

PAPER

View Article Online
View Journal | View Issue

Cite this: *Biomater. Sci.*, 2024, **12**, 3866

Hypothermic and cryogenic preservation of cardiac tissue-engineered constructs†

Jasmijn Janssen,^{‡a} Nino Chirico,^{‡a} Madison J. Ainsworth,^{id b} Gerardo Cedillo-Servin,^b Martina Viola,^{b,c} Inge Dokter,^a Tina Vermonden,^{id c} Pieter A. Doevendans,^{d,e} Margarida Serra,^{f,g} Ilja K. Voets,^{id i} Jos Malda,^{id b,h} Miguel Castilho,^{id j} Linda W. van Laake,^a Joost P. G. Sluijter,^a Vasco Sampaio-Pinto^{*a} and Alain van Mil^{id *a}

Cardiac tissue engineering (cTE) has already advanced towards the first clinical trials, investigating safety and feasibility of cTE construct transplantation in failing hearts. However, the lack of well-established preservation methods poses a hindrance to further scalability, commercialization, and transportation, thereby reducing their clinical implementation. In this study, hypothermic preservation (4 °C) and two methods for cryopreservation (*i.e.*, a slow and fast cooling approach to −196 °C and −150 °C, respectively) were investigated as potential solutions to extend the cTE construct implantation window. The cTE model used consisted of human induced pluripotent stem cell-derived cardiomyocytes and human cardiac fibroblasts embedded in a natural-derived hydrogel and supported by a polymeric melt electrowritten hexagonal scaffold. Constructs, composed of cardiomyocytes of different maturity, were preserved for three days, using several commercially available preservation protocols and solutions. Cardiomyocyte viability, function (beat rate and calcium handling), and metabolic activity were investigated after rewarming. Our observations show that cardiomyocytes' age did not influence post-rewarming viability, however, it influenced construct function. Hypothermic preservation with HypoThermosol® ensured cardiomyocyte viability and function. Furthermore, fast freezing outperformed slow freezing, but both viability and function were severely reduced after rewarming. In conclusion, whereas long-term preservation remains a challenge, hypothermic preservation with HypoThermosol® represents a promising solution for cTE construct short-term preservation and potential transportation, aiding in off-the-shelf availability, ultimately increasing their clinical applicability.

Received 24th November 2023,
Accepted 15th June 2024

DOI: 10.1039/d3bm01908j

rsc.li/biomaterials-science

Introduction

Currently there is no curative treatment for heart failure (HF). The rising number of HF patients (11.8% prevalence in patients above the age of 60 worldwide), associated with

improved survival rates after cardiovascular events such as a myocardial infarction (MI), and the scarcity of available transplantable hearts stresses the need for cardiac regenerative therapies such as tissue engineered constructs.^{1–6} Following a MI, the heart is unable to regenerate functional tissue, leading

^aDepartment of Cardiology, Experimental Cardiology Laboratory, Circulatory Health Research Center, Regenerative Medicine Center Utrecht, University Utrecht, University Medical Center Utrecht, Uppsalalaan 8, Utrecht, 3584 CT, The Netherlands. E-mail: A.vanMil@umcutrecht.nl, V.M.Sampaiopinto@umcutrecht.nl

^bDepartment of Orthopedics, University Medical Center Utrecht, Heidelberglaan 100, Utrecht, 3584 CX, The Netherlands

^cDepartment of Pharmaceutical Sciences, Utrecht Institute for Pharmaceutical Sciences (UIPS), Utrecht University, Universiteitsweg 99, 3508 TB Utrecht, The Netherlands

^dNetherlands Heart Institute (NLHI), Utrecht, 3511 EP, The Netherlands

^eCentraal Militair Hospitaal (CMH), Utrecht, 3584 EZ, The Netherlands

^fiBET, Instituto de Biologia Experimental e Tecnológica, Oeiras, Portugal

^gInstituto de Tecnologia Química e Biológica António Xavier, Universidade Nova de Lisboa, Oeiras, Portugal

^hDepartment of Equine Sciences, Faculty of Veterinary Sciences, Utrecht University, Yalelaan 1, Utrecht, 3584 CL, The Netherlands

ⁱLaboratory of Self-Organizing Soft Matter, Department of Chemical Engineering and Chemistry & Institute of Complex Molecular Systems, Eindhoven University of Technology, Eindhoven 5600 MBPO box 513, The Netherlands

^jDepartment of Biomedical Engineering, Eindhoven University of Technology, Eindhoven, 5612 AE, The Netherlands

†Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3bm01908j>

‡These authors contributed equally to this work and share first authorship.



to an irreversible loss of cardiomyocytes and the formation of noncontractile scar tissue. Subsequent adverse remodeling leads to organ dysfunction and eventually to advanced HF.^{7,8} The field of cardiac tissue engineering (cTE) aims to repair and restore the function of the heart post-MI by administering cardiac cells and biomaterials.⁹ Transplantation of cells embedded in biomaterials has shown to enhance cell engraftment and prevent flush-out in comparison to injections of single cells.^{10–13}

In recent years, several pre-clinical trials reported the successful implantation of cardiac sheets, leading to long-term cell retention, progressive *in vivo* maturation, and overall improvement of cardiac function.^{13,14–17} As a result, the first clinical trials are being conducted, including phase I (ClinicalTrials.gov identifier: NCT04696328) and phase I/II clinical trials (ClinicalTrials.gov identifier: NCT04396899) in 2020.^{18,19} These trials investigate the feasibility and safety of transplanting cTE constructs onto infarcted cardiac areas of patients with advanced HF.

Despite the promise of cTE, the field currently faces challenges, including a lack of maturation of the implanted cells – that promote potential arrhythmias – a needed human leukocyte antigen (HLA) match to prevent rejection of the graft, and the time required for cTE production.^{20–22} A very practical and logistic problem to date, is a lack of preservation methods that will allow transport between production sites and hospitals to facilitate cTE construct application in different environments and further scalability and commercialization of cTE constructs. These challenges are somewhat comparable to those faced by heart transplantation, which has been partially overcome by *ex vivo* organ preservation methods (up to a maximum of eight hours), ultimately increasing the number of available donor hearts by 15–30%.^{23–25} Reviewing several approaches for short- and long-term preservation has identified two main techniques that hold potential to increase cTE accessibility: hypothermic and cryogenic storage.²⁰

Hypothermic storage (subzero to 10 °C) reduces oxygen demand, metabolic activity, and protein synthesis, ultimately slowing down the energy consumption rate, thereby providing a solution for short-term storage of cells, organs, and tissues.^{26–28} To reduce harmful cell processes associated with hypothermic storage, such as ionic imbalance, formation of free radicals, and cytoskeleton damage associated with reduced ATP production, several specialized solutions have been developed, such as University of Wisconsin (UW) solution, histidine-tryptophan-ketoglutarate, Celsior, HypoThermosol® (HTS), and Rokepie (Rp). These solutions work by preventing the depletion of ATP reserves, maintaining ionic, osmotic, and oncotic (protein-induced osmotic pressure) balance, as well as reducing free radicals and providing energy substitutes.^{26,28,29}

In turn, cryopreservation stops metabolism entirely by exposing samples to ultra-low temperatures (<–150 °C), hence potentially allowing indefinite storage.³⁰ However, during the cooling and rewarming phases, intracellular ice crystals might form, thus damaging tissue and cellular integrity and viability.

During cryopreservation, extreme dehydration and intracellular ice-associated damage is prevented by replacing intracellular and extracellular water with cryoprotective agents (CPAs, *e.g.*, glycerol, dimethyl sulfoxide (DMSO)).^{31,32} In cardiac research, cryopreservation efforts have predominantly centred around preserving human induced pluripotent stem cell-derived cardiomyocytes (hiPSC-CMs), including 3D structures, highlighting the practicality and efficacy of this approach.³³

In this study, we evaluated the effect of three static storage preservation strategies, hypothermic preservation, and cryopreservation following a slow- and fast freezing approach, on a previously established cTE construct. This model consists of 90% hiPSC-CMs and 10% human fetal cardiac fibroblast (hfCFs) embedded in a gelatin and collagen-based hydrogel supported by a polycaprolactone (PCL)-based melt-electrowritten fiber (MEW) scaffold.^{34,35} Post-preservation viability, function, metabolic activity, and morphology were analysed in cTE constructs. Lastly, the effects of preservation on the hydrogel and MEW scaffold physical properties were evaluated (hydrogel swelling behavior, elasticity, surface topography, and crystallinity).

Experimental section

Cell culture and differentiation

All experiments were conducted according to the criteria of the code of proper use of human tissue used in the Netherlands and approved by the ethics committees of the University Medical Center Utrecht and Leiden University Medical Center, the Netherlands. Different hiPSCs lines, all deposited in the European Bank for iPSCs (EBiSC, <https://ebisc.org/>) and registered in the online registry for human PSC lines hPSCreg (<https://hpscereg.eu/>), were used: UKKi032-C (NP0141-31B) (female) (Fig. S1†), UKKi037-C (NP0144-41) (female), and NP0143-18 UKKi036-C (male).³⁶ hiPSCs were differentiated into hiPSC-CMs using a GiWi differentiation protocol.³⁷ In short, at day 0, with hiPSCs at 90% confluency, medium was changed to heparin medium (see Table S1† for media compositions) with 4 μM CHIR99021 (Selleck Chemicals). After 48 hours, medium was refreshed with heparin medium with 2 μM Wnt-C59 (Tocris Bioscience). At day 4 and 6, medium was changed with heparin medium. From day 7 onwards, medium was replaced with insulin medium (Table S1†) until purification around day 10. Around day 10, the hiPSC-CMs were beating, and medium was changed with purification medium (Table S1†) every other day until day 15. From day 15 onwards, hiPSC-CMs were expanded following Wnt activation by GSK-3β inhibition with 2.0 to 4.0 μM CHIR99021 and the inhibition of cell-cell contact.^{38,39} Human fetal cardiac fibroblasts (hfCFs) (P6-10) were isolated and cultured as described previously.⁴⁰

Construct preparation

Melt electrowriting (MEW) scaffold fabrication. MEW micro-fiber scaffolds were fabricated as previously described.^{34,35,41}



Briefly, hexagonal MEW scaffolds with a side length of 400 μm were fabricated using a 3DDiscovery (regenHU) and medical-grade PCL granules (Purasorb, Corbion). Constructs with an 8 mm diameter and $\sim 400\ \mu\text{m}$ in thickness comprised 20 layers with a fiber diameter of 11 μm . For tensile analysis $5 \times 10\ \text{mm}$ constructs were prepared.

Construct seeding. The cell suspension was resuspended in a blend of 5% w/v porcine skin gelatin methacryloyl (GelMA, Sigma-Aldrich, G2500) (80% degree of functionalization) and 0.8 mg mL^{-1} type 1 rat tail collagen (RatCol®) supplemented with 0.5 mM tris (2,2'-bipyridyl)dichlororuthenium(II) hexahydrate (Ru, Sigma-Aldrich, 544981) and 5 mM sodium persulfate (SPS, Sigma-Aldrich, S6172) as photo-initiators⁴² and casted within a hexagonal PCL scaffold (thickness $\sim 400\ \mu\text{m}$, diameter = 8 mm).³⁴ Crosslinking was initiated with white light (floodlight) for 4 minutes. Subsequently, the constructs were incubated at 37 °C for collagen thermal crosslinking for 12 minutes.³⁵ 75% of the construct culture media (Table S1†) was changed every other day.

