaminophen-induced liver injury. J Biol Chem 2008;283:13565–13577.
[44] Win S, Than TA, Han D, Petrovic LM, Kaplowitz N. c-Jun N-terminal kinase (JNK)-dependent acute liver injury from acetaminophen or tumor necrosis factor (TNF) requires mitochondrial Sab protein expression in mice. J Biol Chem 2011;286:35071–35078.
[45] Saito C, Lemasters JJ, Jaeschke H. c-Jun N-terminal kinase modulates oxidant stress and peroxynitrite formation independent of inducible nitric oxide synthase in acetaminophen hepatotoxicity. Toxicol Appl Pharmacol 2010;246:8–17.
[46] Karch J, Kwong JQ, Burr AR, Sargent MA, Elrod JW, Peixoto PM, et al. Bax and Bak function as the outer membrane component of the mitochondrial permeability pore in regulating necrotic cell death in mice. Elife 2013;2:e00772.
[47] Norberg E, Orrenius S, Zhivotovsky B. Mitochondrial regulation of cell death: processing of apoptosis-inducing factor (AIF). Biochem Biophys Res Commun 2010;396:95–100.
[48] Bajt ML, Cover C, Lemasters JJ, Jaeschke H. Nuclear translocation of endonuclease G and apoptosis-inducing factor during acetaminophen-induced liver cell injury. Toxicol Sci 2006;94:217–225.
[49] Jaeschke H. Reactive oxygen and mechanisms of inflammatory liver injury: present concepts. J Gastroenterol Hepatol 2011;26(Suppl 1):173–179.
[50] Macias-Rodriguez RU, Inzaugarat ME, Ruiz-Margain A, Nelson LJ, Trautwein C, Cubero FJ. Reclassifying hepatic cell death during liver damage: ferroptosis-A novel form of non-apoptotic cell death? Int J Mol Sci 2020:21.
[51] Ye H, Nelson LJ, Gomez Del Moral M, Martinez-Naves E, Cubero FJ. Dissecting the molecular pathophysiology of drug-induced liver injury. World J Gastroenterol 2018;24:1373–1385.
[52] Uzi D, Barda L, Scaiewicz V, Mills M, Mueller T, Gonzalez-Rodriguez A, et al. CHOP is a critical regulator of acetaminophen-induced hepatotoxicity. J Hepatol 2013;59:495–503.
[53] Hur KY, So JS, Ruda V, Frank-Kamenetsky M, Fitzgerald K, Koteliansky V, et al. IRE1alpha activation protects mice against acetaminophen-induced hepatotoxicity. J Exp Med 2012;209:307–318.
[54] Baulies A, Ribas V, Nunez S, Torres S, Alarcon-Vila C, Martinez L, et al. Lysosomal cholesterol accumulation sensitizes to acetaminophen hepatotoxicity by impairing mitophagy. Sci Rep 2015;5:18017.
[55] Massart J, Begriche K, Moreau C, Fromenty B. Role of nonalcoholic fatty liver disease as risk factor for drug-induced hepatotoxicity. J Clin Transl Res 2017;3:212–232.
[56] **Caballero F**, Fernandez A, De Lacy AM, **Fernandez-Checa JC**, Caballeria J, **Garcia-Ruiz C**. Enhanced free cholesterol, SREBP-2 and StAR expression in human NASH. J Hepatol 2009;50:789–796.
[57] Saberi B, Ybanez MD, Johnson HS, Gaarde WA, Han D, Kaplowitz N. Protein kinase C (PKC) participates in acetaminophen hepatotoxicity through c-jun-N-terminal kinase (JNK)-dependent and -independent signaling pathways. Hepatology 2014;59:1543–1554.
[58] Ramachandran A, McGill MR, Xie Y, Ni HM, Ding WX, Jaeschke H. Receptor interacting protein kinase 3 is a critical early mediator of acetaminophen-induced hepatocyte necrosis in mice. Hepatology 2013;58:2099–2108.
[59] Dara L, Johnson H, Suda J, Win S, Gaarde W, Han D, et al. Receptor interacting protein kinase 1 mediates murine acetaminophen toxicity independent of the necrosome and not through necroptosis. Hepatology 2015;62:1847–1857.
[60] Ramachandran A, Jaeschke H. Mechanisms of acetaminophen hepatotoxicity and their translation to the human pathophysiology. J Clin Transl Res 2017;3:157–169.
[61] Poisson J, Lemoinne S, Boulanger C, Durand F, Moreau R, Valla D, et al. Liver sinusoidal endothelial cells: physiology and role in liver diseases. J Hepatol 2017;66:212–227.
[62] DeLeve LD, Wang X, Kaplowitz N, Shulman HM, Bart JA, van der Hoek A. Sinusoidal endothelial cells as a target for acetaminophen toxicity. Direct action versus requirement for hepatocyte activation in different mouse strains. Biochem Pharmacol 1997;53:1339–1345.

[63] Badmann A, Langsch S, Keogh A, Brunner T, Kaufmann T, Corazza N. TRAIL enhances paracetamol-induced liver sinusoidal endothelial cell death in a Bim- and Bid-dependent manner. Cell Death Dis 2012;3:e447.
[64] Teratani T, Tomita K, Suzuki T, Furuhashi H, Irie R, Hida S, et al. Free cholesterol accumulation in liver sinusoidal endothelial cells exacerbates acetaminophen hepatotoxicity via TLR9 signaling. J Hepatol 2017;67:780–790.
[65] Ni HM, Bockus A, Boggess N, Jaeschke H, Ding WX. Activation of autophagy protects against acetaminophen-induced hepatotoxicity. Hepatology 2012;55:222–232.
[66] Ni HM, McGill MR, Chao X, Du K, Williams JA, Xie Y, et al. Removal of acetaminophen protein adducts by autophagy protects against acetaminophen-induced liver injury in mice. J Hepatol 2016;65:354–362.
[67] Ni HM, Williams JA, Jaeschke H, Ding WX. Zonated induction of autophagy and mitochondrial spheroids limits acetaminophen-induced necrosis in the liver. Redox Biol 2013;1:427–432.
[68] Lin Z, Wu F, Lin S, Pan X, Jin L, Lu T, et al. Adiponectin protects against acetaminophen-induced mitochondrial dysfunction and acute liver injury by promoting autophagy in mice. J Hepatol 2014;61:825–831.
[69] Bryan HK, Olayanju A, Goldring CE, Park BK. The Nrf2 cell defence pathway: Keap1-dependent and -independent mechanisms of regulation. Biochem Pharmacol 2013;85:705–717.
[70] Kaspar JW, Niture SK, Jaiswal AK. Nrf2:INrf2 (Keap1) signaling in oxidative stress. Free Radic Biol Med 2009;47:1304–1309.
[71] Chan K, Han XD, Kan YW. An important function of Nrf2 in combating oxidative stress: detoxification of acetaminophen. Proc Natl Acad Sci U S A 2001;98:4611–4616.
[72] Wang L, Wei W, Xiao Q, Yang H, Ci X. Farrerol ameliorates APAP-induced hepatotoxicity via activation of Nrf2 and autophagy. Int J Biol Sci 2019;15:788–799.