Preservation methods

Hypothermic preservation was conducted on a slowly moving (approx. 15 rpm) rocking platform (Polymax 1040, Heidolph Instruments) at 4 °C for 3 days. Rokepie-FD01® (Rp), HypoThermosol® (HTS), and CoStorSol®/University of Wisconsin (UW) were utilized according to manufacturer instructions (Table S2†). Plates were sealed air-tight with parafilm, preventing a drop in CO_2 to cause an increase in the solution's pH.^{43,44} Standard cryopreservation following a slow freezing protocol was initiated by transferring the construct into a cryovial (Cryo.S™) with 1 mL pre-cooled standard freezing media (4 °C) (Table S2†) and cooled and stored for one day in a CoolCell container (Biocision) at $-80\ ^\circ\text{C}$ with a cooling rate of $-1\ ^\circ\text{C min}^{-1}$. After 24 hours, the vial was transferred to a liquid nitrogen freezer ($-196\ ^\circ\text{C}$), where it stayed for an additional 2 days (3 days of cryogenic preservation in total). Fast freezing of cTE constructs was conducted in aluminium cryotubes (Sanbio). Constructs were incubated in 1 mL pre-cooled (4 °C) standard freezing media (Table S2†) for 30 minutes to allow DMSO permeation.^{45,46} Following incubation, the cryotube was carefully dipped in a $-80\ ^\circ\text{C}$ isopropanol bath for 2.5 minutes. During these 2.5 minutes, the constructs reached an average temperature of $-60\ ^\circ\text{C}$, with freezing rates ranging from $>150\ ^\circ\text{C min}^{-1}$ during the first seconds to $11.4\ ^\circ\text{C min}^{-1}$ from $-50\ ^\circ\text{C}$ to $-60\ ^\circ\text{C}$ as measured with a temperature logging digital thermometer (Fluke, 53 IIB) (Fig. S2†). Thereafter, cryotubes were transferred to a $-150\ ^\circ\text{C}$ freezer, where they were preserved for 3 days. Post-preservation, conventional thawing in a 37 °C water bath was performed as recommended for samples $<2\text{--}3\ \text{mL}$.³¹ Briefly, samples were submerged in the water bath until only a sliver of ice remained (15 s) and constructs were transferred to a 24-well plate filled with 2 mL pre-warmed construct media. Media was refreshed one hour and one-day post-thawing, followed by media changes every other day.

Method of time matching of controls and preserved constructs

For each experiment, cTE constructs were prepared and cultured for 7 days. Following this first week, constructs were randomly allocated to one of the experimental groups: hypothermic preservation for 3 days, cryogenic preservation for 3 days or control culture conditions for 1 day. Due to the low temperatures achieved during preservation, construct metabolism and maturation are stopped for 3 days. After 3 days of preservation, the constructs were rewarmed by transferring them to pre-warmed 37 °C fresh construct medium. After a recovery period of 24 hours, preserved constructs were analysed and compared to their time-matched controls (CTRL) (Fig. 1). Analysis time-points are denoted as D7 + X, representing the age of the constructs (7 days of culturing + X days of culture under standard culture conditions after allocation to their experimental conditions). For instance, D7 + 1 control constructs were cultured continuously for 8 days, while D7 + 1 preserved constructs were cultured for 7 days, preserved for 3 days (during which time stopped), and cultured for one more day to allow for cTE recovery. This time-matching strategy ensured that both controls and experimental groups (hypothermic or cryogenic preservation) were compared after spending an equal time in culture at 37 °C, when cTE metabolism and functionality is highest.

Evaluation of cell viability – LIVE/DEAD analysis

Cellular viability was determined with the LIVE/DEAD viability/cytotoxicity kit (ThermoFisher Scientific, L3224) following manufacturer instructions. The stained constructs were visualized and captured in three different locations – once in the center and twice at the construct periphery – *via* z-stack imaging with Leica Thunder microscopy (z-stack size = 102.94 μm) (Fig. S3†). Maximum projections were obtained from the equal-sized z-stacks and cell viability was quantified by determining the area of red and green fluorescence (dead/live cells, respectively) on the maximum projection images using the ImageJ software. Data were normalized to time-matched CTRL.

Beating rate

Twenty-second videos were taken using a GoPro Black Hero 7 camera connected to a bright field microscope (Olympus CKX41) *via* a c-mount system (Olympus U-TV1X-2). During video acquisition, cTE constructs were kept at 37 °C using a warm base plate.

Cellular metabolic activity, metabolic substrate quantification, and high-sensitivity troponin assay

Cellular metabolic activity was evaluated with the alamarBlue cell viability assay (Thermo Fisher Scientific, DAL1025) following manufacturer instructions. In short, 10% alamarBlue solution in construct medium was added to the constructs. After 4 hours, the reacted solution was transferred to a 96-well plate and fluorescence was recorded using a fluorescence excitation wavelength of 560 nm and an emission wavelength of 590 nm.



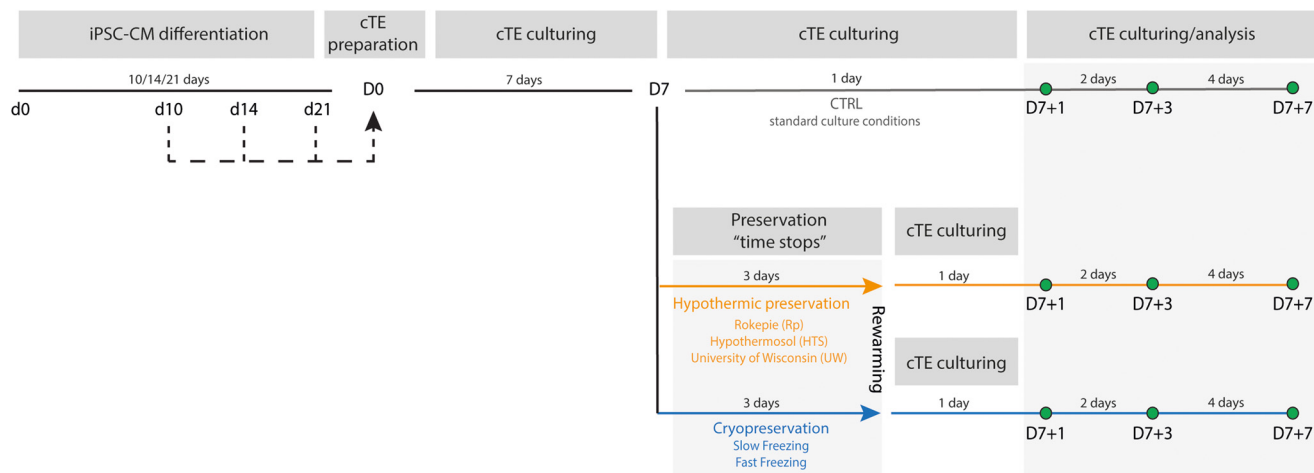


Fig. 1 Experimental timeline. Visual representation of the incorporation of hiPSC-CMs with different ages (d10, d14 and d21, black dotted lines) into the cTE constructs and time-matching of the controls and preserved constructs. After seven days of cTE construct culturing, the constructs were divided into three groups, subjected to the different experimental conditions. Control constructs (grey line) were kept under standard cTE culture conditions during the experiments. Preserved constructs were subjected to 3 days of hypothermic- (orange line) or cryogenic (blue line) conditions. Constructs were analyzed at three different timepoints: D7 + 1, D7 + 3 and D7 + 7.

Absorbance was monitored at 570 nm and presented in arbitrary fluorescent units.

Conditioned culture media was collected at different time points (*i.e.* before preservation, at D7 + 1, D7 + 3, and D7 + 7 post-preservation) and analyzed using an Atellica CH Analyzer (Siemens). Glucose (Atellica CH Glucose Hexokinase₃; REF 11097592), lactate (Atellica CH Lactate₂; REF 11532568), and cardiac troponin-I levels (Atellica IM High-Sensitivity Troponin I; REF 10997840) were measured. To prevent media change-associated differences between different timepoints, media was always refreshed one day prior to media collection for analysis.

Calcium handling

Three days post-preservation (D7 + 3), when preserved cTE constructs re-start contracting (as individual hiPSC-CMs or as small or larger clusters), calcium handling was evaluated. cTE constructs were incubated with construct culture media with 25% fluorobright DMEM (ThermoFisher Scientific, A1896701) supplemented with 1:1000 Cal-520® AM (Abcam, ab171868) and 8% Pluronic F-127 (Sigma-Aldrich) for 45 minutes at 37 °C. Thirty second videos were taken at three different locations – once near the centre and twice at the periphery of the construct – with a Leica Thunder microscope (Fig. S3†) and as previously described.³³ Data analysis was performed using FIJI and Peaks, a custom-written Matlab script (<https://doi.org/10.17605/OSF.IO/86UFE>).

Evaluation of post-preservation hydrogel properties

The effects of preservation on the hydrogel were determined three days post-preservation (D7 + 3) *via* a sol-fraction analysis, percentage of macromer, and the mass swelling ratio. *Hydrogel sol-fraction analysis*: sol-fraction was performed using the pro-

cedure from Blocki *et al.*⁴⁷ Percentage of macromer, mass swelling ratio (q), and the sol-fraction could be determined following the equations below:

$$\% \text{macromer} = \frac{m_{\text{dry}, t_0}}{m_{\text{wet}, t_0}} \times 100\% \quad (1)$$

$$\text{Mass swelling ratio } (q) = \frac{m_{\text{swollen}, t_1}}{m_{\text{dry}, t_1}} \quad (2)$$

$$\text{Sol fraction} = \frac{m_{\text{dry}, t_0} - m_{\text{dry}, t_1}}{m_{\text{dry}, t_0}} \times 100\% \quad (3)$$

Evaluation of post-preservation PCL scaffold properties

Preserved PCL scaffolds were analyzed three days post-preservation (D7 + 3) by scanning electron microscopy (SEM), differential scanning calorimetry (DSC), and tensile stress tests and compared to non-preserved controls. SEM pictures were taken of all groups ($n = 3$), using a backscatter detector with 10 kV acceleration voltage after pre-coating the constructs with a 2 nm gold coating to enhance conductivity. To assess thermophysical properties, CTRL, HTS and fast frozen constructs were analyzed using DSC Q2000 equipment (TA instruments). A cycle of scans (heating-cooling-heating) was performed on the PCL scaffolds (5 mg, loaded into Tzero aluminium pans (TA Instruments)) from 0 to 150 °C with a heating rate of 10 °C min⁻¹ and a cooling rate of 1 °C min⁻¹, under nitrogen flow of 50 mL min⁻¹. Melting temperature (T_m) (°C), enthalpy (J g⁻¹), and crystallinity were determined for both groups of preserved constructs and compared to controls. Melting temperatures (T_m) were determined from the onset of the endothermic peaks. Lastly, uniaxial tensile tests were performed with a 25 N force gauge in a Multitest 2.5-dV mechanical tester (Mecmesin), and data was recorded using VectorPro software (Mecmesin).



Statistical analysis

All statistical analyses were performed in GraphPad Prism v9.4.1 (GraphPad Software Inc., La Jolla, USA) and data expressed as mean $\pm$ standard deviation. Ordinary one way or two-way analysis of variance (ANOVA) with Tukey's multiple comparisons were carried out for all analyses. For the analysis of the calcium transients, a multiple comparison Kruskal–Wallis test was performed. For all analyses, a *p*-value less than 0.05 was considered to be significant.

Results

In this study, long- and short-term preservation techniques and solutions were tested on cTE constructs.^{34,35} The cTE construct represents a complex system with multiple cell types and biomaterials, where cell viability and function are linked to hydrogel and scaffold integrity.^{34,35} This relationship needs to be maintained regardless of the preservation strategy; thus, a wide set of parameters are investigated, at both the cellular and material level.

HypoThermosol® and fast freezing best preserved cTE constructs viability

To determine the impact on and feasibility of preservation of cTE constructs, three commercially available hypothermic preservation solutions were tested: ROKEPIE (Rp), University of Wisconsin solution (UW), and HypoThermosol® (HTS). Additionally, two cryopreservation methods were applied: slow- and fast freezing (Fig. 1).

Seven-day-old cTE constructs were preserved for three days in hypothermic solution (at 4 °C) and subsequently rewarmed and cultured for seven days (Fig. 1 – orange line). Viability analyses at D7 + 3 showed that HTS exhibited a trend of higher cell viability ($102.9\% \pm 20\%$) compared to other solutions (Rp: $84.0\% \pm 4.9\%$, and UW: $95.2\% \pm 2.0\%$) and the time-matched unpreserved constructs (CTRL: $100\% \pm 5.4\%$) (Fig. 2A and B). After rewarming, cTE constructs preserved in the presence of Rp and UW did not resume contractions, while those preserved in HTS showed no signs of functional decline, with a synchronized beating rate comparable to the unpreserved CTRL (Fig. 2C) (Videos S1 and S2†).

Slow- and fast freezing resulted in a significant decrease in construct viability and functional impairment at D7 + 3. Slow freezing resulted in $48.7\% \pm 12.5\%$ survival while fast freezing ensued $73.4\% \pm 15.7\%$ viability after rewarming (Fig. 2D and E). Of note, the beating rate showed a non-significant decrease in cryopreserved constructs compared to CTRL (Fig. 2E and Videos S1, S3, S4†). However, preserved constructs displayed patchy, unsynchronized beating clusters. This effect was even more pronounced in the slowly frozen constructs (Video S3†) that had fewer beating clusters compared to the fast frozen group. Furthermore, after a longer, 7-days of culture after rewarming, no beating clusters could be observed in the slowly frozen constructs, while beating clusters could still be distinguished in fast frozen constructs.

Overall, these results indicate that HTS-based hypothermic preservation and fast freezing were most effective at maintaining cTE construct viability and contractility. For this reason, these conditions were selected for subsequent experiments.

Cardiomyocyte maturation does not influence the viability of cTE constructs

The differentiation day or maturity of cardiomyocytes has been shown to influence their post-preservation viability and functionality in a 2D setting, emphasizing the possibility of an effect of cardiomyocyte age in the context of cTE construct manufacturing and recovery yields after preservation.^{48–50} We investigated the influence of cardiomyocyte maturation on post-preservation viability and beating behaviour following the most promising preservation approaches: HTS and fast freezing.

In this study, we evaluated hiPSC-CM age-dependent recovery by preparing cTE constructs with hiPSC-CMs at differentiation day 10 (d10, initial beating), day 14 (d14, replating day), and day 21 (Fig. 1, black dashed lines). Time-matched controls (CTRL) from the same construct batch were kept in culture for comparison (Fig. 1, grey line). Post-preservation, viability was determined at D7 + 1 and D7 + 3, one and three days after rewarming, respectively (Fig. 1).

Live/dead analysis showed no significant differences between CTRL and preserved samples, irrespective of the analyzed timepoint and hiPSC-CMs age (Fig. 3A and B). However, cardiomyocyte age greatly influenced cell organization and the contractile phenotype of cTE constructs. Constructs prepared with d21 hiPSC-CMs displayed synchronous beating of the whole construct before preservation (D7) (Video S7†). In contrast, pre-preservation constructs prepared with d10 or d14 hiPSC-CMs often showed no or cluster-like contractions and different tissue morphology (Videos S5, S6,† and Fig. 3A). At D7 + 3, d10 hiPSC-CM constructs showed no signs of contraction, d14 hiPSC-CM constructs showed irregular beating behavior or no contraction (Videos S8–10†), and d21 hiPSC-CM constructs showed more consistent beating behavior across all experimental groups (Videos S11–S13†).

Altogether, and in line with previous observations, d21 hiPSC-CMs have a greater ability to re-arrange the supporting hydrogel and connect with each other before preservation, and do not show lower viability compared to younger cells, establishing the basis for subsequent experiments using d21 hiPSC-CMs.³⁵

HypoThermosol® and fast freezing reduced cTE construct metabolism and recovery post-preservation

The impact of HTS and fast freezing on the metabolic activity of cTE constructs were investigated by measuring cellular redox activity, glucose consumption, and lactate production. cTE constructs subjected to fast freezing displayed an acute reduction of metabolic activity ($12.82\% \pm 1.33\%$) at D7 + 1, compared to CTRL. Fast frozen constructs' metabolic activity



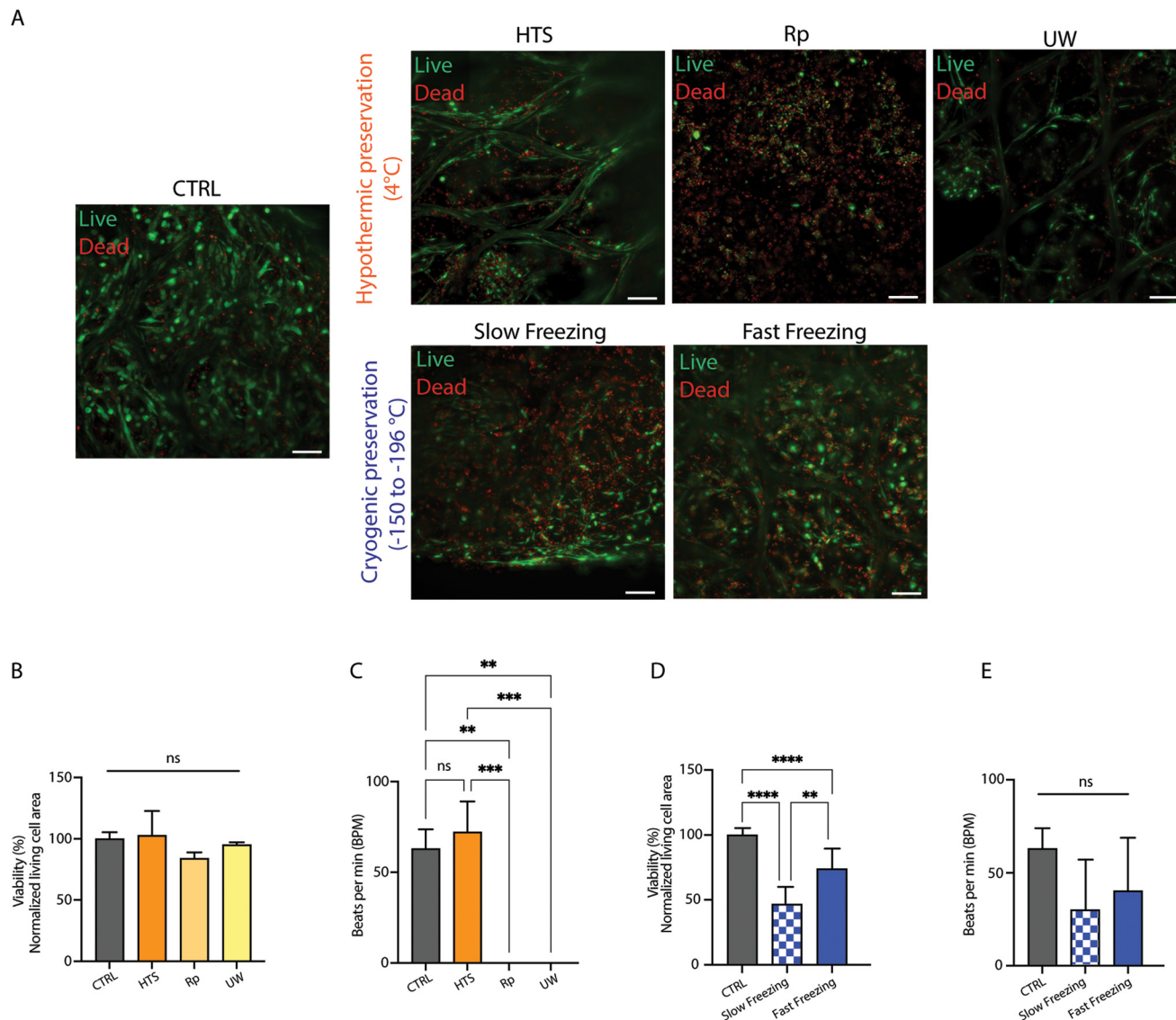


Fig. 2 HypoThermosol® and fast freezing best preserved short- and long-term cTE construct viability. (A) Live/dead imaging 3 days after rewarming (D7 + 3). Living cells, (green/calcein-AM positive), and dead cells (red/ethidium homodimer-1 positive). Channels are overlaid. Scale bar = 150 μ m. Representative images are shown. (B) Live/dead quantification, normalized to time-matched unpreserved CTRL. CTRL = 100% $\pm$ 5.4%, HTS = 102.9% $\pm$ 20%, Rp = 84.0% $\pm$ 4.9%, and UW = 95.2% $\pm$ 2.0% (median age: d14, n = 14, 13, 2 and 2, respectively). (C) Beating rate of cTE constructs preserved under hypothermic conditions. HTS showed signs of functional decline (Video S2†) while no contraction was detected in the Rp and UW groups. (D) Live/dead quantification, normalized to time-matched unpreserved CTRL. CTRL = 100% $\pm$ 5.0%, Slow freezing = 48.7% $\pm$ 12.5%, and fast freezing = 73.4% $\pm$ 15.7% (median age: d21, n = 16, 4, and 16, respectively). (E) Beating rate of cTE constructs subjected to slow- and fast freezing (Videos S3 and S4†). A non-significant decrease was observed in preserved constructs compared to time-matched unpreserved CTRL.

gradually increased over time to 43.44% $\pm$ 13.04% on D7 + 3 and 94.98% $\pm$ 28.18% on D7 + 7. After hypothermic preservation, the metabolic activity of HTS-preserved constructs initially dropped to 61.26% $\pm$ 17.13% on D7 + 1 but was no longer distinguishable from that of CTRL constructs from D7+3 onwards (Fig. 4A).