[73] Royo F, Falcon-Perez JM. Liver extracellular vesicles in health and disease. J Extracell Vesic 2012;1.
[74] Vlassov AV, Magdaleno S, Setterquist R, Conrad R. Exosomes: current knowledge of their composition, biological functions, and diagnostic and therapeutic potentials. Biochim Biophys Acta 2012;1820:940–948.
[75] Mathivanan S, Fahner CJ, Reid GE, Simpson RJ. ExoCarta 2012: database of exosomal proteins, RNA and lipids. Nucl Acids Res 2012;40:D1241–D1244.
[76] Hessvik NP, Llorente A. Current knowledge on exosome biogenesis and release. Cell Mol Life Sci 2018;75:193–208.
[77] Abels ER, Breakefield XO. Introduction to extracellular vesicles: biogenesis, RNA cargo selection, content, release, and uptake. Cell Mol Neurobiol 2016;36:301–312.
[78] Masyuk AI, Masyuk TV, Larusso NF. Exosomes in the pathogenesis, diagnostics and therapeutics of liver diseases. J Hepatol 2013;59:621–625.
[79] Denzer K, van Eijk M, Kleijmeer MJ, Jakobson E, de Groot C, Geuze HJ. Follicular dendritic cells carry MHC class II-expressing microvesicles at their surface. J Immunol 2000;165:1259–1265.
[80] Holme PA, Solum NO, Brosstad F, Egberg N, Lindahl TL. Stimulated Glanzmann's thrombasthenia platelets produced microvesicles. Microvesiculation correlates better to exposure of procoagulant surface than to activation of GPIIb-IIIa. Thromb Haemost 1995;74:1533–1540.
[81] Kalra H, Simpson RJ, Ji H, Aikawa E, Altevogt P, Askenase P, et al. Vesiclepedia: a compendium for extracellular vesicles with continuous community annotation. Plos Biol 2012;10:e1001450.
[82] Conde-Vancells J, Rodriguez-Suarez E, Embade N, Gil D, Matthiesen R, Valle M, et al. Characterization and comprehensive proteome profiling of exosomes secreted by hepatocytes. J Proteome Res 2008;7:5157–5166.
[83] Herrera MB, Fonsato V, Gatti S, Deregibus MC, Sordi A, Cantarella D, et al. Human liver stem cell-derived microvesicles accelerate hepatic regeneration in hepatectomized rats. J Cell Mol Med 2010;14:1605–1618.
[84] Bala S, Petrasek J, Mundkur S, Catalano D, Levin I, Ward J, et al. Circulating microRNAs in exosomes indicate hepatocyte injury and inflammation in alcoholic, drug-induced, and inflammatory liver diseases. Hepatology 2012;56:1946–1957.
[85] Masyuk AI, Huang BQ, Ward CJ, Gradilone SA, Banales JM, Masyuk TV, et al. Biliary exosomes influence cholangiocyte regulatory mechanisms and proliferation through interaction with primary cilia. Am J Physiol Gastrointest Liver Physiol 2010;299:G990–G999.
[86] Povero D, Eguchi A, Li H, Johnson CD, Papouchado BG, Wree A, et al. Circulating extracellular vesicles with specific proteome and liver microRNAs are potential biomarkers for liver injury in experimental fatty liver disease. Plos One 2014;9:e113651.
[87] Palomo L, Mleczko JE, Azkargorta M, Conde-Vancells J, Gonzalez E, Elortza F, et al. Abundance of cytochromes in hepatic extracellular vesicles is altered by drugs related with drug-induced liver injury. Hepatol Commun 2018;2:1064–1079.
[88] Holman NS, Mosedale M, Wolf KK, LeCluyse EL, Watkins PB. Subtoxic alterations in hepatocyte-derived exosomes: an early step in drug-induced liver injury? Toxicol Sci 2016;151:365–375.
[89] Royo F, Schlangen K, Palomo L, Gonzalez E, Conde-Vancells J, Berisa A, et al. Transcriptome of extracellular vesicles released by hepatocytes. Plos One 2013;8:e68693.
[90] Rodriguez-Suarez E, Gonzalez E, Hughes C, Conde-Vancells J, Rudella A, Royo F, et al. Quantitative proteomic analysis of hepatocyte-secreted extracellular vesicles reveals candidate markers for liver toxicity. J Proteomics 2014;103:227–240.
[91] Cho YE, Im EJ, Moon PG, Mezey E, Song BJ, Baek MC. Increased liver-specific proteins in circulating extracellular vesicles as potential biomarkers for drug- and alcohol-induced liver injury. Plos One 2017;12:e0172463.
[92] Yang X, Weng Z, Mendrick DL, Shi Q. Circulating extracellular vesicles as a potential source of new biomarkers of drug-induced liver injury. Toxicol Lett 2014;225:401–406.
[93] Rowland A, Ruanglertboon W, van Dyk M, Wijayakumara D, Wood LS, Meech R, et al. Plasma extracellular nanovesicle (exosome)-derived biomarkers for drug metabolism pathways: a novel approach to characterize variability in drug exposure. Br J Clin Pharmacol 2019;85:216–226.
[94] Kumar S, Sinha N, Gerth KA, Rahman MA, Yallapu MM, Midde NM. Specific packaging and circulation of cytochromes P450, especially 2E1 isozyme, in human plasma exosomes and their implications in cellular communications. Biochem Biophys Res Commun 2017;491:675–680.
[95] Royo F, Moreno L, Mleczko J, Palomo L, Gonzalez E, Cabrera D, et al. Hepatocyte-secreted extracellular vesicles modify blood metabolome and endothelial function by an arginase-dependent mechanism. Sci Rep 2017;7:42798.
[96] Royo F, Cossio U, Ruiz de Angulo A, Llop J, Falcon-Perez JM. Modification of the glycosylation of extracellular vesicles alters their biodistribution in mice. Nanoscale 2019;11:1531–1537.
[97] Rahman MA, Kodidela S, Sinha N, Haque S, Shukla PK, Rao R, et al. Plasma exosomes exacerbate alcohol- and acetaminophen-induced toxicity via CYP2E1 pathway. Sci Rep 2019;9:6571.
[98] Ogese MO, Jenkins RE, Adair K, Tailor A, Meng X, Faulkner L, et al. Exosomal transport of hepatocyte-derived drug-modified proteins to the immune system. Hepatology 2019;70:1732–1749.
[99] Vinken M, Hengstler JG. Characterization of hepatocyte-based in vitro systems for reliable toxicity testing. Arch Toxicol 2018;92:2981–2986.
[100] Daly AK. Using genome-wide association studies to identify genes important in serious adverse drug reactions. Annu Rev Pharmacol Toxicol 2012;52:21–35.
[101] Kohonen P, Parkkinen JA, Willighagen EL, Ceder R, Wennerberg K, Kaski S, et al. A transcriptomics data-driven gene space accurately predicts liver cytopathology and drug-induced liver injury. Nat Commun 2017;8:15932.
[102] Leist M, Ghallab A, Graepel R, Marchan R, Hassan R, Bennekou SH, et al. Adverse outcome pathways: opportunities, limitations and open questions. Arch Toxicol 2017;91:3477–3505.
[103] Watanabe KH, Andersen ME, Basu N, Carvan 3rd MJ, Crofton KM, King KA, et al. Defining and modeling known adverse outcome pathways: domoic acid and neuronal signaling as a case study. Environ Toxicol Chem 2011;30:9–21.
[104] Vinken M. Adverse outcome pathways and drug-induced liver injury testing. Chem Res Toxicol 2015;28:1391–1397.
[105] Woodhead JL, Yang K, Brouwer KL, Siler SQ, Stahl SH, Ambroso JL, et al. Mechanistic modeling reveals the critical knowledge gaps in bile acid-mediated DILI. CPT Pharmacomet Syst Pharmacol 2014;3:e123.
[106] Longo DM, Woodhead JL, Walker P, Heredi-Szabo K, Mogyorosi K, Wolenski FS, et al. Quantitative systems toxicology analysis of in vitro mechanistic assays reveals importance of bile acid accumulation and mitochondrial dysfunction in TAK-875-induced liver injury. Toxicol Sci 2019;167:458–467.
[107] Shoda LK, Battista C, Siler SQ, Pisetsky DS, Watkins PB, Howell BA. Mechanistic modelling of drug-induced liver injury: investigating the role of innate immune responses. Gene Regul Syst Bio 2017;11:1177625017696074.

[108] Khadka KK, Chen M, Liu Z, Tong W, Wang D. Integrating adverse outcome pathways (AOPs) and high throughput in vitro assays for better risk evaluations, a study with drug-induced liver injury (DILI). ALTEX 2020;37:187–196.
[109] Kuepfer L, Clayton O, Thiel C, Cordes H, Nudischer R, Blank LM, et al. A model-based assay design to reproduce in vivo patterns of acute drug-induced toxicity. Arch Toxicol 2018;92:553–555.
[110] Ruden DM, Gurdziel K, Aschner M. Frontiers in toxicogenomics in the twenty-first century-the grand challenge: to understand how the genome and epigenome interact with the toxic environment at the single-cell, whole-organism, and multi-generational level. Front Genet 2017;8:173.
[111] Igarashi Y, Nakatsu N, Yamashita T, Ono A, Ohno Y, Urushidani T, et al. Open TG-GATEs: a large-scale toxicogenomics database. Nucl Acids Res 2015;43:D921–D927.
[112] **Nicoletti P**, **Aithal GP**, Bjornsson ES, Andrade RJ, Sawle A, Arrese M, et al. Association of liver injury from specific drugs, or groups of drugs, with polymorphisms in HLA and other genes in a genome-wide association study. Gastroenterology 2017;152:1078–1089.
[113] Koido M, Kawakami E, Fukumura J, Noguchi Y, Ohori M, Nio Y, et al. Polygenic architecture informs potential vulnerability to drug-induced liver injury. Nat Med 2020;26:1541–1548.
[114] Kolodziejczyk AA, Federici S, Zmora N, Mohapatra G, Dori-Bachash M, Hornstein S, et al. Acute liver failure is regulated by MYC- and microbiome-dependent programs. Nat Med 2020;26:1899–1911.
[115] Cuykx M, Rodrigues RM, Laukens K, Vanhaecke T, Covaci A. In vitro assessment of hepatotoxicity by metabolomics: a review. Arch Toxicol 2018;92:3007–3029.
[116] Dash A, Inman W, Hoffmaster K, Sevidal S, Kelly J, Obach RS, et al. Liver tissue engineering in the evaluation of drug safety. Expert Opin Drug Metab Toxicol 2009;5:1159–1174.
[117] Guillouzo A, Guguen-Guillouzo C. Evolving concepts in liver tissue modeling and implications for in vitro toxicology. Expert Opin Drug Metab Toxicol 2008;4:1279–1294.
[118] Tsiaoussis J, Newsome PN, Nelson LJ, Hayes PC, Plevris JN. Which hepatocyte will it be? Hepatocyte choice for bioartificial liver support systems. Liver Transpl 2001;7:2–10.
[119] Prot JM, Leclerc E. The current status of alternatives to animal testing and predictive toxicology methods using liver microfluidic biochips. Ann Biomed Eng 2012;40:1228–1243.
[120] Sistare FD, Dieterle F, Troth S, Holder DJ, Gerhold D, Andrews-Cleavenger D, et al. Towards consensus practices to qualify safety biomarkers for use in early drug development. Nat Biotechnol 2010;28:446–454.
[121] Nelson LJ, Morgan K, Treskes P, Samuel K, Henderson CJ, LeBled C, et al. Human hepatic HepaRG cells maintain an organotypic phenotype with high intrinsic CYP450 activity/metabolism and significantly outperform standard HepG2/C3A cells for pharmaceutical and therapeutic applications. Basic Clin Pharmacol Toxicol 2017;120:30–37.
[122] Zhou Y, Shen JX, Lauschke VM. Comprehensive evaluation of organotypic and microphysiological liver models for prediction of drug-induced liver injury. Front Pharmacol 2019;10:1093.
[123] Weaver RJ, Blomme EA, Chadwick AE, Copple IM, Gerets HHJ, Goldring CE, et al. Managing the challenge of drug-induced liver injury: a roadmap for the development and deployment of preclinical predictive models. Nat Rev Drug Discov 2020;19:131–148.
[124] Dandri M, Burda MR, Torok E, Pollok JM, Iwanska A, Sommer G, et al. Repopulation of mouse liver with human hepatocytes and in vivo infection with hepatitis B virus. Hepatology 2001;33:981–988.
[125] Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell 2006;126:663–676.
[126] Park IH, Arora N, Huo H, Maherali N, Ahfeldt T, Shimamura A, et al. Disease-specific induced pluripotent stem cells. Cell 2008;134:877–886.
[127] Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL, Tian S, et al. Induced pluripotent stem cell lines derived from human somatic cells. Science 2007;318:1917–1920.
[128] Ding Q, Regan SN, Xia Y, Oostrom LA, Cowan CA, Musunuru K. Enhanced efficiency of human pluripotent stem cell genome editing through replacing TALENs with CRISPRs. Cell Stem Cell 2013;12:393–394.
[129] Shi Y, Inoue H, Wu JC, Yamanaka S. Induced pluripotent stem cell technology: a decade of progress. Nat Rev Drug Discov 2017;16:115–130.
[130] Fiorotto R, Amenduni M, Mariotti V, Fabris L, Spirli C, Strazzabosco M. Liver diseases in the dish: iPSC and organoids as a new approach to modeling liver diseases. Biochim Biophys Acta Mol Basis Dis 2019;1865:920–928.
[131] Song Z, Cai J, Liu Y, Zhao D, Yong J, Duo S, et al. Efficient generation of hepatocyte-like cells from human induced pluripotent stem cells. Cell Res 2009;19:1233–1242.
[132] Sullivan GJ, Hay DC, Park IH, Fletcher J, Hannoun Z, Payne CM, et al. Generation of functional human hepatic endoderm from human induced pluripotent stem cells. Hepatology 2010;51:329–335.
[133] Si-Tayeb K, Noto FK, Nagaoka M, Li J, Battle MA, Duris C, et al. Highly efficient generation of human hepatocyte-like cells from induced pluripotent stem cells. Hepatology 2010;51:297–305.