The iPSC-CMs and hfCFs embedded in the cTE constructs consume metabolic substrates like glucose from the medium, while lactate is released as a by-product of glucose consumption.⁵¹ Before preservation (D7), comparable glucose consumption (3.03 mM to 3.63 mM) and lactate production (6.37 mM

to 7.43 mM) were observed among experimental groups. After preservation and rewarming, on D7 + 1 and D7 + 3, fast frozen constructs presented no appreciable glucose consumption or lactate production. Limited recovery was subsequently observed at D7 + 7 with glucose consumption increasing to 1.08 mM $\pm$ 0.30 mM and lactate production to 4.13 mM $\pm$ 0.84 mM. On the contrary, HTS constructs showed signs of recovery at D7 + 3, for both glucose and lactate, but failed to reach the levels measured in CTRL (Fig. 4B and C). These results indicate that the metabolic activity of cTE constructs is strongly inhibited after preservation, particularly in the fast

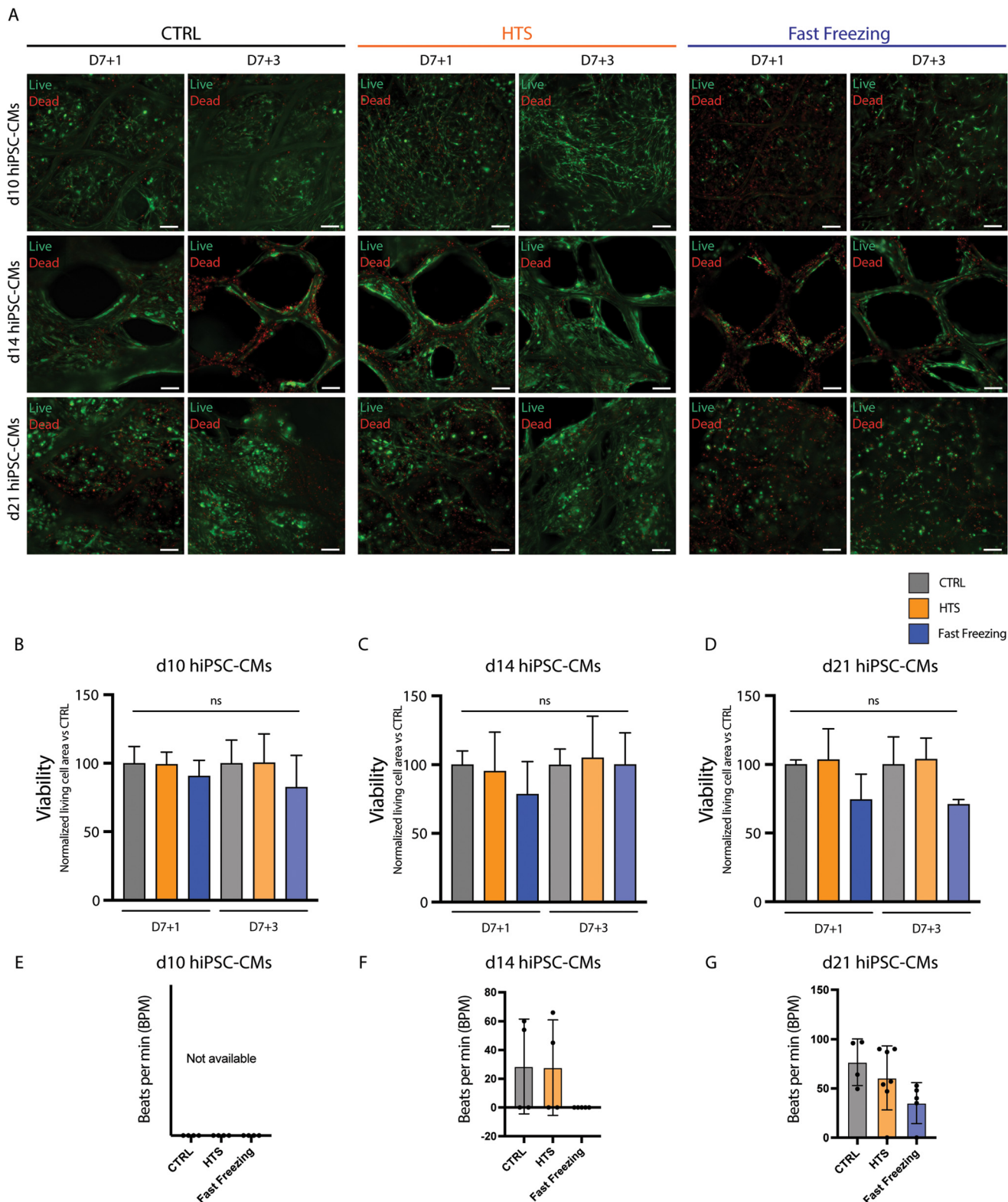


Fig. 3 Cardiomyocyte age does not influence viability of cTE constructs. (A) Live/dead imaging at 1 (D7 + 1) and 3 (D7 + 3) days after rewarming. Living cells, (green/calcein-AM positive), and dead cells (red/ethidium homodimer-1 positive). Channels are overlaid. Scale bar = 150 μ m. Representative images are shown. (B–D) Live/dead quantification, normalized to time-matched unpreserved CTRL for d10, d14 and d21 hiPSC-CMs ($n = 4$). (E–G) Beating rate of cTE constructs with d10, d14, and d21 hiPSC-CMs at D7 + 3 (Videos S8–S13[†]). The incorporation of d10 iPSC-CMs did not yield contractile cTE constructs. No significant differences were observed between preserved constructs and CTRL.



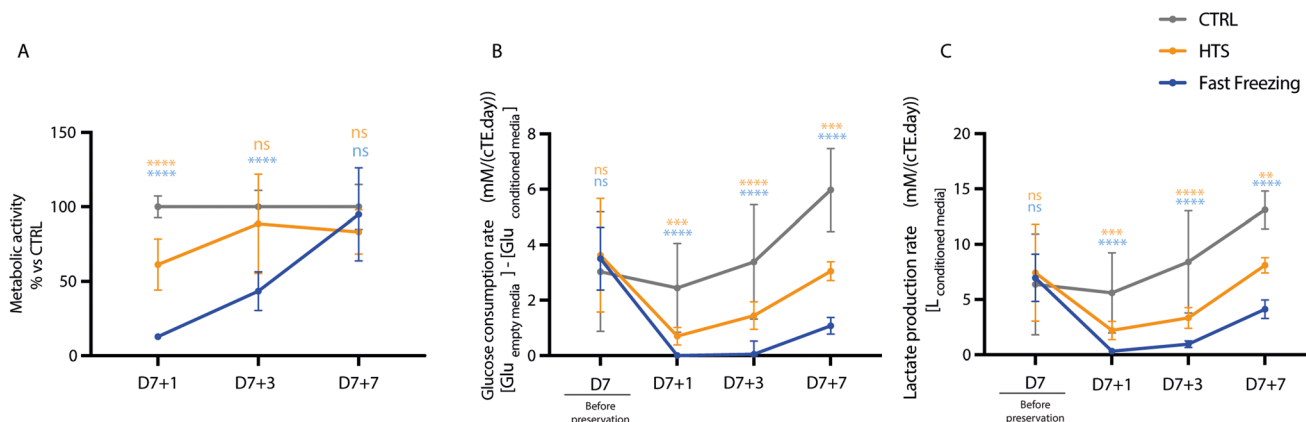


Fig. 4 Hypothermosol® and fast freezing reduced cTE construct metabolism and recovery post-preservation. (A) Redox activity of preserved constructs determined by the Alamar Blue assay at days 1 (D7 + 1), 3 (D7 + 3) and 7 (D7 + 7) after rewarming; HTS-preserved ($n = 7$), fast frozen ($n = 7$) CTRL ($n = 8$). (B) Glucose consumption rate of hypothermically preserved (HTS, $n = 20$) and fast frozen ($n = 18$) constructs, was compared to that of unpreserved controls (CTRL, $n = 16$) at days 1 (D7 + 1), 3 (D7 + 3) and 7 (D7 + 7) after rewarming. (C) Lactate production of HTS-preserved ($n = 20$), fast frozen ($n = 18$) and CTRL ($n = 16$) constructs was compared pre-preservation and at days 1 (D7 + 1), 3 (D7 + 3) and 7 (D7 + 7) after rewarming. ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, and ns indicates a non-significant difference. The color indicates whether the comparison was made between CTRL vs. HTS (orange) or CTRL vs. fast freezing (blue).

frozen group, though partial recovery occurs during culture after rewarming in the HTS group.

The impact of HTS and fast freezing on hiPSC-CMs sarcomere organization and calcium handling

The damage imposed by cryo- and hypothermic preservation resembles reperfusion post-MI, including a loss of sarcomere content, arrangement, and functionality, resulting in disrupted contractility and calcium handling, both critical for cTE construct performance.^{25,48,52} Following fast freezing, D7 + 3 cTE constructs showed a strong reduction in troponin-T expressing cells compared to CTRL and HTS, which showed a comparable amount (Fig. 5A). Additionally, at D7 + 1 troponin-I release to the medium was substantially higher in fast frozen constructs ($4428.1 \text{ ng mL}^{-1} \pm 2925.59 \text{ ng mL}^{-1}$), and to a lesser extent in HTS cTE constructs ($1576.47 \text{ ng mL}^{-1} \pm 1267.42 \text{ ng mL}^{-1}$), compared to CTRL ($91.33 \text{ ng mL}^{-1} \pm 26.73 \text{ ng mL}^{-1}$). Follow-up measurements (D7 + 3 and D7 + 7) showed no differences between HTS, fast freezing or CTRL (Fig. 5B). Calcium handling at D7 + 3 showed a decreased CaT upstroke velocity for both HTS and fast frozen samples (Fig. 5C–E), with fast freezing displaying a slight decrease in CaT amplitude (Fig. 5D) and no propagation between cell clusters, clearly highlighting the damage in the fast frozen samples (Videos S14–S16†).

Effect of preservation on the mechanical properties, surface topography and crystallinity of cTE constructs

Upon implantation, cTE constructs need to withstand the continuous contractions and relaxation of the myocardium. However, as recently reported, rapid cooling/warming of polymers, particularly filaments, can lead to changes in mechanical properties due to alterations in crystallinity.^{20,53} First, the effect of hypothermic preservation and fast freezing on the

cell-supporting hydrogel was investigated. The results reveal no significant differences in the macromer percentage, mass swelling ratio, and sol fraction (Fig. 6A–C).

Additionally, fast freezing may shrink the polymer filaments leading to cell detachment and, ultimately, apoptosis.^{20,53} Concurring with this, live/dead imaging of fast frozen cTE constructs revealed the presence of several dead cells particularly in the vicinity of PCL fibers (Fig. S4†). SEM was used to closely observe the topography of the PCL mesh, but no lasting impact on the rewarmed PCL fibers was observed for either of the two preservation methods (Fig. 7A). Further investigation into PCL network remodeling after preservation focused on its melting curves and crystallinity. DSC measurements reveal how the first heating cycle, which may be influenced by the thermal history of the polymer, presented readings of melting temperature and melting enthalpy that were not statistically different between CTRL, HTS and fast frozen constructs (Fig. 7B). Similarly, the crystallization temperature and enthalpy were essentially the same (Fig. 7C). Comparably, similar trends in the change of enthalpy and melting temperature between the first and second heat cycles were observed in all the groups (Fig. 7D). Finally, analysis of bulk mechanical properties of the PCL scaffold through uniaxial tensile testing showed no significant differences in the elastic stress limit and the elastic strain energy density (Fig. 7E).