[134] Sancho-Bru P, Roelandt P, Narain N, Pauwelyn K, Notelaers T, Shimizu T, et al. Directed differentiation of murine-induced pluripotent stem cells to functional hepatocyte-like cells. J Hepatol 2011;54:98–107.
[135] Siller R, Sullivan GJ. Rapid screening of the endodermal differentiation potential of human pluripotent stem cells. Curr Protoc Stem Cell Biol 2017;43:1G 7 1–1G 7 23.
[136] Siller R, Greenhough S, Naumovska E, Sullivan GJ. Small-molecule-driven hepatocyte differentiation of human pluripotent stem cells. Stem Cell Rep 2015;4:939–952.
[137] Mathapati S, Siller R, Impellizzeri AA, Lycke M, Vegheim K, Almaas R, et al. Small-Molecule-directed hepatocyte-like cell differentiation of human pluripotent stem cells. Curr Protoc Stem Cell Biol 2016;38:1G 6 1–1G 6 18.
[138] Du C, Feng Y, Qiu D, Xu Y, Pang M, Cai N, et al. Highly efficient and expedited hepatic differentiation from human pluripotent stem cells by pure small-molecule cocktails. Stem Cell Res Ther 2018;9:58.
[139] Kvist AJ, Kanebratt KP, Walentinsson A, Palmgren H, O'Hara M, Bjorkbom A, et al. Critical differences in drug metabolic properties of human hepatic cellular models, including primary human hepatocytes, stem cell derived hepatocytes, and hepatoma cell lines. Biochem Pharmacol 2018;155:124–140.
[140] Bell CC, Lauschke VM, Vorrink SU, Palmgren H, Duffin R, Andersson TB, et al. Transcriptional, functional, and mechanistic comparisons of stem cell-derived hepatocytes, HepaRG cells, and three-dimensional human hepatocyte spheroids as predictive in vitro systems for drug-induced liver injury. Drug Metab Disp 2017;45:419–429.
[141] Baxter M, Withey S, Harrison S, Segeritz CP, Zhang F, Atkinson-Dell R, et al. Phenotypic and functional analyses show stem cell-derived hepatocyte-like cells better mimic fetal rather than adult hepatocytes. J Hepatol 2015;62:581–589.
[142] Gieseck 3rd RL, Hannan NR, Bort R, Hanley NA, Drake RA, Cameron GW, et al. Maturation of induced pluripotent stem cell derived hepatocytes by 3D-culture. Plos One 2014;9:e86372.
[143] Shan J, Schwartz RE, Ross NT, Logan DJ, Thomas D, Duncan SA, et al. Identification of small molecules for human hepatocyte expansion and iPS differentiation. Nat Chem Biol 2013;9:514–520.
[144] Beath SV. Hepatic function and physiology in the newborn. Semin Neonatol 2003;8:337–346.
[145] Avior Y, Levy G, Zimerman M, Kitsberg D, Schwartz R, Sadeh R, et al. Microbial-derived lithocholic acid and vitamin K2 drive the metabolic maturation of pluripotent stem cells-derived and fetal hepatocytes. Hepatology 2015;62:265–278.
[146] Boon R, Kumar M, Tricot T, Elia I, Ordovas L, Jacobs F, et al. Amino acid levels determine metabolism and CYP450 function of hepatocytes and hepatoma cell lines. Nat Commun 2020;11:1393.
[147] Dianat N, Dubois-Pot-Schneider H, Steichen C, Desterke C, Leclerc P, Raveux A, et al. Generation of functional cholangiocyte-like cells from human pluripotent stem cells and HepaRG cells. Hepatology 2014;60:700–714.
[148] Sampaziotis F, de Brito MC, Madrigal P, Bertero A, Saeb-Parsy K, Soares FAC, et al. Cholangiocytes derived from human induced pluripotent stem cells for disease modeling and drug validation. Nat Biotechnol 2015;33:845–852.
[149] Ogawa M, Ogawa S, Bear CE, Ahmadi S, Chin S, Li B, et al. Directed differentiation of cholangiocytes from human pluripotent stem cells. Nat Biotechnol 2015;33:853–861.
[150] Tasnim F, Xing J, Huang X, Mo S, Wei X, Tan MH, et al. Generation of mature kupffer cells from human induced pluripotent stem cells. Biomaterials 2019;192:377–391.
[151] Coll M, Perea L, Boon R, Leite SB, Vallverdu J, Mannaerts I, et al. Generation of hepatic stellate cells from human pluripotent stem cells enables in vitro modeling of liver fibrosis. Cell Stem Cell 2018;23. 101-113 e107.

[152] Takebe T, Sekine K, Enomura M, Koike H, Kimura M, Ogaeri T, et al. Vascularized and functional human liver from an iPSC-derived organ bud transplant. Nature 2013;499:481–484.
[153] Guan Y, Xu D, Garfin PM, Ehmer U, Hurwitz M, Enns G, et al. Human hepatic organoids for the analysis of human genetic diseases. JCI Insight 2017;2.
[154] Koui Y, Kido T, Ito T, Oyama H, Chen SW, Katou Y, et al. An in vitro human liver model by iPSC-derived parenchymal and non-parenchymal cells. Stem Cell Rep 2017;9:490–498.
[155] Ouchi R, Togo S, Kimura M, Shinozawa T, Koido M, Koike H, et al. Modeling steatohepatitis in humans with pluripotent stem cell-derived organoids. Cell Metab 2019;30. 374-384 e376.
[156] Gomez-Lechon MJ, Tolosa L. Human hepatocytes derived from pluripotent stem cells: a promising cell model for drug hepatotoxicity screening. Arch Toxicol 2016;90:2049–2061.
[157] Mun SJ, Ryu JS, Lee MO, Son YS, Oh SJ, Cho HS, et al. Generation of expandable human pluripotent stem cell-derived hepatocyte-like liver organoids. J Hepatol 2019;71:970–985.
[158] Andersen ME, Krewski D. Toxicity testing in the 21st century: bringing the vision to life. Toxicol Sci 2009;107:324–330.
[159] Baharvand H, Hashemi SM, Kazemi Ashtiani S, Farrokhi A. Differentiation of human embryonic stem cells into hepatocytes in 2D and 3D culture systems in vitro. Int J Dev Biol 2006;50:645–652.
[160] Davila JC, Cezar GG, Thiede M, Strom S, Miki T, Trosko J. Use and application of stem cells in toxicology. Toxicol Sci 2004;79:214–223.
[161] Giri S, Nieber K, Bader A. Hepatotoxicity and hepatic metabolism of available drugs: current problems and possible solutions in preclinical stages. Expert Opin Drug Metab Toxicol 2010;6:895–917.
[162] Krewski D, Andersen ME, Mantus E, Zeise L. Toxicity testing in the 21st century: implications for human health risk assessment. Risk Anal 2009;29:474–479.
[163] Mandenius CF, Andersson TB, Alves PM, Batzl-Hartmann C, Bjorquist P, Carrondo MJ, et al. Toward preclinical predictive drug testing for metabolism and hepatotoxicity by using in vitro models derived from human embryonic stem cells and human cell lines - a report on the Vitrocellomics EU-project. Altern Lab Anim 2011;39:147–171.
[164] Shukla SJ, Huang R, Austin CP, Xia M. The future of toxicity testing: a focus on in vitro methods using a quantitative high-throughput screening platform. Drug Discov Today 2010;15:997–1007.
[165] Trosko JE. Commentary on "Toxicity testing in the 21st century: a vision and a strategy": stem cells and cell-cell communication as fundamental targets in assessing the potential toxicity of chemicals. Hum Exp Toxicol 2010;29:21–29.
[166] Balls M. Modern alternative approaches to the problem of drug-induced liver injury. Altern Lab Anim 2011;39:103–107.
[167] Lee A, Hudson AR, Shiwarski DJ, Tashman JW, Hinton TJ, Yerneni S, et al. 3D bioprinting of collagen to rebuild components of the human heart. Science 2019;365:482–487.
[168] Bhise NS, Manoharan V, Massa S, Tamayol A, Ghaderi M, Miscuglio M, et al. A liver-on-a-chip platform with bioprinted hepatic spheroids. Biofabrication 2016;8:014101.
[169] Ma X, Qu X, Zhu W, Li YS, Yuan S, Zhang H, et al. Deterministically patterned biomimetic human iPSC-derived hepatic model via rapid 3D bioprinting. Proc Natl Acad Sci U S A 2016;113:2206–2211.
[170] Jang KJ, Otieno MA, Ronxhi J, Lim HK, Ewart L, Kodella KR, et al. Reproducing human and cross-species drug toxicities using a Liver-Chip. Sci Transl Med 2019;11.
[171] Khetani SR, Berger DR, Ballinger KR, Davidson MD, Lin C, Ware BR. Microengineered liver tissues for drug testing. J Lab Auto 2015;20:216–250.
[172] Domansky K, Inman W, Serdy J, Dash A, Lim MH, Griffith LG. Perfused multiwell plate for 3D liver tissue engineering. Lab Chip 2010;10:51–58.
[173] Vernetti LA, Senutovitch N, Boltz R, DeBiasio R, Shun TY, Gough A, et al. A human liver microphysiology platform for investigating physiology, drug safety, and disease models. Exp Biol Med (Maywood) 2016;241:101–114.
[174] Prodanov L, Jindal R, Bale SS, Hegde M, McCarty WJ, Golberg I, et al. Long-term maintenance of a microfluidic 3D human liver sinusoid. Biotechnol Bioeng 2016;113:241–246.
[175] Nelson LJ, Navarro M, Treskes P, Samuel K, Tura-Ceide O, Morley SD, et al. Acetaminophen cytotoxicity is ameliorated in a human liver organotypic co-culture model. Sci Rep 2015;5:17455.
[176] Baudoin R, Prot JM, Nicolas G, Brocheton J, Brochot C, Legallais C, et al. Evaluation of seven drug metabolisms and clearances by cryopreserved human primary hepatocytes cultivated in microfluidic biochips. Xenobiotica 2013;43:140–152.
[177] Bricks T, Paullier P, Legendre A, Fleury MJ, Zeller P, Merlier F, et al. Development of a new microfluidic platform integrating co-cultures of intestinal and liver cell lines. Toxicol In Vitro 2014;28:885–895.
[178] Chen WLK, Edington C, Suter E, Yu J, Velazquez JJ, Velazquez JG, et al. Integrated gut/liver microphysiological systems elucidates inflammatory inter-tissue crosstalk. Biotechnol Bioeng 2017;114:2648–2659.
[179] Choucha-Snouber L, Aninat C, Grsicom L, Madalinski G, Brochot C, Poleni PE, et al. Investigation of ifosfamide nephrotoxicity induced in a liver-kidney co-culture biochip. Biotechnol Bioeng 2013;110:597–608.
[180] Wagner I, Materne EM, Brincker S, Sussbier U, Fradrich C, Busek M, et al. A dynamic multi-organ-chip for long-term cultivation and substance testing proven by 3D human liver and skin tissue co-culture. Lab Chip 2013;13:3538–3547.
[181] Oleaga C, Bernabini C, Smith AS, Srinivasan B, Jackson M, McLamb W, et al. Multi-Organ toxicity demonstration in a functional human in vitro system composed of four organs. Sci Rep 2016;6:20030.
[182] Maschmeyer I, Lorenz AK, Schimek K, Hasenberg T, Ramme AP, Hubner J, et al. A four-organ-chip for interconnected long-term co-culture of human intestine, liver, skin and kidney equivalents. Lab Chip 2015;15:2688–2699.
[183] Skardal A, Murphy SV, Devarasetty M, Mead I, Kang HW, Seol YJ, et al. Multi-tissue interactions in an integrated three-tissue organ-on-a-chip platform. Sci Rep 2017;7:8837.
[184] Edington CD, Chen WLK, Geishecker E, Kassis T, Soenksen LR, Bhushan BM, et al. Interconnected microphysiological systems for quantitative biology and pharmacology studies. Sci Rep 2018;8:4530.
[185] Wink S, Hiemstra SW, Huppelschoten S, Klip JE, van de Water B. Dynamic imaging of adaptive stress response pathway activation for prediction of drug induced liver injury. Arch Toxicol 2018;92:1797–1814.
[186] Hiemstra S, Ramaiahgari SC, Wink S, Callegaro G, Coonen M, Meerman J, et al. High-throughput confocal imaging of differentiated 3D liver-like spheroid cellular stress response reporters for identification of drug-induced liver injury liability. Arch Toxicol 2019;93:2895–2911.
[187] Beriault DR, Werstuck GH. Detection and quantification of endoplasmic reticulum stress in living cells using the fluorescent compound, Thioflavin T. Biochim Biophys Acta 2013;1833:2293–2301.
[188] Lunova M, Smolkova B, Uzhytchak M, Janouskova KZ, Jirsa M, Egorova D, et al. Light-induced modulation of the mitochondrial respiratory chain activity: possibilities and limitations. Cell Mol Life Sci 2020;77:2815–2838.
[189] Hayashi S. Resolution doubling using confocal microscopy via analogy with structured illumination microscopy. Jpn J Appl Phys 2016;55:082501.
[190] Hayashi S, Okada Y. Ultrafast superresolution fluorescence imaging with spinning disk confocal microscope optics. Mol Biol Cell 2015;26:1743–1751.
[191] York AG, Chandris P, Nogare DD, Head J, Wawrzusin P, Fischer RS, et al. Instant super-resolution imaging in live cells and embryos via analog image processing. Nat Meth 2013;10:1122–1126.
[192] Richardson DS, Lichtman JW. Clarifying tissue clearing. Cell 2015;162:246–257.
[193] Richardson DS, Lichtman JW. SnapShot: tissue clearing. Cell 2017;171.496-496 e491.
[194] Rios AC, Fu NY, Lindeman GJ, Visvader JE. In situ identification of bipotent stem cells in the mammary gland. Nature 2014;506:322–327.
[195] Rios AC, Clevers H. Imaging organoids: a bright future ahead. Nat Meth 2018;15:24–26.