Discussion

Despite numerous advances in cTE, several hurdles limit the application of cell-based cTE constructs in the clinical arena. Recent advances in HLA-compatible hiPSC lines would allow the production of cTE constructs upfront, thus making them readily available for biobanking, and implantation as an off-



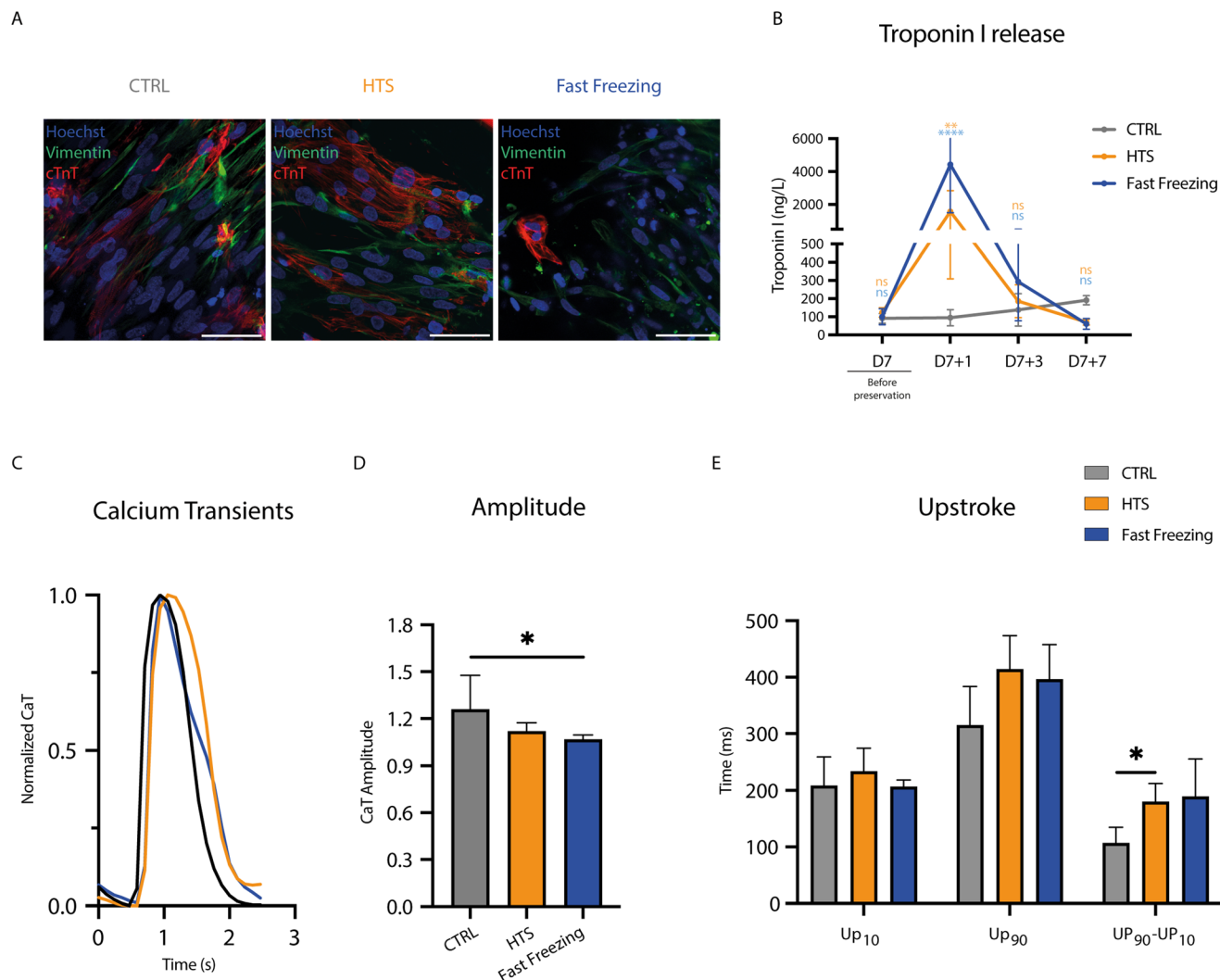


Fig. 5 The impact of HTS and fast freezing on hiPSC-CM sarcomere organization and calcium handling. (A) Representative immunofluorescence images taken at D7 + 3. CTRL and HTS showed comparable presence of well-arranged and dense sarcomere protein expression (cardiac troponin T (cTnT)). Fast frozen constructs showed reduced cTnT expressing cells. Scale bars = 200 μ m. (B) Troponin-I release to the medium ($n = 3$ to 13). (C) Representative normalized calcium transients of constructs at D7 + 3. (D) Calcium transient amplitude at D7 + 3. (E) Calcium transient upstroke kinetics at D7 + 3.

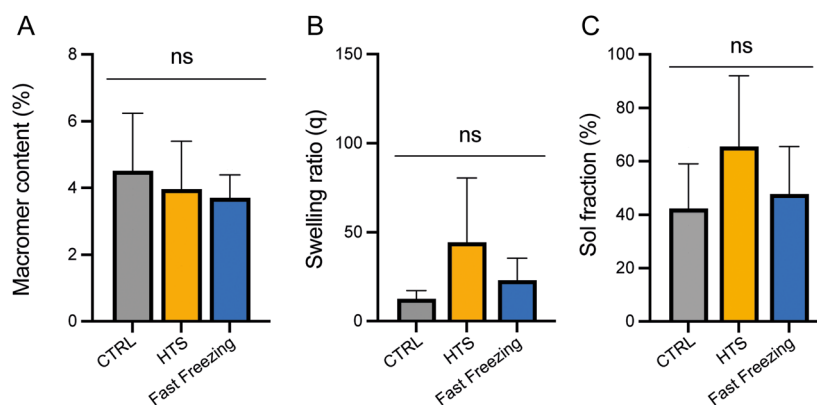


Fig. 6 The effect of hypothermic preservation and fast freezing on the cell-supporting hydrogel at D7 + 3. (A) Macromer percentage, (B) mass swelling ratio, and (C) sol fraction. $n > 4$.



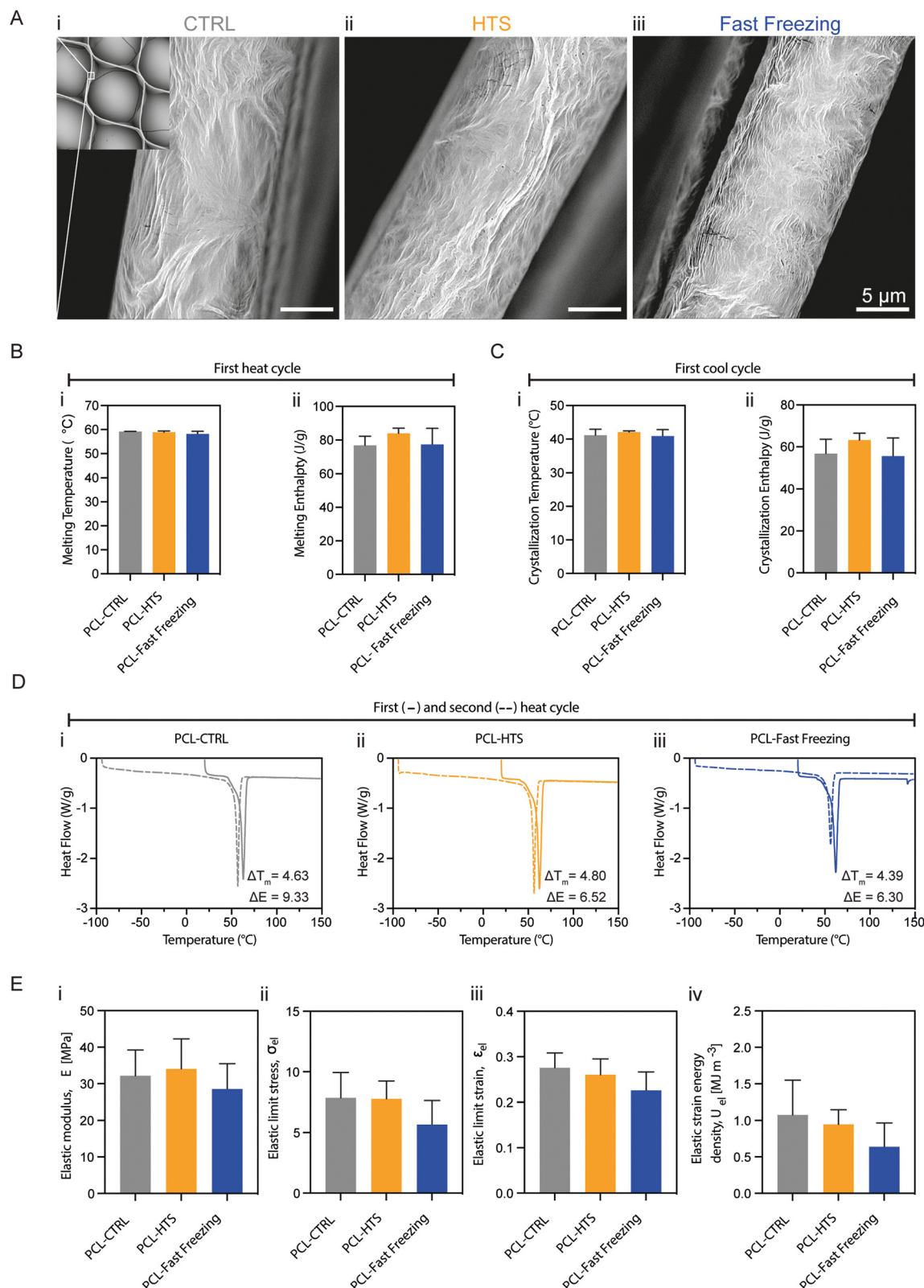


Fig. 7 No significant changes to scaffold mechanical properties, surface topography, and crystallinity of cTE constructs. (A) Scanning electron microscopy images of scaffold fibers at D7 + 3. Top-left corner a wide overview of the scaffold. Representative images are shown. $n = 4$. (B–D) Differential scanning calorimetry analysis of samples. (B) Melting temperature and enthalpy from first heating ramp. Data shown as mean with standard deviation as error bars. $n > 3$. (C) Crystallization temperature and enthalpy from first cooling ramp. Data shown as mean with standard deviation as error bars. $n > 3$. (D) First and second heating ramps plotted per condition with calculated mean change of melting temperature and enthalpy. $n > 3$. (E) Tensile mechanical analysis of samples with calculated elastic modulus, elastic limit stress, elastic limit strain and elastic strain energy density. Individual data points plotted with mean and standard deviation as error bars. $n = 6$.



the-shelf treatment for acute patients.⁵⁴ Thus, efficient preservation and off-the-shelf availability has the potential to revolutionize heart failure treatment by extending the time between production and implantation. Yet, effective preservation methods for cTE constructs are currently lacking, ultimately hindering their scalability, commercialization, and transport. Finding solutions to this challenge will increase the accessibility of tissue engineered approaches and their potential benefits to patients suffering from several diseases not only limited to the heart.²⁰

This study aimed to identify the optimal method for preserving a complex cTE construct. This construct consisted of hiPSC-CMs and hfCFs, seeded in an ECM-mimicking hydrogel composed of GelMA-collagen,⁵⁵ and embedded in a PCL-based MEW microfiber scaffold with hexagonal micro-architecture to promote cellular organization and improved contraction.^{34,35,56} Hence, the cTE constructs used herein represent a large group of cardiac constructs currently under development as disease models or clinical therapy.

The goal of short-term preservation, at hypothermic temperatures, is to maintain construct viability for a few days (three days in this study) to allow for safe transportation. By transporting tissue under hypothermic conditions, one avoids the build-up of gas and stress factors, the depletion of nutrients and growth factors, and the pH changes associated with the secretion of lactic acid and other waste products after >48 hours of transport at 37 °C.⁵⁷ For this application, HTS yielded superior post-preservation recovery among the three commercially available solutions tested. After rewarming, HTS-preserved constructs showed less functional decline, with a beating rate comparable to that of unpreserved CTRL, whereas Rp and UW did not recover contractility after rewarming (Fig. 2C). These results are analogous with previous studies where hiPSC-CMs preserved in HTS as monolayers or aggregates showed a metabolic activity ranging from 60 to 70% after 7 days of storage.