[196] Sahl SJ, Hell SW, Jakobs S. Fluorescence nanoscopy in cell biology. Nat Rev Mol Cell Biol 2017;18:685–701.
[197] Schermelleh L, Heintzmann R, Leonhardt H. A guide to super-resolution fluorescence microscopy. J Cell Biol 2010;190:165–175.
[198] Martucci NJ, Morgan K, Anderson GW, Hayes PC, Plevris JN, Nelson LJ, et al. Nondestructive optical toxicity assays of 3D liver spheroids with optical coherence tomography. Adv Biosyst 2018;2:1700212.
[199] Gwosch KC, Pape JK, Balzarotti F, Hoess P, Ellenberg J, Ries J, et al. MINFLUX nanoscopy delivers 3D multicolor nanometer resolution in cells. Nat Meth 2020;17:217–224.

[200] Hell SW, Wichmann J. Breaking the diffraction resolution limit by stimulated emission: stimulated-emission-depletion fluorescence microscopy. Opt Lett 1994;19:780–782.
[201] Gamal W, Wu H, Underwood I, Jia J, Smith S, Bagnaninchi PO. Impedance-based cellular assays for regenerative medicine. Philos Trans R Soc Lond B Biol Sci 2018;373.
[202] Morgan K, Martucci N, Kozlowska A, Gamal W, Brzeszczynski F, Treskes P, et al. Chlorpromazine toxicity is associated with disruption of cell membrane integrity and initiation of a pro-inflammatory response in the HepaRG hepatic cell line. Biomed Pharmacother 2019;111:1408–1416.
[203] Wu H, Yang Y, Bagnaninchi PO, Jia J. Electrical impedance tomography for real-time and label-free cellular viability assays of 3D tumour spheroids. Analyst 2018;143:4189–4198.
[204] Mancio-Silva L, Fleming HE, Miller AB, Milstein S, Liebow A, Haslett P, et al. Improving drug discovery by nucleic acid delivery in engineered human microlivers. Cell Metab 2019;29. 727-735 e723.
[205] Katoh M, Tateno C, Yoshizato K, Yokoi T. Chimeric mice with humanized liver. Toxicology 2008;246:9–17.
[206] Mercer DF, Schiller DE, Elliott JF, Douglas DN, Hao C, Rinfret A, et al. Hepatitis C virus replication in mice with chimeric human livers. Nat Med 2001;7:927–933.
[207] Tateno C, Yoshizane Y, Saito N, Kataoka M, Utoh R, Yamasaki C, et al. Near completely humanized liver in mice shows human-type metabolic responses to drugs. Am J Pathol 2004;165:901–912.
[208] Hasegawa M, Kawai K, Mitsui T, Taniguchi K, Monnai M, Wakui M, et al. The reconstituted 'humanized liver' in TK-NOG mice is mature and functional. Biochem Biophys Res Commun 2011;405:405–410.
[209] Azuma H, Paulk N, Ranade A, Dorrell C, Al-Dhalimy M, Ellis E, et al. Robust expansion of human hepatocytes in Fah-/-/Rag2-/-/Il2rg-/- mice. Nat Biotechnol 2007;25:903–910.
[210] Yamasaki C, Kataoka M, Kato Y, Kakuni M, Usuda S, Ohzone Y, et al. In vitro evaluation of cytochrome P450 and glucuronidation activities in hepatocytes isolated from liver-humanized mice. Drug Metab Pharmacokinet 2010;25:539–550.
[211] Katoh M, Matsui T, Nakajima M, Tateno C, Kataoka M, Soeno Y, et al. Expression of human cytochromes P450 in chimeric mice with humanized liver. Drug Metab Disp 2004;32:1402–1410.
[212] Hasegawa M, Tahara H, Inoue R, Kakuni M, Tateno C, Ushiki J. Investigation of drug-drug interactions caused by human pregnane X receptor-mediated induction of CYP3A4 and CYP2C subfamilies in chimeric mice with a humanized liver. Drug Metab Disp 2012;40:474–480.
[213] Naritomi Y, Sanoh S, Ohta S. Chimeric mice with humanized liver: application in drug metabolism and pharmacokinetics studies for drug discovery. Drug Metab Pharmacokinet 2018;33:31–39.
[214] Nishimura T, Hu Y, Wu M, Pham E, Suemizu H, Elazar M, et al. Using chimeric mice with humanized livers to predict human drug metabolism and a drug-drug interaction. J Pharmacol Exp Ther 2013;344:388–396.
[215] Bissig KD, Han W, Barzi M, Kovalchuk N, Ding L, Fan X, et al. P450-Humanized and human liver chimeric mouse models for studying xenobiotic metabolism and toxicity. Drug Metab Disp 2018;46:1734–1744.
[216] Ekdahl A, Weidolf L, Baginski M, Morikawa Y, Thompson RA, Wilson ID. The metabolic fate of fenclozic acid in chimeric mice with a humanized liver. Arch Toxicol 2018;92:2819–2828.
[217] Barzi M, Pankowicz FP, Zorman B, Liu X, Legras X, Yang D, et al. A novel humanized mouse lacking murine P450 oxidoreductase for studying human drug metabolism. Nat Commun 2017;8:39.
[218] Sato Y, Yamada H, Iwasaki K, Tateno C, Yokoi T, Yoshizato K, et al. Human hepatocytes can repopulate mouse liver: histopathology of the liver in human hepatocyte-transplanted chimeric mice and toxicologic responses to acetaminophen. Toxicol Pathol 2008;36:581–591.
[219] Kofman AV, Morgan G, Kirschenbaum A, Osbeck J, Hussain M, Swenson S, et al. Dose- and time-dependent oval cell reaction in acetaminophen-induced murine liver injury. Hepatology 2005;41:1252–1261.
[220] Wilson EM, Bial J, Tarlow B, Bial G, Jensen B, Greiner DL, et al. Extensive double humanization of both liver and hematopoiesis in FRGN mice. Stem Cell Res 2014;13:404–412.
[221] Vernetti LA, Vogt A, Gough A, Taylor DL. Evolution of experimental models of the liver to predict human drug hepatotoxicity and efficacy. Clin Liver Dis 2017;21:197–214.
[222] Wang S, Miller SR, Ober EA, Sadler KC. Making it new again: insight into liver development, regeneration, and disease from zebrafish research. Curr Top Dev Biol 2017;124:161–195.
[223] Vliegenthart AD, Tucker CS, Del Pozo J, Dear JW. Zebrafish as model organisms for studying drug-induced liver injury. Br J Clin Pharmacol 2014;78:1217–1227.
[224] Yu Q, Huo J, Zhang Y, Liu K, Cai Y, Xiang T, et al. Tamoxifen-induced hepatotoxicity via lipid accumulation and inflammation in zebrafish. Chemosphere 2020;239:124705.
[225] Nguyen XB, Kislyuk S, Pham DH, Kecskes A, Maes J, Cabooter D, et al. Cell imaging counting as a novel ex vivo approach for investigating drug-induced hepatotoxicity in zebrafish larvae. Int J Mol Sci 2017;18.
[226] Zhang Y, Han L, He Q, Chen W, Sun C, Wang X, et al. A rapid assessment for predicting drug-induced hepatotoxicity using zebrafish. J Pharmacol Toxicol Meth 2017;84:102–110.