HTS outperforming Rp and UW could be linked to its broader range of action compared to RP and UW.^{58,59} HTS acts as a PH buffer, ROS scavenger, energy substrate, and ion, osmotic, and oncotic balancer. Whereas UW only ensures osmolarity and reduced oncotic damage by scavenging free radicals, and Rp increases mitochondrial respiration, thus counteracting slower metabolism.

The goal of long-term preservation at cryogenic temperatures is to maintain construct viability for a hypothetical indefinite span of time. This would ensure the biobanking and effective off-the-shelf availability of cTE constructs. In our model, fast freezing outperformed slow freezing. However, both methods critically impaired constructs viability and functionality. These findings are in line with previous experiments with cardiac aggregates, small tissues, and whole hearts.^{60,61} The superiority of fast freezing compared to slow freezing could be explained by the faster cooling rates partly protecting the constructs from irremediable damage caused by cell dehydration during the cooling phase. During slow freezing, osmotic pressure forces water to exit from the cells. If cooling

rates are fast enough, water efflux cannot occur before freezing. Consequently, the water becomes trapped within the cells, limiting damage caused by cell dehydration and shrinkage.^{31,32,62}

Interestingly, hiPSC-CM maturation status did not have a significant effect on cTE construct viability post-preservation for both HTS or fast freezing (Fig. 3A–D). This contrasts with previously published research on 2D cultures of hESC-CMs, reporting better results after freezing hESC-CMs pre-beating stage, compared to post-beating stage.^{28,48,49} However, we observed that incorporating more mature hiPSC-CMs resulted in synchronous beating, extending throughout the whole scaffold before preservation (D7) and showed improved cell and matrix re-arrangement (Fig. 3A), as reported in previous publications.³⁵ Moreover, storing more mature constructs led to improved preservation of beating characteristics (Fig. 3E and F). In light of these findings, the decision was made to proceed with the preservation of constructs made with d21 hiPSC-CMs.

Characterizing the impact of preservation on viability in complex 3D structures is often challenged by the inability of assessing the full depth of the sample, potentially leading to selection bias. Herein, LIVE/DEAD quantifications were conducted in large z-stacks, that were maintained constant across different experiments and experimental groups, thus providing a good and unbiased overview of cTE viability throughout the construct. In addition, beating rate measurements, used to characterize cTE constructs functionally, are often not sufficient and can yield misleading results. In fact, similar beating rates were observed in preserved and unpreserved constructs, but the underlying dynamics varied significantly, as shown in ESI Videos 1–13.† While CTRL constructs show often a cohesive, synchronized rhythm throughout the whole construct, HTS and cryopreserved constructs mostly display clustered- or single cell beating, respectively. Therefore, the inclusion of representative videos alongside beating rate data is indispensable for providing a comprehensive understanding of the complex dynamics underlying cTE constructs.

Recent studies have described the damage imposed by cryo- and hypothermic preservation to be similar to that caused by reperfusion after an MI, including oxidative stress, damage-associated and antioxidant gene expression, impaired action potential propagation, reduced cell–cell contact, and loss of sarcomere content and organization.^{25,48,52,63,64} Altogether, these findings, in addition to the technical limitations associated with viability and contractility assays in constructs, indicate that the assessment of preservation-associated damage necessitates a multiparameter approach that focuses on construct recovery beyond the mere live and dead and beating rate data.

Indeed, complementing our viability and contractility data with metabolite analyses revealed that the metabolism of HTS-preserved cTE constructs was initially decreased by 40%, but fully recovered at D7 + 3 (Fig. 4A). This was also corroborated by a small increase in glucose consumption and lactate production between D7 + 1 and D7 + 3 (Fig. 4B and C).



Conversely, fast frozen samples exhibited an initial sharp reduction (almost 80%) in redox activity (Fig. 4A) combined with negligible glucose consumption and lactate production shortly after rewarming with only a partial recovery at D7 + 7 (Fig. 4B and C). Of note, glucose is also utilized by hfCFs as a substrate. Considering the lack of beating cells, these data suggest that the increased consumption of glucose for fast frozen constructs may be linked to hfCFs proliferation. In the clinic, enhanced lactate levels in the blood are typically taken as a marker for acute heart damage. However, in this case, lactate is released as a by-product of glucose consumption, a finding also observed in the CTRL constructs.^{65–67} An initial decrease in metabolism and its subsequent improvement might suggest that allowing a longer recovery time would result in total functional recovery. Glucose, as well as other substrates are consumed by the hiPSC-CMs, mainly to provide energy for contraction.⁶⁸ Troponin (T and I), one of the main components of the sarcomeres, is released upon cell damage and is commonly used in the clinic as a biomarker for MI and other forms of cardiac injury.⁶⁹ Immunofluorescence revealed that HTS preserved- and, to a larger extent fast frozen constructs, had an appreciable reduction in the number of troponin-T expressing cells (Fig. 5A). In addition, quantification of troponin-I in the conditioned media of cTE constructs revealed a 44-fold and 15-fold increase for fast frozen- and HTS samples, respectively, compared to controls at D7 + 1 (Fig. 5B). The release of cardiac troponin-I in the plasma happens more quickly and reaches much higher levels.^{70,71} At the cardiomyocyte level, the exocytosis of unbound cardiac troponin I is associated with damaged mitochondria and sarcomere protein removal and is enhanced in cardiomyocytes during stress.⁷² Taken together these data suggest that reduced function and metabolism in HTS-preserved constructs might be linked to cells rearranging their sarcomere structures and loss of cell-cell contacts. The latter could be partially reversed by allowing cTE constructs to recover in culture. Whereas in fast frozen constructs, the significant decrease in cardiac troponin T positive cells might indicate irreversible damage.

The development of a polymeric scaffold with arranged fibers was associated with improved cardiac cell arrangement and properties comparable to the native myocardium.⁷³ Additionally, upon implantation, cTE constructs need to withstand the continuous contractions and distensions of the heart, which can result in altered mechanical properties induced by irreversible crystallinity changes that may occur during cryopreservation.^{20,53} Furthermore, rapid temperature shifts in polymer filaments were associated with shrinking and consequent cell detachment and ultimately apoptosis.²⁰ In this study, the rapid cooling/warming of the PCL filaments did not result in (irreversibly) altered surface topography, or statistically significant differences in crystallinity, glass and melting temperature, or mechanical properties (Fig. 7). Ultimately, ensuring suitable mechanical properties is important for successful implantation and long-term retention. The cTE model used in this study demonstrates how small changes in crystallinity (Fig. 7B) are overpowered by the geometry/

design of the scaffold according to the uni-axial direction (Fig. 7C). Our results show no significant adverse effects of preservation on the polymer scaffolds, providing evidence that scaffold-free constructs of similar thickness would not yield different results. While increasing construct thickness could enhance the therapeutic effect, preserving larger cTE constructs is expected to be increasingly difficult, as bigger tissues often suffer from uneven freezing and thawing. Instead, thin constructs, such as the ones used in this study, bypass these issues and still hold clinical significance as stacking of thin constructs is possible and already applied in an ongoing clinical trial.⁷⁴

Altogether, our results suggest that cryogenic preservation induces severe and irreversible damages to cTE constructs, which display signs of hiPSC-CM dysfunction and death. DMSO toxicity may play a role in the decay of the cTE construct. Additionally, it is conceivable that the low thermal strain of the construct in combination with uneven cooling, results in ECM contraction/expansion and (limited) ice nucleation. This would negatively affect cell-cell and cell-matrix interactions, contributing to increased hiPSC-CM death.^{46,75} As cell death is permanent, cryopreservation technology needs further improvements to achieve preserved post-storage cTE construct viability and functionality. Several approaches could be applied focusing on both the biology and methodology. First, toxic CPAs may be (partially) substituted by non-toxic alternatives, such as ice-binding proteins and other materials that inhibit recrystallization and/or promote nucleation near 0 °C. Second, the field of cryopreservation is moving towards the use of CPA cocktails with higher CPA concentrations (*e.g.*, DP6, VS55, and M22) and ultra-fast cooling and rewarming rates ($>10^7$ °C min⁻¹) to entirely avoid the formation of ice crystals. This method, referred to as vitrification, allows for the samples to acquire a glassy state without ice nucleation, circumventing ice-associated damage to cell-cell junctions and the extracellular matrix.^{50,75–78} Recently, rat kidney vitrification was successfully performed using perfused iron oxide nanoparticles to uniformly heat the sample, resulting in the successful transplantation of the preserved kidney.⁷⁹

In turn, HTS-preserved cTE constructs were correlated with high viability and fast metabolic recovery, with better preservation of hiPSC-CM sarcomere ultrastructure. However, in hypothermia time does not “stop”, but it merely slows down. Therefore, HTS provides a good platform for short-term preservation, extending the time between production and application, allowing construct transport.

Conclusion

Overall, the results of this study provide insights into the preservation-associated changes to cTE constructs and suggest that HTS is a promising solution for their short-term preservation. Further studies are required to optimize promising long-term preservation techniques, such as fast freezing and vitrification, as well as to evaluate their long-term effects on the cTE con-



structs. To conclude, our findings provide a basis for the development of efficient preservation techniques, ultimately improving the accessibility and potential benefits of cTE approaches for patients suffering from heart diseases.

Author contributions

Conceptualization (JJ, LvL, AvM), data curation (NC, JJ, MJA, VSP), formal analysis (NC, JJ, MJA, GCS, MV), investigation (NC, JJ, MJA, GCS, MV, ID), methodology (NC, JJ, MJA, TV, MS, MDC, AvM), writing – original draft (NC, JJ, MA, VSP, AvM), writing – review & editing (JJ, NC, MJA, ID, TV, PAD, MS, IKV, JM, LvL, JPGS, VSP, AvM), funding acquisition (AvM, JPGS, PAD, JM, TV, LvL, IKV, VSP), resources (TV, JM, MDC), software (NC, MA, GCS, VSP), project administration (AvM, JPGS), supervision (AvM, VSP, JPGS, MDC, JM, TV).

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

The authors would like to gratefully acknowledge Tomo Saric (Uniklinik Köln) for providing the human iPSC lines and M.-J. Goumans (LUMC) for providing the hfCFs. This work has been supported by the EU's H2020 Marie Skłodowska-Curie Actions COFUND programme under grant agreement #801540 (RESCUE), the EU H2020 research and innovation programme under grant agreement #874827 (BRAVE), as well as the Gravitation Program "Materials Driven Regeneration", funded by the Netherlands Organization for Scientific Research (024.003.013). The work was funded by the alliance between Eindhoven University of Technology, Utrecht University and the University Medical Center Utrecht (to LvL). This work was supported by European Research Council (ERC) under the EVICARE grant (number 725229) to JPGS. V. S.-P. was supported by a Netherlands Heart Institute postdoctoral fellowship.