[227] Tarantino G, Conca P, Basile V, Gentile A, Capone D, Polichetti G, et al. A prospective study of acute drug-induced liver injury in patients suffering from non-alcoholic fatty liver disease. Hepatol Res 2007;37:410–415.
[228] Michaut A, Moreau C, Robin MA, Fromenty B. Acetaminophen-induced liver injury in obesity and nonalcoholic fatty liver disease. Liver Int 2014;34:e171–e179.
[229] Bessone F, Dirchwolf M, Rodil MA, Razori MV, Roma MG. Review article: drug-induced liver injury in the context of nonalcoholic fatty liver disease - a physiopathological and clinical integrated view. Aliment Pharmacol Ther 2018;48:892–913.
[230] Lammert C, Imler T, Teal E, Chalasani N. Patients with chronic liver disease suggestive of nonalcoholic fatty liver disease may Be at higher risk for drug-induced liver injury. Clin Gastroenterol Hepatol 2019;17:2814–2815.
[231] Li X, Gao P, Niu J. Metabolic comorbidities and risk of development and severity of drug-induced liver injury. Biomed Res Int 2019;2019:8764093.
[232] Garcia-Roman R, Frances R. Acetaminophen-induced liver damage in hepatic steatosis. Clin Pharmacol Ther 2020;107:1068–1081.
[233] Allard J, Le Guillou D, Begriche K, Fromenty B. Drug-induced liver injury in obesity and nonalcoholic fatty liver disease. Adv Pharmacol 2019;85:75–107.
[234] Meunier L, Larrey D. Chemotherapy-associated steatohepatitis. Ann Hepatol 2020.
[235] Regev A, Palmer M, Avigan MI, Dimick-Santos L, Treem WR, Marcinak JF, et al. Consensus: guidelines: best practices for detection, assessment and management of suspected acute drug-induced liver injury during clinical trials in patients with nonalcoholic steatohepatitis. Aliment Pharmacol Ther 2019;49:702–713.
[236] Le Guillou D, Bucher S, Begriche K, Hoet D, Lombes A, Labbe G, et al. Drug-induced alterations of mitochondrial DNA homeostasis in steatotic and nonsteatotic HepaRG cells. J Pharmacol Exp Ther 2018;365:711–726.
[237] Aubert J, Begriche K, Knockaert L, Robin MA, Fromenty B. Increased expression of cytochrome P450 2E1 in nonalcoholic fatty liver disease: mechanisms and pathophysiological role. Clin Res Hepatol Gastroenterol 2011;35:630–637.
[238] Brill MJ, Diepstraten J, van Rongen A, van Kralingen S, van den Anker JN, Knibbe CA. Impact of obesity on drug metabolism and elimination in adults and children. Clin Pharmacokinet 2012;51:277–304.
[239] Cobbina E, Akhlaghi F. Non-alcoholic fatty liver disease (NAFLD) - pathogenesis, classification, and effect on drug metabolizing enzymes and transporters. Drug Metab Rev 2017;49:197–211.
[240] Bucher S, Le Guillou D, Allard J, Pinon G, Begriche K, Tete A, et al. Possible involvement of mitochondrial dysfunction and oxidative stress in a cellular model of NAFLD progression induced by benzo[a]pyrene/ethanol CoExposure. Oxid Med Cell Longev 2018;2018:4396403.
[241] Haczeyni F, Yeh MM, Ioannou GN, Leclercq IA, Goldin R, Dan YY, et al. Mouse models of non-alcoholic steatohepatitis: a reflection on recent literature. J Gastroenterol Hepatol 2018;33:1312–1320.
[242] Santhekadur PK, Kumar DP, Sanyal AJ. Preclinical models of non-alcoholic fatty liver disease. J Hepatol 2018;68:230–237.

[243] Trak-Smayra V, Paradis V, Massart J, Nasser S, Jebara V, Fromenty B. Pathology of the liver in obese and diabetic ob/ob and db/db mice fed a standard or high-calorie diet. Int J Exp Pathol 2011;92:413–421.
[244] Aubert J, Begriche K, Delannoy M, Morel I, Pajaud J, Ribault C, et al. Differences in early acetaminophen hepatotoxicity between obese ob/ob and db/db mice. J Pharmacol Exp Ther 2012;342:676–687.
[245] Arao Y, Kawai H, Kamimura K, Kobayashi T, Nakano O, Hayatsu M, et al. Effect of methionine/choline-deficient diet and high-fat diet-induced steatohepatitis on mitochondrial homeostasis in mice. Biochem Biophys Res Commun 2020;527:365–371.
[246] Denk H, Abuja PM, Zatloukal K. Animal models of NAFLD from the pathologist's point of view. Biochim Biophys Acta Mol Basis Dis 2019;1865:929–942.
[247] Bucher S, Tete A, Podechard N, Liamin M, Le Guillou D, Chevanne M, et al. Co-exposure to benzo[a]pyrene and ethanol induces a pathological progression of liver steatosis in vitro and in vivo. Sci Rep 2018;8:5963.
[248] Luo Y, Rana P, Will Y. Palmitate increases the susceptibility of cells to drug-induced toxicity: an in vitro method to identify drugs with potential contraindications in patients with metabolic disease. Toxicol Sci 2012;129:346–362.
[249] Breher-Esch S, Sahini N, Trincone A, Wallstab C, Borlak J. Genomics of lipid-laden human hepatocyte cultures enables drug target screening for the treatment of non-alcoholic fatty liver disease. BMC Med Genomics 2018;11:111.
[250] Tanner N, Kubik L, Luckert C, Thomas M, Hofmann U, Zanger UM, et al. Regulation of drug metabolism by the interplay of inflammatory signaling, steatosis, and xeno-sensing receptors in HepaRG cells. Drug Metab Disp 2018;46:326–335.
[251] Pant A, Rondini EA, Kocarek TA. Farnesol induces fatty acid oxidation and decreases triglyceride accumulation in steatotic HepaRG cells. Toxicol Appl Pharmacol 2019;365:61–70.
[252] Wang Y, Wang H, Deng P, Tao T, Liu H, Wu S, et al. Modeling human nonalcoholic fatty liver disease (NAFLD) with an organoids-on-a-chip system. ACS Biomater Sci Eng 2020;6:5734–5743.
[253] Duwaerts CC, Le Guillou D, Her CL, Phillips NJ, Willenbring H, Mattis AN, et al. Induced pluripotent stem cell-derived hepatocytes from patients with nonalcoholic fatty liver disease display a disease-specific gene expression profile. Gastroenterology 2021.
[254] Gurevich I, Burton SA, Munn C, Ohshima M, Goedland ME, Czysz K, et al. iPSC-derived hepatocytes generated from NASH donors provide a valuable platform for disease modeling and drug discovery. Biol Open 2020;9.
[255] Wilkening S, Stahl F, Bader A. Comparison of primary human hepatocytes and hepatoma cell line Hepg2 with regard to their biotransformation properties. Drug Metab Disp 2003;31:1035–1042.