References

- 1 M. J. Leening, S. Siregar, I. Vaartjes, M. L. Bots, M. I. Versteegh, R. J. van Geuns, *et al.*, Heart disease in the Netherlands: a quantitative update, *Neth. Heart J.*, 2014, **22**(1), 3–10.
- 2 M. Fuchs, D. Schibilsky, W. Zeh, M. Berchtold-Herz, F. Beyersdorf and M. Siepe, Does the heart transplant have a future?, *Eur. J. Cardiothorac. Surg.*, 2019, **55**(Suppl 1), i38–i48.
- 3 T. Resch, B. Cardini, R. Oberhuber, A. Weissenbacher, J. Dumfarth, C. Krapf, *et al.*, Transplanting Marginal Organs in the Era of Modern Machine Perfusion and Advanced Organ Monitoring, *Front. Immunol.*, 2020, **11**, 631.
- 4 G. S. Bleumink, A. M. Knetsch, M. C. Sturkenboom, S. M. Straus, A. Hofman, J. W. Deckers, *et al.*, Quantifying the heart failure epidemic: prevalence, incidence rate, lifetime risk and prognosis of heart failure The Rotterdam Study, *Eur. Heart J.*, 2004, **25**(18), 1614–1619.
- 5 E. E. van Riet, A. W. Hoes, K. P. Wagenaar, A. Limburg, M. A. Landman and F. H. Rutten, Epidemiology of heart failure: the prevalence of heart failure and ventricular dysfunction in older adults over time. A systematic review, *Eur. J. Heart Failure*, 2016, **18**(3), 242–252.
- 6 V. L. Roger, Epidemiology of Heart Failure, *Circ. Res.*, 2013, **113**(6), 646–659.
- 7 J. Wang, S. Liu, T. Heallen and J. F. Martin, The Hippo pathway in the heart: pivotal roles in development, disease, and regeneration, *Nat. Rev. Cardiol.*, 2018, **15**(11), 672–684.
- 8 A. Ferrini, M. M. Stevens, S. Sattler and N. Rosenthal, Toward Regeneration of the Heart: Bioengineering Strategies for Immunomodulation, *Front. Cardiovasc. Med.*, 2019, **6**, 26.
- 9 M. Rheault-Henry, I. White, D. Grover and R. Atoui, Stem cell therapy for heart failure: Medical breakthrough, or dead end?, *World J. Stem Cells*, 2021, **13**(4), 236–259.
- 10 F. van den Akker, D. A. Feyen, P. van den Hoogen, L. W. van Laake, E. C. van Eeuwijk, I. Hoefer, *et al.*, Intramyocardial stem cell injection: go(ne) with the flow, *Eur. Heart J.*, 2017, **38**(3), 184–186.
- 11 R. Gaetani, D. A. Feyen, V. Verhage, R. Slaats, E. Messina, K. L. Christman, *et al.*, Epicardial application of cardiac progenitor cells in a 3D-printed gelatin/hyaluronic acid patch preserves cardiac function after myocardial infarction, *Biomaterials*, 2015, **61**, 339–348.
- 12 Y. Shiba, T. Gomibuchi, T. Seto, Y. Wada, H. Ichimura, Y. Tanaka, *et al.*, Allogeneic transplantation of iPS cell-derived cardiomyocytes regenerates primate hearts, *Nature*, 2016, **538**(7625), 388–391.
- 13 R. J. Jabbour, T. J. Owen, P. Pandey, M. Reinsch, B. Wang, O. King, *et al.*, In vivo grafting of large engineered heart tissue patches for cardiac repair, *JCI Insight*, 2021, **6**(15), e144068.
- 14 L. Gao, Z. R. Gregorich, W. Zhu, S. Mattapally, Y. Oduk, X. Lou, *et al.*, Large Cardiac Muscle Patches Engineered From Human Induced-Pluripotent Stem Cell-Derived Cardiac Cells Improve Recovery From Myocardial Infarction in Swine, *Circulation*, 2018, **137**(16), 1712–1730.
- 15 Y. Yoshikawa, S. Miyagawa, K. Toda, A. Saito, Y. Sakata and Y. Sawa, Myocardial regenerative therapy using a scaffold-free skeletal-muscle-derived cell sheet in patients with dilated cardiomyopathy even under a left ventricular assist device: a safety and feasibility study, *Surg Today*, 2018, **48**(2), 200–210.
- 16 H. Narita, F. Shima, J. Yokoyama, S. Miyagawa, Y. Tsukamoto, Y. Takamura, *et al.*, Engraftment and morphological development of vascularized human iPS cell-



- derived 3D-cardiomyocyte tissue after xenotransplantation, *Sci. Rep.*, 2017, **7**(1), 13708.
- 17 M. Kawamura, S. Miyagawa, K. Miki, A. Saito, S. Fukushima, T. Higuchi, *et al.*, Feasibility, safety, and therapeutic efficacy of human induced pluripotent stem cell-derived cardiomyocyte sheets in a porcine ischemic cardiomyopathy model, *Circulation*, 2012, **126**(11 Suppl 1), S29–S37.
 - 18 Therapeutics H, University CS., Allogeneic iPSC-derived Cardiomyocyte Therapy in Patients With Worsening Ischemic Heart Failure. <https://ClinicalTrials.gov/show/NCT05566600>, 2022.
 - 19 Goettingen UMC, Herz-Kreislauf-Forschung DZf, Freiburg UMC, Safety and Efficacy of Induced Pluripotent Stem Cell-derived Engineered Human Myocardium as Biological Ventricular Assist Tissue in Terminal Heart Failure. <https://ClinicalTrials.gov/show/NCT04396899>, 2020.
 - 20 V. Sampaio-Pinto, J. Janssen, N. Chirico, M. Serra, P. M. Alves, P. A. Doevendans, *et al.*, A Roadmap to Cardiac Tissue-Engineered Construct Preservation: Insights from Cells, Tissues, and Organs, *Adv. Mater.*, 2021, **33**(27), e2008517.
 - 21 S. Freitas-Ribeiro, R. L. Reis and R. P. Pirraco, Long-term and short-term preservation strategies for tissue engineering and regenerative medicine products: state of the art and emerging trends, *PNAS Nexus*, 2022, **1**(4), pgac212.
 - 22 K. Takuji, I. Yoshito, I. Emiko, T. Maki, M. Tsubasa, T. Takura, *et al.*, Safety confirmation of induced pluripotent stem cell-derived cardiomyocyte patch transplantation for ischemic cardiomyopathy: first three case reports, *Front. Cardiovasc. Med.*, 2023, **10**, 3389.
 - 23 P. Niederberger, E. Farine, M. Raillard, M. Dornbierer, D. H. Freed, S. R. Large, *et al.*, Heart Transplantation With Donation After Circulatory Death, *Circ.: Heart Failure*, 2019, **12**(4), e005517.
 - 24 K. K. Dhital, A. Iyer, M. Connellan, H. C. Chew, L. Gao, A. Doyle, *et al.*, Adult heart transplantation with distant procurement and *ex vivo* preservation of donor hearts after circulatory death: a case series, *Lancet*, 2015, **385**(9987), 2585–2591.
 - 25 M. J. M. Silvis, S. E. Kaffka Genaamd Dengler, C. A. Odille, M. Mishra, N. P. van der Kaaij, P. A. Doevendans, *et al.*, Damage-Associated Molecular Patterns in Myocardial Infarction and Heart Transplantation: The Road to Translational Success, *Front. Immunol.*, 2020, **11**, 599511.
 - 26 S. Freitas-Ribeiro, A. F. Carvalho, M. Costa, M. T. Cerqueira, A. P. Marques, R. L. Reis, *et al.*, Strategies for the hypothermic preservation of cell sheets of human adipose stem cells, *PLoS One*, 2019, **14**(10), e0222597.
 - 27 B. Rubinsky, Principles of low temperature cell preservation, *Heart Failure Rev.*, 2003, **8**(3), 277–284.
 - 28 C. Correia, A. Koshkin, M. Carido, N. Espinha, T. Saric, P. A. Lima, *et al.*, Effective Hypothermic Storage of Human Pluripotent Stem Cell-Derived Cardiomyocytes Compatible With Global Distribution of Cells for Clinical Applications and Toxicology Testing, *Stem Cells Transl. Med.*, 2016, **5**(5), 658–669.
 - 29 N. Latchana, J. R. Peck, B. Whitson and S. M. Black, Preservation solutions for cardiac and pulmonary donor grafts: a review of the current literature, *J. Thorac. Dis.*, 2014, **6**(8), 1143–1149.
 - 30 P. Mazur, Freezing of living cells: mechanisms and implications, *Am. J. Physiol.*, 1984, **247**(3 Pt 1), C125–C142.
 - 31 E. B. Finger and J. C. Bischof, Cryopreservation by vitrification: a promising approach for transplant organ banking, *Curr. Opin. Organ Transplant.*, 2018, **23**(3), 353–360.
 - 32 A. Chiu-Lam, E. Staples, C. J. Pepine and C. Rinaldi, Perfusion, cryopreservation, and nanowarming of whole hearts using colloiddally stable magnetic cryopreservation agent solutions, *Sci. Adv.*, 2021, **7**(2), eabe3005.
 - 33 R. G. C. Maas, T. Beekink, N. Chirico, C. J. B. Snijders Blok, I. Dokter, V. Sampaio-Pinto, *et al.*, Generation, High-Throughput Screening, and Biobanking of Human-Induced Pluripotent Stem Cell-Derived Cardiac Spheroids, *J. Visualized Exp.*, 2023, (193), e64365.
 - 34 M. Castilho, A. van Mil, M. Maher, C. H. G. Metz, G. Hochleitner, J. Groll, *et al.*, Melt Electrowriting Allows Tailored Microstructural and Mechanical Design of Scaffolds to Advance Functional Human Myocardial Tissue Formation, *Adv. Funct. Mater.*, 2018, **28**(40), 1803151.
 - 35 M. J. Ainsworth, N. Chirico, M. de Ruijter, A. Hrynevich, I. Dokter, J. P. G. Sluijter, *et al.*, Convergence of melt electrowriting and extrusion-based bioprinting for vascular patterning of a myocardial construct, *Biofabrication*, 2023, **15**(3), 035025.
 - 36 S. Hamad, D. Derichsweiler, S. Papadopoulos, F. Nguemo, T. Šarić, A. Sachinidis, *et al.*, Generation of human induced pluripotent stem cell-derived cardiomyocytes in 2D monolayer and scalable 3D suspension bioreactor cultures with reduced batch-to-batch variations, *Theranostics*, 2019, **9**(24), 7222–7238.
 - 37 N. Chirico, E. L. Kessler, R. G. C. Maas, J. Fang, J. Qin, I. Dokter, *et al.*, Small molecule-mediated rapid maturation of human induced pluripotent stem cell-derived cardiomyocytes, *Stem Cell Res. Ther.*, 2022, **13**(1), 531.
 - 38 R. G. C. Maas, S. Lee, M. Harakalova, C. J. B. Snijders Blok, W. R. Goodyer, J. Hjortnaes, *et al.*, Massive expansion and cryopreservation of functional human induced pluripotent stem cell-derived cardiomyocytes, *STAR Protoc.*, 2021, **2**(1), 100334.
 - 39 J. Buikema, S. Lee, W. Goodyer, R. Maas, O. Chirikian, G. Li, *et al.*, Wnt Activation and Reduced Cell-Cell Contact Synergistically Induce Massive Expansion of Functional Human iPSC-Derived Cardiomyocytes, *Cell Stem Cell*, 2020, **27**(1), 50–63.
 - 40 T. C. L. Bracco Gartner, S. Crnko, L. Leiteris, I. Van Adrichem, L. W. Van Laake, C. V. C. Bouten, *et al.*, Pirfenidone Has Anti-fibrotic Effects in a Tissue-Engineered Model of Human Cardiac Fibrosis, *Front. Cardiovasc. Med.*, 2022, **9**, 854314.
 - 41 M. Castilho, D. Feyen, M. Flandes-Iparraguirre, G. Hochleitner, J. Groll, P. A. F. Doevendans, *et al.*, Melt Electrospinning Writing of Poly-Hydroxymethylglycolide-



- co-epsilon-Caprolactone-Based Scaffolds for Cardiac Tissue Engineering, *Adv. Healthcare Mater.*, 2017, **6**(18), 1700311.
- 42 K. S. Lim, B. J. Klotz, G. C. J. Lindberg, F. P. W. Melchels, G. J. Hooper, J. Malda, *et al.*, Visible Light Cross-Linking of Gelatin Hydrogels Offers an Enhanced Cell Microenvironment with Improved Light Penetration Depth, *Macromol. Biosci.*, 2019, **19**(6), e1900098.
 - 43 Rokepie Cell Protection by Hibernation [available from: https://www.rokepie.com/wp-content/uploads/2016/12/Application_Note_ROKEPIE.pdf].
 - 44 Rokepie Hibernation Technology for Cells [available from: <https://www.rokepie.com/wp-content/uploads/2016/12/Instructions-ROKEPIE-FD01.pdf>].
 - 45 A. Vasquez-Rivera, K. K. Sommer, H. Oldenhof, A. Z. Higgins, K. G. M. Brockbank, A. Hilfiker, *et al.*, Simultaneous monitoring of different vitrification solution components permeating into tissues, *Analyst*, 2018, **143**(2), 420–428.
 - 46 L. J. Fry, S. Querol, S. G. Gomez, S. McArdle, R. Rees and J. A. Madrigal, Assessing the toxic effects of DMSO on cord blood to determine exposure time limits and the optimum concentration for cryopreservation, *Vox Sang.*, 2015, **109**(2), 181–190.
 - 47 A. Blocki, F. Loper, N. Chirico, A. T. Neffe, F. Jung, C. Stamm, *et al.*, Engineering of cell-laden gelatin-based microgels for cell delivery and immobilization in regenerative therapies, *Clin. Hemorheol. Microcirc.*, 2017, **67**(3–4), 251–259.
 - 48 Y. Y. Kim, S. Y. Ku, H. C. Liu, H. J. Cho, S. K. Oh, S. Y. Moon, *et al.*, Cryopreservation of human embryonic stem cells derived-cardiomyocytes induced by BMP2 in serum-free condition, *Reprod. Sci.*, 2011, **18**(3), 252–260.
 - 49 H. S. Hwang, D. O. Kryshal, T. K. Feaster, V. Sanchez-Freire, J. Zhang, T. J. Kamp, *et al.*, Comparable calcium handling of human iPSC-derived cardiomyocytes generated by multiple laboratories, *J. Mol. Cell. Cardiol.*, 2015, **85**, 79–88.
 - 50 M. K. Preininger, M. Singh and C. H. Xu, Cryopreservation of Human Pluripotent Stem Cell-Derived Cardiomyocytes: Strategies, Challenges, and Future Directions, *Adv. Exp. Med. Biol.*, 2016, **951**, 123–135.
 - 51 P. Rana, B. Anson, S. Engle and Y. Will, Characterization of Human-Induced Pluripotent Stem Cell-Derived Cardiomyocytes: Bioenergetics and Utilization in Safety Screening, *Toxicol. Sci.*, 2012, **130**(1), 117–131.
 - 52 J. Z. Zhang, N. Belbachir, T. Zhang, Y. Liu, R. Shrestha and J. C. Wu, Effects of Cryopreservation on Human Induced Pluripotent Stem Cell-Derived Cardiomyocytes for Assessing Drug Safety Response Profiles, *Stem Cell Rep.*, 2021, **16**(1), 168–181.
 - 53 O. Batnyam, S.-I. Suye and S. Fujita, Direct cryopreservation of adherent cells on an elastic nanofiber sheet featuring a low glass-transition temperature, *RSC Adv.*, 2017, **7**(81), 51264–51271.
 - 54 S. Sullivan, P. J. Fairchild, S. G. E. Marsh, C. R. Muller, M. L. Turner, J. Song, *et al.*, Haplobanking induced pluripotent stem cells for clinical use, *Stem Cell Res.*, 2020, **49**, 102035.
 - 55 G. B. Basara, G. Ozcebe, B. W. Ellis, G. Ronan and P. Zorlutuna, Myocardial infarction from a tissue engineering and regenerative medicine point of view: A comprehensive review on models and treatments, *Biophys. Rev.*, 2022, **3**(3), 031305.
 - 56 R. Augustine, P. Dan, A. Hasan, I. M. Khalaf, P. Prasad, K. Ghosal, *et al.*, Stem cell-based approaches in cardiac tissue engineering: controlling the microenvironment for autologous cells, *Biomed. Pharmacother.*, 2021, **138**, 111425.
 - 57 M. Vis, K. Ito and S. Hofmann, Impact of Culture Medium on Cellular Interactions in in vitro Co-culture Systems, *Front. Bioeng. Biotechnol.*, 2020, **8**, 911.
 - 58 A. Ostrowska, K. Gu, D. C. Bode and R. G. Van Buskirk, Hypothermic storage of isolated human hepatocytes: a comparison between University of Wisconsin solution and a hypothermosol platform, *Arch. Toxicol.*, 2009, **83**(5), 493–502.
 - 59 G. Hajmoussa, P. Vogelaar, L. A. Brouwer, A. C. van der Graaf, R. H. Henning and G. Krenning, The 6-chromanol derivate SUL-109 enables prolonged hypothermic storage of adipose tissue-derived stem cells, *Biomaterials*, 2017, **119**, 43–52.
 - 60 P. Joshi, L. E. Ehrlich, Z. Gao, J. C. Bischof and Y. Rabin, Thermal Analyses of Nanowarming-Assisted Recovery of the Heart From Cryopreservation by Vitrification, *J. Heat Transfer*, 2022, **144**(3), 031202.
 - 61 Z. Gao, B. Namsrai, Z. Han, P. Joshi, J. S. Rao, V. Ravikumar, *et al.*, Vitrification and Rewarming of Magnetic Nanoparticle-Loaded Rat Hearts, *Adv. Mater. Technol.*, 2022, **7**(3), 2100873.
 - 62 E. B. Finger and J. C. Bischof, Cryopreservation by vitrification: a promising approach for transplant organ banking, *Curr. Opin. Organ Transplant.*, 2018, **23**(3), 353–360.
 - 63 A. Braik, M. Lahouel, R. Merabet, M. R. Djebbar and D. Morin, Myocardial protection by propolis during prolonged hypothermic preservation, *Cryobiology*, 2019, **88**, 29–37.
 - 64 E. S. Teh, F. Zal, V. Polard, P. Menasche and D. J. Chambers, HEMO(2)life as a protective additive to Celsior solution for static storage of donor hearts prior to transplantation, *Artif. Cells, Nanomed., Biotechnol.*, 2017, **45**(4), 717–722.
 - 65 C. Lazzeri, S. Valente, M. Chiostri and G. F. Gensini, Clinical significance of lactate in acute cardiac patients, *World J. Cardiol.*, 2015, **7**(8), 483–489.
 - 66 R. Zymlinski, J. Biegus, M. Sokolski, S. Nawrocka, P. Siwolowski, J. Szachniewicz, *et al.*, Elevated serum lactate is a marker of organ damage related to impaired hemodynamics and indicates poor prognosis in acute heart failure, *Eur. Heart J.*, 2013, **34**, 294.
 - 67 S. Vuckovic, R. Dinani, E. E. Nollet, D. W. D. Kuster, J. W. Buikema, R. H. Houtkooper, *et al.*, Characterization of cardiac metabolism in iPSC-derived cardiomyocytes:



- lessons from maturation and disease modeling, *Stem Cell Res. Ther.*, 2022, **13**(1), 332.
- 68 C. A. Henderson, C. G. Gomez, S. M. Novak, L. Mi-Mi and C. C. Gregorio, Overview of the Muscle Cytoskeleton, *Compr. Physiol.*, 2017, **7**(3), 891–944.
 - 69 D. Kraus, B. von Jeinsen, S. Tzikas, L. Palapies, T. Zeller, C. Bickel, *et al.*, Cardiac Troponins for the Diagnosis of Acute Myocardial Infarction in Chronic Kidney Disease, *J. Am. Heart Assoc.*, 2018, **7**(19), e008032.
 - 70 A. Arnadottir, S. Pedersen, R. Bo Hasselbalch, J. P. Goetze, L. J. Friis-Hansen, A. M. Bloch-Munster, *et al.*, Temporal Release of High-Sensitivity Cardiac Troponin T and I and Copeptin After Brief Induced Coronary Artery Balloon Occlusion in Humans, *Circulation*, 2021, **143**(11), 1095–1104.
 - 71 O. Hammarsten, M. Wernbom, N. L. Mills and C. Mueller, How is cardiac troponin released from cardiomyocytes?, *Eur. Heart J. Acute Cardiovasc. Care*, 2022, **11**(9), 718–720.
 - 72 J. A. Nicolas-Avila, A. V. Lechuga-Vieco, L. Esteban-Martinez, M. Sanchez-Diaz, E. Diaz-Garcia, D. J. Santiago, *et al.*, A Network of Macrophages Supports Mitochondrial Homeostasis in the Heart, *Cell*, 2020, **183**(1), 94–109.
 - 73 M. Kristen, M. J. Ainsworth, N. Chirico, C. F. T. van der Ven, P. A. Doevendans, J. P. G. Sluijter, *et al.*, Fiber Scaffold Patterning for Mending Hearts: 3D Organization Bringing the Next Step, *Adv. Healthcare Mater.*, 2020, **9**(1), e1900775.
 - 74 Heart patch in clinical trial: Dose finding completed (BioVAT-DZHK20 study) 2024 [available from: <https://dzhk.de/en/news/latest-news/article/heart-patch-in-clinical-trial-dose-finding-completed-biovat-dzhk20-study-1/>].
 - 75 R. D. Rodriguez, B. Van Hoeck, S. De Gelas, F. Blancke, R. Ngakam, K. Bogaerts, *et al.*, Determination of residual dimethylsulfoxide in cryopreserved cardiovascular allografts, *Cell Tissue Banking*, 2017, **18**(2), 263–270.
 - 76 H. Ohkawara, S. Miyagawa, S. Fukushima, S. Yajima, A. Saito, H. Nagashima, *et al.*, Development of a vitrification method for preserving human myoblast cell sheets for myocardial regeneration therapy, *BMC Biotechnol.*, 2018, **18**, 56.
 - 77 J. B. Hopkins, R. Badeau, M. Warkentin and R. E. Thorne, Effect of common cryoprotectants on critical warming rates and ice formation in aqueous solutions, *Cryobiology*, 2012, **65**(3), 169–178.
 - 78 S. Phatak, H. Natesan, J. Choi, K. G. M. Brockbank and J. C. Bischof, Measurement of Specific Heat and Crystallization in VS55, DP6, and M22 Cryoprotectant Systems With and Without Sucrose, *Biopreserv. Biobanking*, 2018, **16**(4), 270–277.
 - 79 Z. Han, J. S. Rao, L. Gangwar, B.-E. Namsrai, J. L. Pasek-Allen, M. L. Etheridge, *et al.*, Vitrification and nanowarming enable long-term organ cryopreservation and life-sustaining kidney transplantation in a rat model, *Nat. Commun.*, 2023, **14**(1), 3407.