[256] Choi S, Sainz Jr B, Corcoran P, Uprichard S, Jeong H. Characterization of increased drug metabolism activity in dimethyl sulfoxide (DMSO)-treated Huh7 hepatoma cells. Xenobiotica 2009;39:205–217.
[257] Michaut A, Le Guillou D, Moreau C, Bucher S, McGill MR, Martinais S, et al. A cellular model to study drug-induced liver injury in nonalcoholic fatty liver disease: application to acetaminophen. Toxicol Appl Pharmacol 2016;292:40–55.
[258] Horvath S, Erhart W, Brosch M, Ammerpohl O, von Schonfels W, Ahrens M, et al. Obesity accelerates epigenetic aging of human liver. Proc Natl Acad Sci U S A 2014;111:15538–15543.
[259] Stine JG, Sateesh P, Lewis JH. Drug-induced liver injury in the elderly. Curr Gastroenterol Rep 2013;15:299.
[260] Lucena MI, Sanabria J, Garcia-Cortes M, Stephens C, Andrade RJ. Drug-induced liver injury in older people. Lancet Gastroenterol Hepatol 2020;5:862–874.
[261] Waring RH, Harris RM, Mitchell SC. Drug metabolism in the elderly: a multifactorial problem? Maturitas 2017;100:27–32.
[262] Tanimizu N, Ichinohe N, Suzuki H, Mitaka T. Prolonged oxidative stress and delayed tissue repair exacerbate acetaminophen-induced liver injury in aged mice. Aging (Albany NY) 2020;12:18907–18927.
[263] Pernelle K, Le Guevel R, Glaise D, Stasio CG, Le Charpentier T, Bouaita B, et al. Automated detection of hepatotoxic compounds in human hepatocytes using HepaRG cells and image-based analysis of mitochondrial dysfunction with JC-1 dye. Toxicol Appl Pharmacol 2011;254:256–266.
[264] Gregg SQ, Gutierrez V, Robinson AR, Woodell T, Nakao A, Ross MA, et al. A mouse model of accelerated liver aging caused by a defect in DNA repair. Hepatology 2012;55:609–621.
[265] Ogrodnik M, Miwa S, Tchkonia T, Tiniakos D, Wilson CL, Lahat A, et al. Cellular senescence drives age-dependent hepatic steatosis. Nat Commun 2017;8:15691.
[266] Godoy P, Widera A, Schmidt-Heck W, Campos G, Meyer C, Cadenas C, et al. Gene network activity in cultivated primary hepatocytes is highly similar to diseased mammalian liver tissue. Arch Toxicol 2016;90:2513–2529.
[267] Deharde D, Schneider C, Hiller T, Fischer N, Kegel V, Lubberstedt M, et al. Bile canaliculi formation and biliary transport in 3D sandwich-cultured hepatocytes in dependence of the extracellular matrix composition. Arch Toxicol 2016;90:2497–2511.
[268] Bell CC, Hendriks DF, Moro SM, Ellis E, Walsh J, Renblom A, et al. Characterization of primary human hepatocyte spheroids as a model system for drug-induced liver injury, liver function and disease. Sci Rep 2016;6:25187.
[269] Lubberstedt M, Muller-Vieira U, Mayer M, Biemel KM, Knospel F, Knobeloch D, et al. HepaRG human hepatic cell line utility as a surrogate for primary human hepatocytes in drug metabolism assessment in vitro. J Pharmacol Toxicol Meth 2011;63:59–68.
[270] Miranda JP, Rodrigues A, Tostoes RM, Leite S, Zimmerman H, Carrondo MJ, et al. Extending hepatocyte functionality for drug-testing applications using high-viscosity alginate-encapsulated three-dimensional cultures in bioreactors. Tissue Eng C Meth 2010;16:1223–1232.
[271] Mueller D, Tascher G, Muller-Vieira U, Knobeloch D, Nuessler AK, Zeilinger K, et al. In-depth physiological characterization of primary human hepatocytes in a 3D hollow-fiber bioreactor. J Tissue Eng Regen Med 2011;5:e207–e218.
[272] Serras AS, Rodrigues JS, Cipriano M, Rodrigues AV, Oliveira NG, Miranda JP. A critical perspective on 3D liver models for drug metabolism and toxicology studies. Front Cell Dev Biol 2021;9:626805.
[273] Zeilinger K, Schreiter T, Darnell M, Soderdahl T, Lubberstedt M, Dillner B, et al. Scaling down of a clinical three-dimensional perfusion multicompartment hollow fiber liver bioreactor developed for extracorporeal liver support to an analytical scale device useful for hepatic pharmacological in vitro studies. Tissue Eng Part C Meth 2011;17:549–556.
[274] Darnell M, Schreiter T, Zeilinger K, Urbaniak T, Soderdahl T, Rossberg I, et al. Cytochrome P450-dependent metabolism in HepaRG cells cultured in a dynamic three-dimensional bioreactor. Drug Metab Disp 2011;39:1131–1138.
[275] Darnell M, Ulvestad M, Ellis E, Weidolf L, Andersson TB. In vitro evaluation of major in vivo drug metabolic pathways using primary human hepatocytes and HepaRG cells in suspension and a dynamic three-dimensional bioreactor system. J Pharmacol Exp Ther 2012;343:134–144.
[276] Hoffmann SA, Muller-Vieira U, Biemel K, Knobeloch D, Heydel S, Lubberstedt M, et al. Analysis of drug metabolism activities in a miniaturized liver cell bioreactor for use in pharmacological studies. Biotechnol Bioeng 2012;109:3172–3181.
[277] Freyer N, Knospel F, Strahl N, Amini L, Schrade P, Bachmann S, et al. Hepatic differentiation of human induced pluripotent stem cells in a perfused three-dimensional multicompartment bioreactor. Biores Open Access 2016;5:235–248.
[278] Novik E, Maguire TJ, Chao P, Cheng KC, Yarmush ML. A microfluidic hepatic coculture platform for cell-based drug metabolism studies. Biochem Pharmacol 2010;79:1036–1044.
[279] Lauschke VM, Hendriks DF, Bell CC, Andersson TB, Ingelman-Sundberg M. Novel 3D culture systems for studies of human liver function and assessments of the hepatotoxicity of drugs and drug candidates. Chem Res Toxicol 2016;29:1936–1955.
[280] Bavli D, Prill S, Ezra E, Levy G, Cohen M, Vinken M, et al. Real-time monitoring of metabolic function in liver-on-chip microdevices tracks the dynamics of mitochondrial dysfunction. Proc Natl Acad Sci U S A 2016;113:E2231–E2240.
[281] Tsamandouras N, Kostrzewski T, Stokes CL, Griffith LG, Hughes DJ, Cirit M. Quantitative assessment of population variability in hepatic drug metabolism using a perfused three-dimensional human liver microphysiological system. J Pharmacol Exp Ther 2017;360:95–105.
[282] Ma C, Zhao L, Zhou EM, Xu J, Shen S, Wang J. On-chip construction of liver lobule-like microtissue and its application for adverse drug reaction assay. Anal Chem 2016;88:1719–1727.
[283] Soldatow VY, Lecluyse EL, Griffith LG, Rusyn I. In vitro models for liver toxicity testing. Toxicol Res (Camb) 2013;2:23–39.